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(54) Title: LIGAND

(57) Abstract: The invention provides a dual-specific ligand comprising a first immunoglobulin variable domain having a first binding specificity and a complementary or non-complementary immunoglobulin variable domain having a second binding specificity.

Ligand

The present invention relates to dual specific ligands. In particular, the invention provides a method for the preparation of dual-specific ligands comprising a first immunoglobulin single variable domain binding to a first antigen or epitope, and a second immunoglobulin single variable domain binding to a second antigen or epitope. More particularly, the invention relates to dual-specific ligands wherein binding to at least one of the first and second antigens or epitopes acts to increase the half-life of the ligand *in vivo*. Open and closed conformation ligands comprising more than one binding specificity are described.

Introduction

The antigen binding domain of an antibody comprises two separate regions: a heavy chain variable domain (V_H) and a light chain variable domain (V_L : which can be either V_K or V_λ). The antigen binding site itself is formed by six polypeptide loops: three from V_H domain (H1, H2 and H3) and three from V_L domain (L1, L2 and L3). A diverse primary repertoire of V genes that encode the V_H and V_L domains is produced by the combinatorial rearrangement of gene segments. The V_H gene is produced by the recombination of three gene segments, V_H , D and J_H . In humans, there are approximately 51 functional V_H segments (Cook and Tomlinson (1995) *Immunol Today*, 16: 237), 25 functional D segments (Corbett *et al.* (1997) *J. Mol. Biol.*, 268: 69) and 6 functional J_H segments (Ravetch *et al.* (1981) *Cell*, 27: 583), depending on the haplotype. The V_H segment encodes the region of the polypeptide chain which forms the first and second antigen binding loops of the V_H domain (H1 and H2), whilst the V_H , D and J_H segments combine to form the third antigen binding loop of the V_H domain (H3). The V_L gene is produced by the recombination of only two gene segments, V_L and J_L . In humans, there are approximately 40 functional V_K segments (Schäble and Zachau (1993) *Biol. Chem. Hoppe-Seyler*, 374: 1001), 31 functional V_λ segments (Williams *et al.* (1996) *J. Mol. Biol.*, 264: 220; Kawasaki *et al.* (1997) *Genome Res.*, 7: 250), 5 functional J_K segments (Hieter *et al.* (1982) *J. Biol. Chem.*, 257: 1516) and 4 functional J_λ segments (Vasicek

and Leder (1990) *J. Exp. Med.*, 172: 609), depending on the haplotype. The V_L segment encodes the region of the polypeptide chain which forms the first and second antigen binding loops of the V_L domain (L1 and L2), whilst the V_L and J_L segments combine to form the third antigen binding loop of the V_L domain (L3). Antibodies selected from this 5 primary repertoire are believed to be sufficiently diverse to bind almost all antigens with at least moderate affinity. High affinity antibodies are produced by "affinity maturation" of the rearranged genes, in which point mutations are generated and selected by the immune system on the basis of improved binding.

10 Analysis of the structures and sequences of antibodies has shown that five of the six antigen binding loops (H1, H2, L1, L2, L3) possess a limited number of main-chain conformations or canonical structures (Chothia and Lesk (1987) *J. Mol. Biol.*, 196: 901; Chothia *et al.* (1989) *Nature*, 342: 877). The main-chain conformations are determined by (i) the length of the antigen binding loop, and (ii) particular residues, or types of residue, 15 at certain key position in the antigen binding loop and the antibody framework. Analysis of the loop lengths and key residues has enabled us to predict the main-chain conformations of H1, H2, L1, L2 and L3 encoded by the majority of human antibody sequences (Chothia *et al.* (1992) *J. Mol. Biol.*, 227: 799; Tomlinson *et al.* (1995) *EMBO J.*, 14: 4628; Williams *et al.* (1996) *J. Mol. Biol.*, 264: 220). Although the H3 region is 20 much more diverse in terms of sequence, length and structure (due to the use of D segments), it also forms a limited number of main-chain conformations for short loop lengths which depend on the length and the presence of particular residues, or types of residue, at key positions in the loop and the antibody framework (Martin *et al.* (1996) *J. Mol. Biol.*, 263: 800; Shirai *et al.* (1996) *FEBS Letters*, 399: 1).

25 Bispecific antibodies comprising complementary pairs of V_H and V_L regions are known in the art. These bispecific antibodies must comprise two pairs of V_H and V_L s, each V_H/V_L pair binding to a single antigen or epitope. Methods described involve hybrid hybridomas (Milstein & Cuello AC, *Nature* 305:537-40), minibodies (Hu *et al.*, (1996) *Cancer Res* 30:3055-3061;), diabodies (Holliger *et al.*, (1993) *Proc. Natl. Acad. Sci. USA* 90, 6444-6448; WO 94/13804), chelating recombinant antibodies (CRAbs; (Neri *et al.*, (1995) *J. Mol. Biol.* 246, 367-373), biscFv (e.g. Atwell *et al.*, (1996) *Mol. Immunol.* 33, 1301-1312), "knobs in holes" stabilised antibodies (Carter *et al.*, (1997) *Protein Sci.* 6, 781-

788). In each case each antibody species comprises two antigen-binding sites, each fashioned by a complementary pair of V_H and V_L domains. Each antibody is thereby able to bind to two different antigens or epitopes at the same time, with the binding to EACH antigen or epitope mediated by a V_H and its complementary V_L domain. Each of these 5 techniques presents its particular disadvantages; for instance in the case of hybrid hybridomas, inactive V_H/V_L pairs can greatly reduce the fraction of bispecific IgG. Furthermore, most bispecific approaches rely on the association of the different V_H/V_L pairs or the association of V_H and V_L chains to recreate the two different V_H/V_L binding sites. It is therefore impossible to control the ratio of binding sites to each antigen or 10 epitope in the assembled molecule and thus many of the assembled molecules will bind to one antigen or epitope but not the other. In some cases it has been possible to engineer the heavy or light chains at the sub-unit interfaces (Carter *et al.*, 1997) in order to improve the number of molecules which have binding sites to both antigens or epitopes but this never results in all molecules having binding to both antigens or epitopes.

15 There is some evidence that two different antibody binding specificities might be incorporated into the same binding site, but these generally represent two or more specificities that correspond to structurally related antigens or epitopes or to antibodies that are broadly cross-reactive.. For example, cross-reactive antibodies have been 20 described, usually where the two antigens are related in sequence and structure, such as hen egg white lysozyme and turkey lysozyme (McCafferty *et al.*, WO 92/01047) or to free hapten and to hapten conjugated to carrier (Griffiths AD *et al.* *EMBO J* 1994 13:14 3245-60). In a further example, WO 02/02773 (Abbott Laboratories) describes antibody 25 molecules with "dual specificity". The antibody molecules referred to are antibodies raised or selected against multiple antigens, such that their specificity spans more than a single antigen. Each complementary V_H/V_L pair in the antibodies of WO 02/02773 specifies a single binding specificity for two or more structurally related antigens; the V_H and V_L domains in such complementary pairs do not each possess a separate specificity. The antibodies thus have a broad single specificity which encompasses two antigens, 30 which are structurally related. Furthermore natural autoantibodies have been described that are polyreactive (Casali & Notkins, *Ann. Rev. Immunol.* 7, 515-531), reacting with at least two (usually more) different antigens or epitopes that are not structurally related. It

has also been shown that selections of random peptide repertoires using phage display technology on a monoclonal antibody will identify a range of peptide sequences that fit the antigen binding site. Some of the sequences are highly related, fitting a consensus sequence, whereas others are very different and have been termed mimotopes (Lane & 5 Stephen, Current Opinion in Immunology, 1993, 5, 268-271). It is therefore clear that a natural four-chain antibody, comprising associated and complementary V_H and V_L domains, has the potential to bind to many different antigens from a large universe of known antigens. It is less clear how to create a binding site to two given antigens in the same antibody, particularly those which are not necessarily structurally related.

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Protein engineering methods have been suggested that may have a bearing on this. For example it has also been proposed that a catalytic antibody could be created with a binding activity to a metal ion through one variable domain, and to a hapten (substrate) through contacts with the metal ion and a complementary variable domain (Barbas et al., 15 1993 Proc. Natl. Acad. Sci USA 90, 6385-6389). However in this case, the binding and catalysis of the substrate (first antigen) is proposed to require the binding of the metal ion (second antigen). Thus the binding to the V_H/V_L pairing relates to a single but multi-component antigen.

20

Methods have been described for the creation of bispecific antibodies from camel antibody heavy chain single domains in which binding contacts for one antigen are created in one variable domain, and for a second antigen in a second variable domain. However the variable domains were not complementary. Thus a first heavy chain variable domain is selected against a first antigen, and a second heavy chain variable domain 25 against a second antigen, and then both domains are linked together on the same chain to give a bispecific antibody fragment (Conrath et al., J. Biol. Chem. 270, 27589-27594). However the camel heavy chain single domains are unusual in that they are derived from natural camel antibodies which have no light chains, and indeed the heavy chain single domains are unable to associate with camel light chains to form complementary V_H and 30 V_L pairs.

Single heavy chain variable domains have also been described, derived from natural antibodies which are normally associated with light chains (from monoclonal antibodies

or from repertoires of domains; see EP-A-0368684). These heavy chain variable domains have been shown to interact specifically with one or more related antigens but have not been combined with other heavy or light chain variable domains to create a ligand with a specificity for two or more different antigens. Furthermore, these single 5 domains have been shown to have a very short *in vivo* half-life. Therefore such domains are of limited therapeutic value.

It has been suggested to make bispecific antibody fragments by linking heavy chain variable domains of different specificity together (as described above). The disadvantage 10 with this approach is that isolated antibody variable domains may have a hydrophobic interface that normally makes interactions with the light chain and is exposed to solvent and may be "sticky" allowing the single domain to bind to hydrophobic surfaces. Furthermore, in the absence of a partner light chain the combination of two or more 15 different heavy chain variable domains and their association, possibly via their hydrophobic interfaces, may prevent them from binding to one in not both of the ligands they are able to bind in isolation. Moreover, in this case the heavy chain variable domains would not be associated with complementary light chain variable domains and thus may be less stable and readily unfold (Worn & Pluckthun, 1998 Biochemistry 37, 13120-7).

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Summary of the invention

The inventors have described, in their copending international patent application WO 03/002609 as well as copending unpublished UK patent application 0230203.2, dual 25 specific immunoglobulin ligands which comprise immunoglobulin single variable domains which each have different specificities. The domains may act in competition with each other or independently to bind antigens or epitopes on target molecules.

In a first configuration, the present invention provides a further improvement in dual 30 specific ligands as developed by the present inventors, in which one specificity of the ligand is directed towards a protein or polypeptide present *in vivo* in an organism which can act to increase the half-life of the ligand by binding to it.

Accordingly, in a first aspect, there is provided a dual-specific ligand comprising a first immunoglobulin single variable domain having a binding specificity to a first antigen or epitope and a second complementary immunoglobulin single variable domain having a binding activity to a second antigen or epitope, wherein one or both of said antigens or epitopes acts to increase the half-life of the ligand *in vivo* and wherein said first and second domains lack mutually complementary domains which share the same specificity, provided that said dual specific ligand does not consist of an anti-HSA V_H domain and an anti- β galactosidase V_K domain. Preferably, that neither of the first or second variable domains binds to human serum albumin (HSA).

10

Antigens or epitopes which increase the half-life of a ligand as described herein are advantageously present on proteins or polypeptides found in an organism *in vivo*. Examples include extracellular matrix proteins, blood proteins, and proteins present in various tissues in the organism. The proteins act to reduce the rate of ligand clearance from the blood, for example by acting as bulking agents, or by anchoring the ligand to a desired site of action. Examples of antigens/epitopes which increase half-life *in vivo* are given in Annex 1 below.

Increased half-life is useful in *in vivo* applications of immunoglobulins, especially 20 antibodies and most especially antibody fragments of small size. Such fragments (Fvs, disulphide bonded Fvs, Fabs, scFvs, dAbs) suffer from rapid clearance from the body; thus, whilst they are able to reach most parts of the body rapidly, and are quick to produce and easier to handle, their *in vivo* applications have been limited by their only brief persistence *in vivo*. The invention solves this problem by providing increased half-life of 25 the ligands *in vivo* and consequently longer persistence times in the body of the functional activity of the ligand.

Methods for pharmacokinetic analysis and determination of ligand half-life will be 30 familiar to those skilled in the art. Details may be found in *Kenneth, A et al: Chemical Stability of Pharmaceuticals: A Handbook for Pharmacists* and in *Peters et al, Pharmacokinetic analysis: A Practical Approach* (1996). Reference is also made to "Pharmacokinetics", M Gibaldi & D Perron, published by Marcel Dekker, 2nd Rev. ex

edition (1982), which describes pharmacokinetic parameters such as t alpha and t beta half lives and area under the curve (AUC).

Half lives ($t_{1/2}$ alpha and $t_{1/2}$ beta) and AUC can be determined from a curve of serum concentration of ligand against time. The WinNonlin analysis package (available from Pharsight Corp., Mountain View, CA94040, USA) can be used, for example, to model the curve. In a first phase (the alpha phase) the ligand is undergoing mainly distribution in the patient, with some elimination. A second phase (beta phase) is the terminal phase when the ligand has been distributed and the serum concentration is decreasing as the ligand is cleared from the patient. The t alpha half life is the half life of the first phase and the t beta half life is the half life of the second phase. Thus, advantageously, the present invention provides a ligand or a composition comprising a ligand according to the invention having a $t_{1/2}$ half-life in the range of 15 minutes or more. In one embodiment, the lower end of the range is 30 minutes, 45 minutes, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 7 hours, 10 hours, 11 hours or 12 hours. In addition, or alternatively, a ligand or composition according to the invention will have a $t_{1/2}$ half life in the range of up to and including 12 hours. In one embodiment, the upper end of the range is 11, 10, 9, 8, 7, 6 or 5 hours. An example of a suitable range is 1 to 6 hours, 2 to 5 hours or 3 to 4 hours.

Advantageously, the present invention provides a ligand or a composition comprising a ligand according to the invention having a $t_{1/2}$ half-life in the range of 2.5 hours or more. In one embodiment, the lower end of the range is 3 hours, 4 hours, 5 hours, 6 hours, 7 hours, 10 hours, 11 hours, or 12 hours. In addition, or alternatively, a ligand or composition according to the invention has a $t_{1/2}$ half-life in the range of up to and including 21 days. In one embodiment, the upper end of the range is 12 hours, 24 hours, 2 days, 3 days, 5 days, 10 days, 15 days or 20 days. Advantageously a ligand or composition according to the invention will have a $t_{1/2}$ half life in the range 12 to 60 hours. In a further embodiment, it will be in the range 12 to 48 hours. In a further embodiment still, it will be in the range 12 to 26 hours.

In addition, or alternatively to the above criteria, the present invention provides a ligand or a composition comprising a ligand according to the invention having an AUC

value (area under the curve) in the range of 1 mg.min/ml or more. In one embodiment, the lower end of the range is 5, 10, 15, 20, 30, 100, 200 or 300mg.min/ml. In addition, or alternatively, a ligand or composition according to the invention has an AUC in the range of up to 600 mg.min/ml. In one embodiment, the upper end of the range is 500, 400, 300, 5 200, 150, 100, 75 or 50 mg.min/ml. Advantageously a ligand according to the invention will have a AUC in the range selected from the group consisting of the following: 15 to 150mg.min/ml, 15 to 100 mg.min/ml, 15 to 75 mg.min/ml, and 15 to 50mg.min/ml.

In a first embodiment, the dual specific ligand comprises two complementary variable domains, i.e. two variable domains that, in their natural environment, are capable of operating together as a cognate pair or group even if in the context of the present invention they bind separately to their cognate epitopes. For example, the complementary variable domains may be immunoglobulin heavy chain and light chain variable domains (V_H and V_L). V_H and V_L domains are advantageously provided by scFv or Fab antibody 10 fragments. Variable domains may be linked together to form multivalent ligands by, for example: provision of a hinge region at the C-terminus of each V domain and disulphide bonding between cysteines in the hinge regions; or provision of dAbs each with a cysteine at the C-terminus of the domain, the cysteines being disulphide bonded together; or 15 production of V-CH & V-CL to produce a Fab format; or use of peptide linkers (for example Gly₄Ser linkers discussed hereinbelow) to produce dimers, trimers and further 20 multimers.

The inventors have found that the use of complementary variable domains allows the two domain surfaces to pack together and be sequestered from the solvent. Furthermore the 25 complementary domains are able to stabilise each other. In addition, it allows the creation of dual-specific IgG antibodies without the disadvantages of hybrid hybridomas as used in the prior art, or the need to engineer heavy or light chains at the sub-unit interfaces. The dual-specific ligands of the first aspect of the present invention have at least one V_H/V_L pair. A bispecific IgG according to this invention will therefore comprise two 30 such pairs, one pair on each arm of the Y-shaped molecule. Unlike conventional bispecific antibodies or diabodies, therefore, where the ratio of chains used is determinative in the success of the preparation thereof and leads to practical difficulties, the dual specific ligands of the invention are free from issues of chain balance. Chain

imbalance in conventional bi-specific antibodies results from the association of two different V_L chains with two different V_H chains, where V_L chain 1 together with V_H chain 1 is able to bind to antigen or epitope 1 and V_L chain 2 together with V_H chain 2 is able to bind to antigen or epitope 2 and the two correct pairings are in some way linked to 5 one another. Thus, only when V_L chain 1 is paired with V_H chain 1 and V_L chain 2 is paired with V_H chain 2 in a single molecule is bi-specificity created. Such bi-specific molecules can be created in two different ways. Firstly, they can be created by association 10 of two existing V_H/V_L pairings that each bind to a different antigen or epitope (for example, in a bi-specific IgG). In this case the V_H/V_L pairings must come all together in a 1:1 ratio in order to create a population of molecules all of which are bi-specific. This never occurs (even when complementary CH domain is enhanced by "knobs into holes" engineering) leading to a mixture of bi-specific molecules and molecules that are only able to bind to one antigen or epitope but not the other. The second way of creating a bi-specific antibody is by the simultaneous association of two different V_H chain with two 15 different V_L chains (for example in a bi-specific diabody). In this case, although there tends to be a preference for V_L chain 1 to pair with V_H chain 1 and V_L chain 2 to pair with V_H chain 2 (which can be enhanced by "knobs into holes" engineering of the V_L and V_H domains), this paring is never achieved in all molecules, leading to a mixed formulation whereby incorrect pairings occur that are unable to bind to either antigen or epitope.

20 Bi-specific antibodies constructed according to the dual-specific ligand approach according to the first aspect of the present invention overcome all of these problems because the binding to antigen or epitope 1 resides within the V_H or V_L domain and the binding to antigen or epitope 2 resides with the complementary V_L or V_H domain, 25 respectively. Since V_H and V_L domains pair on a 1:1 basis all V_H/V_L pairings will be bi-specific and thus all formats constructed using these V_H/V_L pairings (Fv, scFvs, Fabs, minibodies, IgGs etc) will have 100% bi-specific activity.

30 In the context of the present invention, first and second "epitopes" are understood to be epitopes which are not the same and are not bound by a single monospecific ligand. In the first configuration of the invention, they are advantageously on different antigens, one of which acts to increase the half-life of the ligand *in vivo*. Likewise, the first and second antigens are advantageously not the same.

The dual specific ligands of the invention do not include ligands as described in WO 02/02773. Thus, the ligands of the present invention do not comprise complementary V_H/V_L pairs which bind any one or more antigens or epitopes co-operatively. Instead, the 5 ligands according to the first aspect of the invention comprise a V_H/V_L complementary pair, wherein the V domains have different specificities.

Moreover, the ligands according to the first aspect of the invention comprise V_H/V_L complementary pairs having different specificities for non-structurally related epitopes or 10 antigens. Structurally related epitopes or antigens are epitopes or antigens which possess sufficient structural similarity to be bound by a conventional V_H/V_L complementary pair which acts in a co-operative manner to bind an antigen or epitope; in the case of structurally related epitopes, the epitopes are sufficiently similar in structure that they “fit” into the same binding pocket formed at the antigen binding site of the V_H/V_L dimer.

15 In a second aspect, the present invention provides a ligand comprising a first immunoglobulin variable domain having a first antigen or epitope binding specificity and a second immunoglobulin variable domain having a second antigen or epitope binding specificity wherein one or both of said first and second variable domains bind to an 20 antigen which increases the half-life of the ligand *in vivo*, and the variable domains are not complementary to one another.

In one embodiment, binding to one variable domain modulates the binding of the ligand to the second variable domain.

25 In this embodiment, the variable domains may be, for example, pairs of V_H domains or pairs of V_L domains. Binding of antigen at the first site may modulate, such as enhance or inhibit, binding of an antigen at the second site. For example, binding at the first site at least partially inhibits binding of an antigen at a second site. In such an embodiment, the 30 ligand may for example be maintained in the body of a subject organism *in vivo* through binding to a protein which increases the half-life of the ligand until such a time as it becomes bound to the second target antigen and dissociates from the half-life increasing protein.

Modulation of binding in the above context is achieved as a consequence of the structural proximity of the antigen binding sites relative to one another. Such structural proximity can be achieved by the nature of the structural components linking the two or more 5 antigen binding sites, eg by the provision of a ligand with a relatively rigid structure that holds the antigen binding sites in close proximity. Advantageously, the two or more antigen binding sites are in physically close proximity to one another such that one site modulates the binding of antigen at another site by a process which involves steric hindrance and/or conformational changes within the immunoglobulin molecule.

10

The first and the second antigen binding domains may be associated either covalently or non-covalently. In the case that the domains are covalently associated, then the association may be mediated for example by disulphide bonds or by a polypeptide linker such as $(\text{Gly}_4\text{Ser})_n$, where $n =$ from 1 to 8, eg, 2, 3, 4, 5 or 7.

15

Ligands according to the invention may be combined into non-immunoglobulin multi-ligand structures to form multivalent complexes, which bind target molecules with the same antigen, thereby providing superior avidity, while at least one variable domain binds an antigen to increase the half life of the multimer. For example natural bacterial 20 receptors such as SpA have been used as scaffolds for the grafting of CDRs to generate ligands which bind specifically to one or more epitopes. Details of this procedure are described in US 5,831,012. Other suitable scaffolds include those based on fibronectin and affibodies. Details of suitable procedures are described in WO 98/58965. Other suitable scaffolds include lipocalin and CTLA4, as described in van den Beuken *et al.*, J. 25 Mol. Biol. (2001) 310, 591-601, and scaffolds such as those described in WO0069907 (Medical Research Council), which are based for example on the ring structure of bacterial GroEL or other chaperone polypeptides.

30 Protein scaffolds may be combined; for example, CDRs may be grafted on to a CTLA4 scaffold and used together with immunoglobulin V_H or V_L domains to form a ligand. Likewise, fibronectin, lipocalin and other scaffolds may be combined.

In the case that the variable domains are selected from V-gene repertoires selected for instance using phage display technology as herein described, then these variable domains can comprise a universal framework region, such that is they may be recognised by a specific generic ligand as herein defined. The use of universal frameworks, generic 5 ligands and the like is described in WO99/20749. In the present invention, reference to phage display includes the use of both phage and/or phagemids.

Where V-gene repertoires are used variation in polypeptide sequence is preferably located within the structural loops of the variable domains. The polypeptide sequences of either 10 variable domain may be altered by DNA shuffling or by mutation in order to enhance the interaction of each variable domain with its complementary pair.

In a preferred embodiment of the invention the 'dual-specific ligand' is a single chain Fv fragment. In an alternative embodiment of the invention, the 'dual-specific ligand' 15 consists of a Fab region of an antibody. The term "Fab region" includes a Fab-like region where two VH or two VL domains are used.

The variable regions may be derived from antibodies directed against target antigens or epitopes. Alternatively they may be derived from a repertoire of single antibody domains 20 such as those expressed on the surface of filamentous bacteriophage. Selection may be performed as described below.

In a third aspect, the invention provides a method for producing a ligand comprising a first immunoglobulin single variable domain having a first binding specificity and a 25 second single immunoglobulin single variable domain having a second (different) binding specificity, one or both of the binding specificities being specific for an antigen which increases the half-life of the ligand *in vivo*, the method comprising the steps of:

- (a) selecting a first variable domain by its ability to bind to a first epitope,
- (b) selecting a second variable region by its ability to bind to a second epitope,
- 30 (c) combining the variable domains; and
- (d) selecting the ligand by its ability to bind to said first epitope and to said second epitope.

The ligand can bind to the first and second epitopes either simultaneously or, where there is competition between the binding domains for epitope binding, the binding of one domain may preclude the binding of another domain to its cognate epitope. In one embodiment, therefore, step (d) above requires simultaneous binding to both first and 5 second (and possibly further) epitopes; in another embodiment, the binding to the first and second epitopes is not simultaneous.

The epitopes are preferably on separate antigens.

10 Ligands advantageously comprise V_H/V_L combinations, or V_H/V_H or V_L/V_L combinations of immunoglobulin variable domains, as described above. The ligands may moreover comprise camelid V_{HH} domains, provided that the V_{HH} domain which is specific for an antigen which increases the half-life of the ligand *in vivo* does not bind Hen egg white lysozyme (HEL), porcine pancreatic alpha-amylase or NmC-A; hcg, BSA-linked RR6 azo 15 dye or *S. mutans* HG982 cells, as described in Conrath *et al.*, (2001) JBC 276:7346-7350 and WO99/23221, neither of which describe the use of a specificity for an antigen which increases half-life to increase the half life of the ligand *in vivo*.

20 In one embodiment, said first variable domain is selected for binding to said first epitope in absence of a complementary variable domain. In a further embodiment, said first variable domain is selected for binding to said first epitope/antigen in the presence of a third variable domain in which said third variable domain is different from said second variable domain and is complementary to the first domain. Similarly, the second domain may be selected in the absence or presence of a complementary variable domain.

25

The antigens or epitopes targeted by the ligands of the invention, in addition to the half-life enhancing protein, may be any antigen or epitope but advantageously is an antigen or epitope that is targeted with therapeutic benefit. The invention provides ligands, including open conformation, closed conformation and isolated dAb monomer ligands, 30 specific for any such target, particularly those targets further identified herein. Such targets may be, or be part of, polypeptides, proteins or nucleic acids, which may be naturally occurring or synthetic. In this respect, the ligand of the invention may bind the epitope or antigen and act as an antagonist or agonist (eg, EPO receptor agonist). One

skilled in the art will appreciate that the choice is large and varied. They may be for instance human or animal proteins, cytokines, cytokine receptors, enzymes co-factors for enzymes or DNA binding proteins. Suitable cytokines and growth factors include but are not limited to: ApoE, Apo-SAA, BDNF, Cardiotrophin-1, EGF, EGF receptor, ENA-78, 5 Eotaxin, Eotaxin-2, Exodus-2, EpoR, FGF-acidic, FGF-basic, fibroblast growth factor-10, FLT3 ligand, Fractalkine (CX3C), GDNF, G-CSF, GM-CSF, GF- β 1, insulin, IFN- γ , IGF-I, IGF-II, IL-1 α , IL-1 β , IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8 (72 a.a.), IL-8 (77 a.a.), IL-9, IL-10, IL-11, IL-12, IL-13, IL-15, IL-16, IL-17, IL-18 (IGIF), Inhibin α , Inhibin β , IP-10, keratinocyte growth factor-2 (KGF-2), KGF, Leptin, LIF, Lymphotactin, 10 Mullerian inhibitory substance, monocyte colony inhibitory factor, monocyte attractant protein, M-CSF, MDC (67 a.a.), MDC (69 a.a.), MCP-1 (MCAF), MCP-2, MCP-3, MCP-4, MDC (67 a.a.), MDC (69 a.a.), MIG, MIP-1 α , MIP-1 β , MIP-3 α , MIP-3 β , MIP-4, myeloid progenitor inhibitor factor-1 (MPIF-1), NAP-2, Neurturin, Nerve growth factor, β -NGF, NT-3, NT-4, Oncostatin M, PDGF-AA, PDGF-AB, PDGF-BB, PF-4, RANTES, 15 SDF1 α , SDF1 β , SCF, SCGF, stem cell factor (SCF), TARC, TGF- α , TGF- β , TGF- β 2, TGF- β 3, tumour necrosis factor (TNF), TNF- α , TNF- β , TNF receptor I, TNF receptor II, TNIL-1, TPO, VEGF, VEGF receptor 1, VEGF receptor 2, VEGF receptor 3, GCP-2, GRO/MGSA, GRO- β , GRO- γ , HCC1, 1-309, HER 1, HER 2, HER 3 and HER 4. Cytokine receptors include receptors for the foregoing cytokines. It will be appreciated 20 that this list is by no means exhaustive.

In one embodiment of the invention, the variable domains are derived from a respective antibody directed against the antigen or epitope. In a preferred embodiment the variable domains are derived from a repertoire of single variable antibody domains.

25 In a further aspect, the present invention provides one or more nucleic acid molecules encoding at least a dual-specific ligand as herein defined. The dual specific ligand may be encoded on a single nucleic acid molecule; alternatively, each domain may be encoded by a separate nucleic acid molecule. Where the ligand is encoded by a single nucleic acid 30 molecule, the domains may be expressed as a fusion polypeptide, in the manner of a scFv molecule, or may be separately expressed and subsequently linked together, for example using chemical linking agents. Ligands expressed from separate nucleic acids will be linked together by appropriate means.

The nucleic acid may further encode a signal sequence for export of the polypeptides from a host cell upon expression and may be fused with a surface component of a filamentous bacteriophage particle (or other component of a selection display system) 5 upon expression.

In a further aspect the present invention provides a vector comprising nucleic acid encoding a dual specific ligand according to the present invention.

10 In a yet further aspect, the present invention provides a host cell transfected with a vector encoding a dual specific ligand according to the present invention.

Expression from such a vector may be configured to produce, for example on the surface of a bacteriophage particle, variable domains for selection. This allows selection of 15 displayed variable regions and thus selection of 'dual-specific ligands' using the method of the present invention.

The present invention further provides a kit comprising at least a dual-specific ligand according to the present invention.

20 Dual-Specific ligands according to the present invention preferably comprise combinations of heavy and light chain domains. For example, the dual specific ligand may comprise a V_H domain and a V_L domain, which may be linked together in the form of an scFv. In addition, the ligands may comprise one or more C_H or C_L domains. For 25 example, the ligands may comprise a C_H1 domain, C_H2 or C_H3 domain, and/or a C_L domain, $C_{\mu}1$, $C_{\mu}2$, $C_{\mu}3$ or $C_{\mu}4$ domains, or any combination thereof. A hinge region domain may also be included. Such combinations of domains may, for example, mimic natural antibodies, such as IgG or IgM, or fragments thereof, such as Fv, scFv, Fab or $F(ab')_2$ molecules. Other structures, such as a single arm of an IgG molecule comprising 30 V_H , V_L , C_H1 and C_L domains, are envisaged.

In a preferred embodiment of the invention, the variable regions are selected from single domain V gene repertoires. Generally the repertoire of single antibody domains is

displayed on the surface of filamentous bacteriophage. In a preferred embodiment each single antibody domain is selected by binding of a phage repertoire to antigen.

In a preferred embodiment of the invention each single variable domain may be selected
5 for binding to its target antigen or epitope in the absence of a complementary variable region. In an alternative embodiment, the single variable domains may be selected for binding to its target antigen or epitope in the presence of a complementary variable region. Thus the first single variable domain may be selected in the presence of a third complementary variable domain, and the second variable domain may be selected in the
10 presence of a fourth complementary variable domain. The complementary third or fourth variable domain may be the natural cognate variable domain having the same specificity as the single domain being tested, or a non-cognate complementary domain – such as a “dummy” variable domain.

15 Preferably, the dual specific ligand of the invention comprises only two variable domains although several such ligands may be incorporated together into the same protein, for example two such ligands can be incorporated into an IgG or a multimeric immunoglobulin, such as IgM. Alternatively, in another embodiment a plurality of dual specific ligands are combined to form a multimer. For example, two different dual
20 specific ligands are combined to create a tetra-specific molecule.

It will be appreciated by one skilled in the art that the light and heavy variable regions of a dual-specific ligand produced according to the method of the present invention may be on the same polypeptide chain, or alternatively, on different polypeptide chains. In the
25 case that the variable regions are on different polypeptide chains, then they may be linked via a linker, generally a flexible linker (such as a polypeptide chain), a chemical linking group, or any other method known in the art.

In a further aspect, the present invention provides a composition comprising a dual-specific ligand, obtainable by a method of the present invention, and a pharmaceutically acceptable carrier, diluent or excipient.
30

Moreover, the present invention provides a method for the treatment and/or prevention of disease using a 'dual-specific ligand' or a composition according to the present invention.

5 In a second configuration, the present invention provides multispecific ligands which comprise at least two non-complementary variable domains. For example, the ligands may comprise a pair of V_H domains or a pair of V_L domains. Advantageously, the domains are of non-camelid origin; preferably they are human domains or comprise 10 human framework regions (FWs) and one or more heterologous CDRs. CDRs and framework regions are those regions of an immunoglobulin variable domain as defined in the Kabat database of Sequences of Proteins of Immunological Interest.

Preferred human framework regions are those encoded by germline gene segments DP47 and DPK9. Advantageously, FW1, FW2 and FW3 of a V_H or V_L domain have the 15 sequence of FW1, FW2 or FW3 from DP47 or DPK9. The human frameworks may optionally contain mutations, for example up to about 5 amino acid changes or up to about 10 amino acid changes collectively in the human frameworks used in the ligands of the invention.

20 The variable domains in the multispecific ligands according to the second configuration of the invention may be arranged in an open or a closed conformation; that is, they may be arranged such that the variable domains can bind their cognate ligands independently and simultaneously, or such that only one of the variable domains may bind its cognate ligand at any one time.

25 The inventors have realised that under certain structural conditions, non-complementary variable domains (for example two light chain variable domains or two heavy chain variable domains) may be present in a ligand such that binding of a first epitope to a first variable domain inhibits the binding of a second epitope to a second variable domain, 30 even though such non-complementary domains do not operate together as a cognate pair.

Advantageously, the ligand comprises two or more pairs of variable domains; that is, it comprises at least four variable domains. Advantageously, the four variable domains comprise frameworks of human origin.

5 In a preferred embodiment, the human frameworks are identical to those of human germline sequences.

The present inventors consider that such antibodies will be of particular use in ligand binding assays for therapeutic and other uses.

10

Thus, in a first aspect of the second configuration, the present invention provides a method for producing a multispecific ligand comprising the steps of:

a) selecting a first epitope binding domain by its ability to bind to a first epitope,
b) selecting a second epitope binding domain by its ability to bind to a second
15 epitope,
c) combining the epitope binding domains; and
d) selecting the closed conformation multispecific ligand by its ability to bind to said first second epitope and said second epitope.

20

In a further aspect of the second configuration, the invention provides method for preparing a closed conformation multi-specific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity, wherein the first and second binding specificities compete for epitope binding such that the closed 25 conformation multi-specific ligand may not bind both epitopes simultaneously, said method comprising the steps of:

a) selecting a first epitope binding domain by its ability to bind to a first epitope,
b) selecting a second epitope binding domain by its ability to bind to a second
30 epitope,
c) combining the epitope binding domains such that the domains are in a closed conformation; and

d) selecting the closed conformation multispecific ligand by its ability to bind to said first second epitope and said second epitope, but not to both said first and second epitopes simultaneously.

5 Moreover, the invention provides a closed conformation multi-specific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity, wherein the first and second binding specificities compete for epitope binding such that the closed conformation multi-specific ligand may not bind both epitopes
10 simultaneously.

An alternative embodiment of the above aspect of the of the second configuration of the invention optionally comprises a further step (b1) comprising selecting a third or further epitope binding domain. In this way the multi-specific ligand produced, whether of open
15 or closed conformation, comprises more than two epitope binding specificities. In a preferred aspect of the second configuration of the invention, where the multi-specific ligand comprises more than two epitope binding domains, at least two of said domains are in a closed conformation and compete for binding; other domains may compete for binding or may be free to associate independently with their cognate epitope(s).

20 According to the present invention the term 'multi-specific ligand' refers to a ligand which possesses more than one epitope binding specificity as herein defined.

As herein defined the term 'closed conformation' (multi-specific ligand) means that the
25 epitope binding domains of the ligand are attached to or associated with each other, optionally by means of a protein skeleton, such that epitope binding by one epitope binding domain competes with epitope binding by another epitope binding domain. That is, cognate epitopes may be bound by each epitope binding domain individually but not simultaneously. The closed conformation of the ligand can be achieved using methods
30 herein described.

"Open conformation" means that the epitope binding domains of the ligand are attached to or associated with each other, optionally by means of a protein skeleton, such that

epitope binding by one epitope binding domain does not compete with epitope binding by another epitope binding domain.

As referred to herein, the term 'competes' means that the binding of a first epitope to its 5 cognate epitope binding domain is inhibited when a second epitope is bound to its cognate epitope binding domain. For example, binding may be inhibited sterically, for example by physical blocking of a binding domain or by alteration of the structure or environment of a binding domain such that its affinity or avidity for an epitope is reduced.

10 In a further embodiment of the second configuration of the invention, the epitopes may displace each other on binding. For example, a first epitope may be present on an antigen which, on binding to its cognate first binding domain, causes steric hindrance of a second binding domain, or a conformational change therein, which displaces the epitope bound to the second binding domain.

15 Advantageously, binding is reduced by 25% or more, advantageously 40%, 50%, 60%, 70%, 80%, 90% or more, and preferably up to 100% or nearly so, such that binding is completely inhibited. Binding of epitopes can be measured by conventional antigen binding assays, such as ELISA, by fluorescence based techniques, including FRET, or by 20 techniques such as surface plasmon resonance which measure the mass of molecules.

According to the method of the present invention, advantageously, each epitope binding domain is of a different epitope binding specificity.

25 In the context of the present invention, first and second "epitopes" are understood to be epitopes which are not the same and are not bound by a single monospecific ligand. They may be on different antigens or on the same antigen, but separated by a sufficient distance that they do not form a single entity that could be bound by a single mono-specific V_H/V_L binding pair of a conventional antibody. Experimentally, if both of the individual 30 variable domains in single chain antibody form (domain antibodies or dAbs) are separately competed by a monospecific V_H/V_L ligand against two epitopes then those two epitopes are not sufficiently far apart to be considered separate epitopes according to the present invention.

The closed conformation multispecific ligands of the invention do not include ligands as described in WO 02/02773. Thus, the ligands of the present invention do not comprise complementary V_H/V_L pairs which bind any one or more antigens or epitopes co-operatively. Instead, the ligands according to the invention preferably comprise non-complementary V_H-V_H or V_L-V_L pairs. Advantageously, each V_H or V_L domain in each V_H-V_H or V_L-V_L pair has a different epitope binding specificity, and the epitope binding sites are so arranged that the binding of an epitope at one site competes with the binding of an epitope at another site.

10

According to the present invention, advantageously, each epitope binding domain comprises an immunoglobulin variable domain. More advantageously, each immunoglobulin variable domain will be either a variable light chain domain (V_L) or a variable heavy chain domain V_H . In the second configuration of the present invention, 15 the immunoglobulin domains when present on a ligand according to the present invention are non-complementary, that is they do not associate to form a V_H/V_L antigen binding site. Thus, multi-specific ligands as defined in the second configuration of the invention comprise immunoglobulin domains of the same sub-type, that is either variable light chain domains (V_L) or variable heavy chain domains (V_H). Moreover, where the 20 ligand according to the invention is in the closed conformation, the immunoglobulin domains may be of the camelid V_{HH} type.

In an alternative embodiment, the ligand(s) according to the invention do not comprise a camelid V_{HH} domain. More particularly, the ligand(s) of the invention do not comprise 25 one or more amino acid residues that are specific to camelid V_{HH} domains as compared to human V_H domains.

Advantageously, the single variable domains are derived from antibodies selected for 30 binding activity against different antigens or epitopes. For example, the variable domains may be isolated at least in part by human immunisation. Alternative methods are known in the art, including isolation from human antibody libraries and synthesis of artificial antibody genes.

The variable domains advantageously bind superantigens, such as protein A or protein L. Binding to superantigens is a property of correctly folded antibody variable domains, and allows such domains to be isolated from, for example, libraries of recombinant or mutant domains.

5

Epitope binding domains according to the present invention comprise a protein scaffold and epitope interaction sites (which are advantageously on the surface of the protein scaffold).

10 Epitope binding domains may also be based on protein scaffolds or skeletons other than immunoglobulin domains. For example natural bacterial receptors such as SpA have been used as scaffolds for the grafting of CDRs to generate ligands which bind specifically to one or more epitopes. Details of this procedure are described in US 5,831,012. Other suitable scaffolds include those based on fibronectin and affibodies. Details of suitable procedures are described in WO 98/58965. Other suitable scaffolds include lipocalin and
15 CTLA4, as described in van den Beuken *et al.*, *J. Mol. Biol.* (2001) 310, 591-601, and scaffolds such as those described in WO0069907 (Medical Research Council), which are based for example on the ring structure of bacterial GroEL or other chaperone polypeptides.

20

Protein scaffolds may be combined; for example, CDRs may be grafted on to a CTLA4 scaffold and used together with immunoglobulin V_H or V_L domains to form a multivalent ligand. Likewise, fibronectin, lipocalin and other scaffolds may be combined.

25 It will be appreciated by one skilled in the art that the epitope binding domains of a closed conformation multispecific ligand produced according to the method of the present invention may be on the same polypeptide chain, or alternatively, on different polypeptide chains. In the case that the variable regions are on different polypeptide chains, then they may be linked via a linker, advantageously a flexible linker (such as a polypeptide chain),
30 a chemical linking group, or any other method known in the art.

The first and the second epitope binding domains may be associated either covalently or non-covalently. In the case that the domains are covalently associated, then the association may be mediated for example by disulphide bonds.

5 In the second configuration of the invention, the first and the second epitopes are preferably different. They may be, or be part of, polypeptides, proteins or nucleic acids, which may be naturally occurring or synthetic. In this respect, the ligand of the invention may bind an epitope or antigen and act as an antagonist or agonist (eg, EPO receptor agonist). The epitope binding domains of the ligand in one embodiment have the same 10 epitope specificity, and may for example simultaneously bind their epitope when multiple copies of the epitope are present on the same antigen. In another embodiment, these epitopes are provided on different antigens such that the ligand can bind the epitopes and bridge the antigens. One skilled in the art will appreciate that the choice of epitopes and antigens is large and varied. They may be for instance human or animal proteins, 15 cytokines, cytokine receptors, enzymes co-factors for enzymes or DNA binding proteins. Suitable cytokines and growth factors include but are not limited to: ApoE, Apo-SAA, BDNF, Cardiotrophin-1, EGF, EGF receptor, ENA-78, Eotaxin, Eotaxin-2, Exodus-2, EpoR, FGF-acidic, FGF-basic, fibroblast growth factor-10, FLT3 ligand, Fractalkine (CX3C), GDNF, G-CSF, GM-CSF, GF- β 1, insulin, IFN- γ , IGF-I, IGF-II, IL-1 α , IL-1 β , 20 IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8 (72 a.a.), IL-8 (77 a.a.), IL-9, IL-10, IL-11, IL-12, IL-13, IL-15, IL-16, IL-17, IL-18 (IGIF), Inhibin α , Inhibin β , IP-10, keratinocyte growth factor-2 (KGF-2), KGF, Leptin, LIF, Lymphotoxin, Mullerian inhibitory substance, monocyte colony inhibitory factor, monocyte attractant protein, M-CSF, MDC (67 a.a.), MDC (69 a.a.), MCP-1 (MCAF), MCP-2, MCP-3, MCP-4, MDC (67 a.a.), MDC (69 a.a.), MIG, MIP-1 α , MIP-1 β , MIP-3 α , MIP-3 β , MIP-4, myeloid progenitor inhibitor 25 factor-1 (MPIF-1), NAP-2, Neurturin, Nerve growth factor, β -NGF, NT-3, NT-4, Oncostatin M, PDGF-AA, PDGF-AB, PDGF-BB, PF-4, RANTES, SDF1 α , SDF1 β , SCF, SCGF, stem cell factor (SCF), TARC, TGF- α , TGF- β , TGF- β 2, TGF- β 3, tumour necrosis factor (TNF), TNF- α , TNF- β , TNF receptor I, TNF receptor II, TNIL-1, TPO, VEGF, 30 VEGF receptor 1, VEGF receptor 2, VEGF receptor 3, GCP-2, GRO/MGSA, GRO- β , GRO- γ , HCC1, 1-309, HER 1, HER 2, HER 3, HER 4, TACE recognition site, TNF BP-I and TNF BP-II, as well as any target disclosed in Annex 2 or Annex 3 hereto, whether in combination as set forth in the Annexes, in a different combination or individually.

Cytokine receptors include receptors for the foregoing cytokines, e.g. IL-1 R1; IL-6R; IL-10R; IL-18R, as well as receptors for cytokines set forth in Annex 2 or Annex 3 and also receptors disclosed in Annex 2 and 3. It will be appreciated that this list is by no means exhaustive. Where the multispecific ligand binds to two epitopes (on the same or 5 different antigens), the antigen(s) may be selected from this list.

Advantageously, dual specific ligands may be used to target cytokines and other molecules which cooperate synergistically in therapeutic situations in the body of an organism. The invention therefore provides a method for synergising the activity of two 10 or more cytokines, comprising administering a dual specific ligand capable of binding to said two or more cytokines. In this aspect of the invention, the dual specific ligand may be any dual specific ligand, including a ligand composed of complementary and/or non-complementary domains, a ligand in an open conformation, and a ligand in a closed conformation. For example, this aspect of the invention relates to combinations of V_H 15 domains and V_L domains, V_H domains only and V_L domains only.

Synergy in a therapeutic context may be achieved in a number of ways. For example, target combinations may be therapeutically active only if both targets are targeted by the ligand, whereas targeting one target alone is not therapeutically effective. In another 20 embodiment, one target alone may provide some low or minimal therapeutic effect, but together with a second target the combination provides a synergistic increase in therapeutic effect.

Preferably, the cytokines bound by the dual specific ligands of this aspect of the invention 25 are selected from the list shown in Annex 2.

Moreover, dual specific ligands may be used in oncology applications, where one specificity targets CD89, which is expressed by cytotoxic cells, and the other is tumour specific. Examples of tumour antigens which may be targeted are given in Annex 3.

30 In one embodiment of the second configuration of the invention, the variable domains are derived from an antibody directed against the first and/or second antigen or epitope. In a preferred embodiment the variable domains are derived from a repertoire of single

variable antibody domains. In one example, the repertoire is a repertoire that is not created in an animal or a synthetic repertoire. In another example, the single variable domains are not isolated (at least in part) by animal immunisation. Thus, the single domains can be isolated from a naïve library.

5

The second configuration of the invention, in another aspect, provides a multi-specific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity. The first and second binding specificities may be the same or

10 different.

In a further aspect, the present invention provides a closed conformation multi-specific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity wherein the first and second binding specificities are capable of competing for epitope binding such that the closed conformation multi-specific ligand cannot bind both epitopes simultaneously.

15

In a still further aspect, the invention provides open conformation ligands comprising non-complementary binding domains, wherein the domains are specific for a different epitope on the same target. Such ligands bind to targets with increased avidity. Similarly, the invention provides multivalent ligands comprising non-complementary binding domains specific for the same epitope and directed to targets which comprise multiple copies of said epitope, such as IL-5, PDGF-AA, PDGF-BB, TGF beta, TGF 25 beta2, TGF beta3 and TNF α , for example human TNF Receptor 1 and human TNF α .

20

In a similar aspect, ligands according to the invention can be configured to bind individual epitopes with low affinity, such that binding to individual epitopes is not therapeutically significant; but the increased avidity resulting from binding to two epitopes provides a therapeutic benefit. In a particular example, epitopes may be targeted which are present individually on normal cell types, but present together only on abnormal or diseased cells, such as tumour cells. In such a situation, only the abnormal or

30

diseased cells are effectively targetted by the bispecific ligands according to the invention.

Ligand specific for multiple copies of the same epitope, or adjacent epitopes, on the same

5 target (known as chelating dAbs) may also be trimeric or polymeric (tertrameric or more) ligands comprising three, four or more non-complementary binding domains. For example, ligands may be constructed comprising three or four V_H domains or V_L domains.

10 Moreover, ligands are provided which bind to multisubunit targets, wherein each binding domain is specific for a subunit of said target. The ligand may be dimeric, trimeric or polymeric.

Preferably, the multi-specific ligands according to the above aspects of the invention are

15 obtainable by the method of the first aspect of the invention.

According to the above aspect of the second configuration of the invention, advantageously the first epitope binding domain and the second epitope binding domains are non-complementary immunoglobulin variable domains, as herein defined. That is

20 either V_H-V_H or V_L-V_L variable domains.

Chelating dAbs in particular may be prepared according to a preferred aspect of the invention, namely the use of anchor dAbs, in which a library of dimeric, trimeric or multimeric dAbs is constructed using a vector which comprises a constant dAb upstream or downstream of a linker sequence, with a repertoire of second, third and further dAbs being inserted on the other side of the linker. For example, the anchor or guiding dAb may be TAR1-5 (V κ), TAR1-27(V κ), TAR2h-5(VH) or TAR2h-6(V κ).

In alternative methodologies, the use of linkers may be avoided, for example by the use of 30 non-covalent bonding or natural affinity between binding domains such as V_H and V κ . The invention accordingly provides a method for preparing a chelating multimeric ligand comprising the steps of:

(a) providing a vector comprising a nucleic acid sequence encoding a single binding domain specific for a first epitope on a target;

(b) providing a vector encoding a repertoire comprising second binding domains specific for a second epitope on said target, which epitope can be the same or different to the first epitope, said second epitope being adjacent to said first epitope; and

5 (c) expressing said first and second binding domains; and

(d) isolating those combinations of first and second binding domains which combine together to produce a target-binding dimer.

10 The first and second epitopes are adjacent such that a multimeric ligand is capable of binding to both epitopes simultaneously. This provides the ligand with the advantages of increased avidity if binding. Where the epitopes are the same, the increased avidity is obtained by the presence of multiple copies of the epitope on the target, allowing at least two copies to be simultaneously bound in order to obtain the increased avidity effect.

15 The binding domains may be associated by several methods, as well as the use of linkers. For example, the binding domains may comprise cys residues, avidin and streptavidin groups or other means for non-covalent attachment post-synthesis; those combinations which bind to the target efficiently will be isolated. Alternatively, a linker may be present
20 between the first and second binding domains, which are expressed as a single polypeptide from a single vector, which comprises the first binding domain, the linker and a repertoire of second binding domains, for instance as described above.

25 In a preferred aspect, the first and second binding domains associate naturally when bound to antigen; for example, V_H and V_{κ} domains, when bound to adjacent epitopes, will naturally associate in a three-way interaction to form a stable dimer. Such associated proteins can be isolated in a target binding assay. An advantage of this procedure is that only binding domains which bind to closely adjacent epitopes, in the correct conformation, will associate and thus be isolated as a result of their increased avidity for
30 the target.

In an alternative embodiment of the above aspect of the second configuration of the invention, at least one epitope binding domain comprises a non-immunoglobulin 'protein

'scaffold' or 'protein skeleton' as herein defined. Suitable non-immunoglobulin protein scaffolds include but are not limited to any of those selected from the group consisting of: SpA, fibronectin, GroEL and other chaperones, lipocalin, CCLTA4 and affibodies, as set forth above.

5

According to the above aspect of the second configuration of the invention, advantageously, the epitope binding domains are attached to a 'protein skeleton'. Advantageously, a protein skeleton according to the invention is an immunoglobulin skeleton.

10

According to the present invention, the term 'immunoglobulin skeleton' refers to a protein which comprises at least one immunoglobulin fold and which acts as a nucleus for one or more epitope binding domains, as defined herein.

15

Preferred immunoglobulin skeletons as herein defined includes any one or more of those selected from the following: an immunoglobulin molecule comprising at least (i) the CL (kappa or lambda subclass) domain of an antibody; or (ii) the CH1 domain of an antibody heavy chain; an immunoglobulin molecule comprising the CH1 and CH2 domains of an antibody heavy chain; an immunoglobulin molecule comprising the CH1, CH2 and CH3 domains of an antibody heavy chain; or any of the subset (ii) in conjunction with the CL (kappa or lambda subclass) domain of an antibody. A hinge region domain may also be included. Such combinations of domains may, for example, mimic natural antibodies, such as IgG or IgM, or fragments thereof, such as Fv, scFv, Fab or F(ab')₂ molecules. Those skilled in the art will be aware that this list is not intended to be exhaustive.

20

Linking of the skeleton to the epitope binding domains, as herein defined may be achieved at the polypeptide level, that is after expression of the nucleic acid encoding the skeleton and/or the epitope binding domains. Alternatively, the linking step may be performed at the nucleic acid level. Methods of linking a protein skeleton according to the present invention, to the one or more epitope binding domains include the use of protein chemistry and/or molecular biology techniques which will be familiar to those skilled in the art and are described herein.

Advantageously, the closed conformation multispecific ligand may comprise a first domain capable of binding a target molecule, and a second domain capable of binding a molecule or group which extends the half-life of the ligand. For example, the molecule or group may be a bulky agent, such as HSA or a cell matrix protein. As used herein, the 5 phrase "molecule or group which extends the half-life of a ligand" refers to a molecule or chemical group which, when bound by a dual-specific ligand as described herein increases the in vivo half-life of such dual specific ligand when administered to an animal, relative to a ligand that does not bind that molecule or group. Examples of molecules or groups that extend the half-life of a ligand are described hereinbelow. In a 10 preferred embodiment, the closed conformation multispecific ligand may be capable of binding the target molecule only on displacement of the half-life enhancing molecule or group. Thus, for example, a closed conformation multispecific ligand is maintained in circulation in the bloodstream of a subject by a bulky molecule such as HSA. When a target molecule is encountered, competition between the binding domains of the closed 15 conformation multispecific ligand results in displacement of the HSA and binding of the target.

Ligands according to any aspect of the present invention, as well as dAb monomers useful in constructing such ligands, may advantageously dissociate from their cognate 20 target(s) with a K_d of 300nM to 5pM (ie, 3×10^{-7} to 5×10^{-12} M), preferably 50nM to 20pM, or 5nM to 200pM or 1nM to 100pM, 1×10^{-7} M or less, 1×10^{-8} M or less, 1×10^{-9} M or less, 1×10^{-10} M or less, 1×10^{-11} M or less; and/or a K_{off} rate constant of 5×10^{-1} to 1×10^{-7} S⁻¹, preferably 1×10^{-2} to 1×10^{-6} S⁻¹, or 5×10^{-3} to 1×10^{-5} S⁻¹, or 5×10^{-1} S⁻¹ or less, or 1×10^{-2} S⁻¹ or less, or 1×10^{-3} S⁻¹ or less, or 1×10^{-4} S⁻¹ or less, or 1×10^{-5} S⁻¹ 25 or less, or 1×10^{-6} S⁻¹ or less as determined by surface plasmon resonance. The K_d rate constant is defined as K_{off}/K_{on} .

In particular the invention provides an anti-TNF α dAb monomer (or dual specific ligand comprising such a dAb), homodimer, heterodimer or homotrimer ligand, wherein each 30 dAb binds TNF α . The ligand binds to TNF α with a K_d of 300nM to 5pM (ie, 3×10^{-7} to 5×10^{-12} M), preferably 50nM to 20pM, more preferably 5nM to 200pM and most preferably 1nM to 100pM; expressed in an alternative manner, the K_d is 1×10^{-7} M or less, preferably 1×10^{-8} M or less, more preferably 1×10^{-9} M or less, advantageously 1 x

10⁻¹⁰ M or less and most preferably 1 x 10⁻¹¹ M or less; and/or a K_{off} rate constant of 5 x 10⁻¹ to 1 x 10⁻⁷ S⁻¹, preferably 1 x 10⁻² to 1 x 10⁻⁶ S⁻¹, more preferably 5 x 10⁻³ to 1 x 10⁻⁵ S⁻¹, for example 5 x 10⁻¹ S⁻¹ or less, preferably 1 x 10⁻² S⁻¹ or less, more preferably 1 x 10⁻³ S⁻¹ or less, advantageously 1 x 10⁻⁴ S⁻¹ or less, further advantageously 1 x 10⁻⁵ S⁻¹ or less, 5 and most preferably 1 x 10⁻⁶ S⁻¹ or less, as determined by surface plasmon resonance.

Preferably, the ligand neutralises TNF α in a standard L929 assay with an ND50 of 500nM to 50pM, preferably or 100nM to 50pM, advantageously 10nM to 100pM, more preferably 1nM to 100pM; for example 50nM or less, preferably 5nM or less, 10 advantageously 500pM or less, more preferably 200pM or less and most preferably 100pM or less.

Preferably, the ligand inhibits binding of TNF alpha to TNF alpha Receptor I (p55 receptor) with an IC50 of 500nM to 50pM, preferably 100nM to 50pM, more preferably 15 10nM to 100pM, advantageously 1nM to 100pM; for example 50nM or less, preferably 5nM or less, more preferably 500pM or less, advantageously 200pM or less, and most preferably 100pM or less. Preferably, the TNF α is Human TNF α .

Furthermore, the invention provides a an anti-TNF Receptor I dAb monomer, or dual 20 specific ligand comprising such a dAb, that binds to TNF Receptor I with a K_d of 300nM to 5pM (ie, 3 x 10⁻⁷ to 5 x 10⁻¹² M), preferably 50nM to 20pM, more preferably 5nM to 200pM and most preferably 1nM to 100pM, for example 1 x 10⁻⁷ M or less, preferably 1 x 10⁻⁸ M or less, more preferably 1 x 10⁻⁹ M or less, advantageously 1 x 10⁻¹⁰ M or less and most preferably 1 x 10⁻¹¹ M or less; and/or a K_{off} rate constant of 5 x 10⁻¹ to 1 x 10⁻⁷ S⁻¹, preferably 1 x 10⁻² to 1 x 10⁻⁶ S⁻¹, more preferably 5 x 10⁻³ to 1 x 10⁻⁵ S⁻¹, for 25 example 5 x 10⁻¹ S⁻¹ or less, preferably 1 x 10⁻² S⁻¹ or less, advantageously 1 x 10⁻³ S⁻¹ or less, more preferably 1 x 10⁻⁴ S⁻¹ or less, still more preferably 1 x 10⁻⁵ S⁻¹ or less, and most preferably 1 x 10⁻⁶ S⁻¹ or less as determined by surface plasmon resonance.

30 Preferably, the dAb monomer or ligand neutralises TNF α in a standard assay (eg, the L929 or HeLa assays described herein) with an ND50 of 500nM to 50pM, preferably 100nM to 50pM, more preferably 10nM to 100pM, advantageously 1nM to 100pM; for

example 50nM or less, preferably 5nM or less, more preferably 500pM or less, advantageously 200pM or less, and most preferably 100pM or less.

Preferably, the dAb monomer or ligand inhibits binding of TNF alpha to TNF alpha Receptor I (p55 receptor) with an IC₅₀ of 500nM to 50pM, preferably 100nM to 50pM, more preferably 10nM to 100pM, advantageously 1nM to 100pM; for example 50nM or less, preferably 5nM or less, more preferably 500pM or less, advantageously 200pM or less, and most preferably 100pM or less. Preferably, the TNF Receptor I target is Human TNF α .

10

Furthermore, the invention provides a dAb monomer(or dual specific ligand comprising such a dAb) that binds to serum albumin (SA) with a K_d of 1nM to 500 μ M (ie, $\times 10^{-9}$ to 5×10^{-4}), preferably 100nM to 10 μ M. Preferably, for a dual specific ligand comprising a first anti-SA dAb and a second dAb to another target, the affinity (eg K_d and/or K_{off} as measured by surface plasmon resonance, eg using BiaCore) of the second dAb for its target is from 1 to 100000 times (preferably 100 to 100000, more preferably 1000 to 100000, or 10000 to 100000 times) the affinity of the first dAb for SA. For example, the first dAb binds SA with an affinity of approximately 10 μ M, while the second dAb binds its target with an affinity of 100pM. Preferably, the serum albumin is human serum albumin (HSA).

In one embodiment, the first dAb (or a dAb monomer) binds SA (eg, HSA) with a K_d of approximately 50, preferably 70, and more preferably 100, 150 or 200 nM.

25 The invention moreover provides dimers, trimers and polymers of the aforementioned dAb monomers, in accordance with the foregoing aspect of the present invention.

Ligands according to the invention, including dAb monomers, dimers and trimers, can be linked to an antibody Fc region, comprising one or both of C_H2 and C_H3 domains, and 30 optionally a hinge region. For example, vectors encoding ligands linked as a single nucleotide sequence to an Fc region may be used to prepare such polypeptides.

In a further aspect of the second configuration of the invention, the present invention provides one or more nucleic acid molecules encoding at least a multispecific ligand as herein defined. In one embodiment, the ligand is a closed conformation ligand. In another embodiment, it is an open conformation ligand. The multispecific ligand may be 5 encoded on a single nucleic acid molecule; alternatively, each epitope binding domain may be encoded by a separate nucleic acid molecule. Where the ligand is encoded by a single nucleic acid molecule, the domains may be expressed as a fusion polypeptide, or may be separately expressed and subsequently linked together, for example using chemical linking agents. Ligands expressed from separate nucleic acids will be linked 10 together by appropriate means.

The nucleic acid may further encode a signal sequence for export of the polypeptides from a host cell upon expression and may be fused with a surface component of a filamentous bacteriophage particle (or other component of a selection display system) 15 upon expression. Leader sequences, which may be used in bacterial expression and/or phage or phagemid display, include pelB, stII, ompA, phoA, bla and pelA.

In a further aspect of the second configuration of the invention the present invention provides a vector comprising nucleic acid according to the present invention.

20 In a yet further aspect, the present invention provides a host cell transfected with a vector according to the present invention.

25 Expression from such a vector may be configured to produce, for example on the surface of a bacteriophage particle, epitope binding domains for selection. This allows selection of displayed domains and thus selection of 'multispecific ligands' using the method of the present invention.

30 In a preferred embodiment of the second configuration of the invention, the epitope binding domains are immunoglobulin variable regions and are selected from single domain V gene repertoires. Generally the repertoire of single antibody domains is displayed on the surface of filamentous bacteriophage. In a preferred embodiment each single antibody domain is selected by binding of a phage repertoire to antigen.

The present invention further provides a kit comprising at least a multispecific ligand according to the present invention, which may be an open conformation or closed conformation ligand. Kits according to the invention may be, for example, diagnostic 5 kits, therapeutic kits, kits for the detection of chemical or biological species, and the like.

In a further aspect still of the second configuration of the invention, the present invention provides a homogenous immunoassay using a ligand according to the present invention.

- 10 In a further aspect still of the second configuration of the invention, the present invention provides a composition comprising a closed conformation multispecific ligand, obtainable by a method of the present invention, and a pharmaceutically acceptable carrier, diluent or excipient.
- 15 Moreover, the present invention provides a method for the treatment of disease using a 'closed conformation multispecific ligand' or a composition according to the present invention.

In a preferred embodiment of the invention the disease is cancer or an inflammatory 20 disease, eg rheumatoid arthritis, asthma or Crohn's disease.

- 25 In a further aspect of the second configuration of the invention, the present invention provides a method for the diagnosis, including diagnosis of disease using a closed conformation multispecific ligand, or a composition according to the present invention.
- Thus in general the binding of an analyte to a closed conformation multispecific ligand may be exploited to displace an agent, which leads to the generation of a signal on displacement. For example, binding of analyte (second antigen) could displace an enzyme (first antigen) bound to the antibody providing the basis for an immunoassay, especially if the enzyme were held to the antibody through its active site.

30 Thus in a final aspect of the second configuration, the present invention provides a method for detecting the presence of a target molecule, comprising:

- (a) providing a closed conformation multispecific ligand bound to an agent, said ligand being specific for the target molecule and the agent, wherein the agent which is bound by the ligand leads to the generation of a detectable signal on displacement from the ligand;
- (b) exposing the closed conformation multispecific ligand to the target molecule; and
- 5 (c) detecting the signal generated as a result of the displacement of the agent.

According to the above aspect of the second configuration of the invention, advantageously, the agent is an enzyme, which is inactive when bound by the closed conformation multi-specific ligand. Alternatively, the agent may be any one or more 10 selected from the group consisting of the following: the substrate for an enzyme, and a fluorescent, luminescent or chromogenic molecule which is inactive or quenched when bound by the ligand.

Sequences similar or homologous (e.g., at least about 70% sequence identity) to the 15 sequences disclosed herein are also part of the invention. In some embodiments, the sequence identity at the amino acid level can be about 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or higher. At the nucleic acid level, the sequence identity can be about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or higher. Alternatively, substantial identity exists when the nucleic acid 20 segments will hybridize under selective hybridization conditions (e.g., very high stringency hybridization conditions), to the complement of the strand. The nucleic acids may be present in whole cells, in a cell lysate, or in a partially purified or substantially pure form.

25 Calculations of "homology" or "sequence identity" or "similarity" between two sequences (the terms are used interchangeably herein) are performed as follows. The sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes).

30 In a preferred embodiment, the length of a reference sequence aligned for comparison purposes is at least 30%, preferably at least 40%, more preferably at least 50%, even more preferably at least 60%, and even more preferably at least 70%, 80%, 90%, 100% of the length of the reference sequence. The amino acid residues or nucleotides at

corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

5

10 Advantageously, the BLAST algorithm (version 2.0) is employed for sequence alignment, with parameters set to default values. The BLAST algorithm is described in detail at the world wide web site ("www") of the National Center for Biotechnology Information ("ncbi") of the National Institutes of Health ("nih") of the U.S. government ("gov"), in the "/Blast/" directory, in the "blast_help.html" file. The search parameters are defined as

15 follows, and are advantageously set to the defined default parameters.

BLAST (Basic Local Alignment Search Tool) is the heuristic search algorithm employed by the programs blastp, blastn, blastx, tblastn, and tblastx; these programs ascribe significance to their findings using the statistical methods of Karlin and Altschul, 1990, 20 Proc. Natl. Acad. Sci. USA 87(6):2264-8 (see the "blast_help.html" file, as described above) with a few enhancements. The BLAST programs were tailored for sequence similarity searching, for example to identify homologues to a query sequence. The programs are not generally useful for motif-style searching. For a discussion of basic issues in similarity searching of sequence databases, see Altschul et al. (1994).

25

The five BLAST programs available at the National Center for Biotechnology Information web site perform the following tasks:

"blastp" compares an amino acid query sequence against a protein sequence database;

"blastn" compares a nucleotide query sequence against a nucleotide sequence database;

30 "blastx" compares the six-frame conceptual translation products of a nucleotide query sequence (both strands) against a protein sequence database;

"tblastn" compares a protein query sequence against a nucleotide sequence database dynamically translated in all six reading frames (both strands).

“tblastx” compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database.

BLAST uses the following search parameters:

HISTOGRAM Display a histogram of scores for each search; default is yes. (See parameter H in the BLAST Manual).

DESCRIPTIONS Restricts the number of short descriptions of matching sequences reported to the number specified; default limit is 100 descriptions. (See parameter V in the manual page). See also EXPECT and CUTOFF.

ALIGNMENTS Restricts database sequences to the number specified for which high-scoring segment pairs (HSPs) are reported; the default limit is 50. If more database sequences than this happen to satisfy the statistical significance threshold for reporting (see EXPECT and CUTOFF below), only the matches ascribed the greatest statistical significance are reported. (See parameter B in the BLAST Manual).

EXPECT The statistical significance threshold for reporting matches against database sequences; the default value is 10, such that 10 matches are expected to be found merely by chance, according to the stochastic model of Karlin and Altschul (1990). If the statistical significance ascribed to a match is greater than the EXPECT threshold, the match will not be reported. Lower EXPECT thresholds are more stringent, leading to fewer chance matches being reported. Fractional values are acceptable. (See parameter E in the BLAST Manual).

CUTOFF Cutoff score for reporting high-scoring segment pairs. The default value is calculated from the EXPECT value (see above). HSPs are reported for a database sequence only if the statistical significance ascribed to them is at least as high as would be ascribed to a lone HSP having a score equal to the CUTOFF value. Higher CUTOFF values are more stringent, leading to fewer chance matches being reported. (See parameter S in the BLAST Manual). Typically, significance thresholds can be more intuitively managed using EXPECT.

MATRIX Specify an alternate scoring matrix for BLASTP, BLASTX, TBLASTN and TBLASTX. The default matrix is BLOSUM62 (Henikoff & Henikoff, 1992, Proc. Natl. Acad. Sci. USA 89(22):10915-9). The valid alternative choices include: PAM40, PAM120, PAM250 and IDENTITY. No alternate scoring matrices are available for BLASTN; specifying the MATRIX directive in BLASTN requests returns an error response.

STRAND Restrict a TBLASTN search to just the top or bottom strand of the database sequences; or restrict a BLASTN, BLASTX or TBLASTX search to just reading frames on the top or bottom strand of the query sequence.

FILTER Mask off segments of the query sequence that have low compositional

5 complexity, as determined by the SEG program of Wootton & Federhen (1993) Computers and Chemistry 17:149-163, or segments consisting of short-periodicity internal repeats, as determined by the XNU program of Claverie & States, 1993, Computers and Chemistry 17:191-201, or, for BLASTN, by the DUST program of Tatusov and Lipman (see the world wide web site of the NCBI). Filtering can eliminate 10 statistically significant but biologically uninteresting reports from the blast output (e.g., hits against common acidic-, basic- or proline-rich regions), leaving the more biologically interesting regions of the query sequence available for specific matching against database sequences.

15 Low complexity sequence found by a filter program is substituted using the letter "N" in nucleotide sequence (e.g., "N" repeated 13 times) and the letter "X" in protein sequences (e.g., "X" repeated 9 times).

Filtering is only applied to the query sequence (or its translation products), not to 20 database sequences. Default filtering is DUST for BLASTN, SEG for other programs.

It is not unusual for nothing at all to be masked by SEG, XNU, or both, when applied to sequences in SWISS-PROT, so filtering should not be expected to always yield an effect. Furthermore, in some cases, sequences are masked in their entirety, indicating that the 25 statistical significance of any matches reported against the unfiltered query sequence should be suspect.

NCBI-gi Causes NCBI gi identifiers to be shown in the output, in addition to the accession and/or locus name.

30 Most preferably, sequence comparisons are conducted using the simple BLAST search algorithm provided at the NCBI world wide web site described above, in the "/BLAST" directory.

Brief Description of the Figures

5 Figure 1 shows the diversification of V_H /HSA at positions H50, H52, H52a, H53, H55, H56, H58, H95, H96, H97, H98 (DVT or NNK encoded respectively) which are in the antigen binding site of V_H HSA. The sequence of V_K is diversified at positions L50, L53.

10 Figure 2 shows Library 1: Germline V_K /DVT V_H ,
Library 2: Germline V_K /NNK V_H ,
Library 3: Germline V_H /DVT V_K
Library 4: Germline V_H /NNK V_K

15 In phage display/ScFv format. These libraries were pre-selected for binding to generic ligands protein A and protein L so that the majority of the clones and selected libraries are functional. Libraries were selected on HSA (first round) and β -gal (second round) or HSA β -gal selection or on β -gal (first round) and HSA (second round) β -gal HSA selection. Soluble scFv from these clones of PCR are amplified in the sequence. One clone encoding a dual specific antibody K8 was chosen for further work.

20 Figure 3 shows an alignment of V_H chains and V_K chains.

25 Figure 4 shows the characterisation of the binding properties of the K8 antibody, the binding properties of the K8 antibody characterised by monoclonal phage ELISA, the dual specific K8 antibody was found to bind HSA and β -gal and displayed on the surface of the phage with absorbant signals greater than 1.0. No cross reactivity with other proteins was detected.

30 Figure 5 shows soluble scFv ELISA performed using known concentrations of the K8 antibody fragment. A 96-well plate was coated with 100 μ g of HSA, BSA and β -gal at 10 μ g/ml and 100 μ g/ml of Protein A at 1 μ g/ml concentration. 50 μ g of the serial dilutions of the K8 scFv was applied and the bound antibody fragments were detected with Protein L-HRP. ELISA results confirm the dual specific nature of the K8 antibody.

5 Figure 6 shows the binding characteristics of the clone K8V_K/dummy V_H analysed using soluble scFv ELISA. Production of the soluble scFv fragments was induced by IPTG as described by Harrison *et al*, Methods Enzymol.

10 1996;267:83-109 and the supernatant containing scFv assayed directly. Soluble scFv ELISA is performed as described in example 1 and the bound scFvs were detected with Protein L-HRP. The ELISA results revealed that this clone was still able to bind β -gal, whereas binding BSA was abolished.

15 Figure 7 shows the sequence of variable domain vectors 1 and 2.

20 Figure 8 is a map of the C_H vector used to construct a V_H1/V_H2 multispecific ligand.

25 Figure 9 is a map of the V_K vector used to construct a V_K1/V_K2 multispecific ligand.

30 Figure 10 TNF receptor assay comparing TAR1-5 dimer 4, TAR1-5-19 dimer 4 and TAR1-5-19 monomer.

35 Figure 11 TNF receptor assay comparing TAR1-5 dimers 1-6. All dimers have been FPLC purified and the results for the optimal dimeric species are shown.

40 Figure 12 TNF receptor assay of TAR1-5 19 homodimers in different formats: dAb-linker-dAb format with 3U, 5U or 7U linker, Fab format and cysteine hinge linker format.

45 Figure 13 Dummy VH sequence for library 1. The sequence of the VH framework based on germline sequence DP47 - JH4b. Positions where NNK randomisation (N=A or T or C or G nucleotides; K=G or T nucleotides) has been incorporated into library 1 are indicated in bold underlined text.

50 Figure 14 Dummy VH sequence for library 2. The sequence of the VH framework based on germline sequence DP47 - JH4b. Positions where NNK

randomisation (N=A or T or C or G nucleotides; K = G or T nucleotides) has been incorporated into library 2 are indicated in bold underlined text.

5 Figure 15 Dummy $V\kappa$ sequence for library 3. The sequence of the $V\kappa$ framework based on germline sequence DP $K9$ - J $\kappa 1$. Positions where NNK randomisation (N=A or T or C or G nucleotides; K = G or T nucleotides) has been incorporated into library 3 are indicated in bold underlined text.

10 Figure 16 Nucleotide and amino acid sequence of anti MSA dAbs MSA 16 and MSA 26.

15 Figure 17 Inhibition biacore of MSA 16 and 26. Purified dAbs MSA16 and MSA26 were analysed by inhibition biacore to determine K_d . Briefly, the dAbs were tested to determine the concentration of dAb required to achieve 200RUs of response on a biacore CM5 chip coated with a high density of MSA. Once the required concentrations of dAb had been determined, MSA antigen at a range of concentrations around the expected K_d was premixed with the dAb and incubated overnight. Binding to the MSA coated biacore chip of dAb in each of the premixes was then measured at a 20 high flow-rate of 30 μ l/minute.

25 Figure 18 Serum levels of MSA16 following injection. Serum half life of the dAb MSA16 was determined in mouse. MSA16 was dosed as single i.v. injections at approx 1.5mg/kg into CD1 mice. Modelling with a 2 compartment model showed MSA16 had a $t_{1/2\alpha}$ of 0.98hr, a $t_{1/2\beta}$ of 36.5hr and an AUC of 913hr.mg/ml. MSA16 had a considerably lengthened half life compared with HEL4 (an anti-hen egg white lysozyme dAb) which had a $t_{1/2\alpha}$ of 0.06hr and a $t_{1/2\beta}$ of 0.34hr.

30 Figure 19 ELISA (a) and TNF receptor assay (c) showing inhibition of TNF binding with a Fab-like fragment comprising MSA26Ck and TAR1-5-19CH. Addition of MSA with the Fab-like fragment reduces the level of inhibition. An ELISA plate coated with 1 μ g/ml TNF α was probed with

5 dual specific $V\kappa C_H$ and $V\kappa C\kappa$ Fab like fragment and also with a control TNF α binding dAb at a concentration calculated to give a similar signal on the ELISA. Both the dual specific and control dAb were used to probe the ELISA plate in the presence and in the absence of 2mg/ml MSA. The signal in the dual specific well was reduced by more than 50% but the signal in the dAb well was not reduced at all (see figure 19a). The same dual specific protein was also put into the receptor assay with and without MSA and competition by MSA was also shown (see figure 19c). This demonstrates that binding of MSA to the dual specific is competitive with binding to TNF α .

10

Figure 20 TNF receptor assay showing inhibiton of TNF binding with a disulphide bonded heterodimer of TAR1-5-19 dAb and MSA16 dAb. Addition of MSA with the dimer reduces the level of inhibiton in a dose dependant manner. The TNF receptor assay (figure 19 (b)) was conducted in the presence of a constant concentration of heterodimer (18nM) and a dilution series of MSA and HSA. The presence of HSA at a range of concentrations (up to 2 mg/ml) did not cause a reduction in the ability of the dimer to inhibit TNF α . However, the addition of MSA caused a dose dependant reduction in the ability of the dimer to inhibit TNF α (figure 19a). This demonstrates that MSA and TNF α compete for binding to the cys bonded TAR1-5-19, MSA16 dimer. MSA and HSA alone did not have an effect on the TNF binding level in the assay.

25

Detailed Description of the Invention

Definitions

30 **Complementary** Two immunoglobulin domains are “complementary” where they belong to families of structures which form cognate pairs or groups or are derived from such families and retain this feature. For example, a V_H domain and a V_L domain of an antibody are complementary; two V_H domains are not complementary, and two V_L

domains are not complementary. Complementary domains may be found in other members of the immunoglobulin superfamily, such as the V_α and V_β (or γ and δ) domains of the T-cell receptor. In the context of the second configuration of the present invention, non-complementary domains do not bind a target molecule cooperatively, but act 5 independently on different target epitopes which may be on the same or different molecules. Domains which are artificial, such as domains based on protein scaffolds which do not bind epitopes unless engineered to do so, are non-complementary. Likewise, two domains based on (for example) an immunoglobulin domain and a fibronectin domain are not complementary.

10

Immunoglobulin This refers to a family of polypeptides which retain the immunoglobulin fold characteristic of antibody molecules, which contains two β sheets and, usually, a conserved disulphide bond. Members of the immunoglobulin superfamily are involved in many aspects of cellular and non-cellular interactions *in vivo*, including 15 widespread roles in the immune system (for example, antibodies, T-cell receptor molecules and the like), involvement in cell adhesion (for example the ICAM molecules) and intracellular signalling (for example, receptor molecules, such as the PDGF receptor). The present invention is applicable to all immunoglobulin superfamily molecules which possess binding domains. Preferably, the present invention relates to antibodies.

20

Combining Variable domains according to the invention are combined to form a group of domains; for example, complementary domains may be combined, such as V_L domains being combined with V_H domains. Non-complementary domains may also be combined. Domains may be combined in a number of ways, involving linkage of the domains by 25 covalent or non-covalent means.

Domain A domain is a folded protein structure which retains its tertiary structure independently of the rest of the protein. Generally, domains are responsible for discrete functional properties of proteins, and in many cases may be added, removed or 30 transferred to other proteins without loss of function of the remainder of the protein and/or of the domain. By single antibody variable domain is meant a folded polypeptide domain comprising sequences characteristic of antibody variable domains. It therefore includes complete antibody variable domains and modified variable domains, for example

in which one or more loops have been replaced by sequences which are not characteristic of antibody variable domains, or antibody variable domains which have been truncated or comprise N- or C-terminal extensions, as well as folded fragments of variable domains which retain at least in part the binding activity and specificity of the full-length domain.

5

Repertoire A collection of diverse variants, for example polypeptide variants which differ in their primary sequence. A library used in the present invention will encompass a repertoire of polypeptides comprising at least 1000 members.

10 **Library** The term library refers to a mixture of heterogeneous polypeptides or nucleic acids. The library is composed of members, each of which have a single polypeptide or nucleic acid sequence. To this extent, *library* is synonymous with *repertoire*. Sequence differences between library members are responsible for the diversity present in the library. The library may take the form of a simple mixture of 15 polypeptides or nucleic acids, or may be in the form of organisms or cells, for example bacteria, viruses, animal or plant cells and the like, transformed with a library of nucleic acids. Preferably, each individual organism or cell contains only one or a limited number of library members. Advantageously, the nucleic acids are incorporated into expression vectors, in order to allow expression of the polypeptides encoded by the nucleic acids. In 20 a preferred aspect, therefore, a library may take the form of a population of host organisms, each organism containing one or more copies of an expression vector containing a single member of the library in nucleic acid form which can be expressed to produce its corresponding polypeptide member. Thus, the population of host organisms has the potential to encode a large repertoire of genetically diverse polypeptide variants.

25

30 A 'closed conformation multi-specific ligand' describes a multi-specific ligand as herein defined comprising at least two epitope binding domains as herein defined. The term 'closed conformation' (multi-specific ligand) means that the epitope binding domains of the ligand are arranged such that epitope binding by one epitope binding domain competes with epitope binding by another epitope binding domain. That is, cognate epitopes may be bound by each epitope binding domain individually but not simultaneously. The closed conformation of the ligand can be achieved using methods herein described.

5 **Antibody** An antibody (for example IgG, IgM, IgA, IgD or IgE) or fragment (such as a Fab , F(ab')₂, Fv, disulphide linked Fv, scFv, closed conformation multispecific antibody, disulphide-linked scFv, diabody) whether derived from any species naturally producing an antibody, or created by recombinant DNA technology; whether isolated from serum, B-cells, hybridomas, transfectomas, yeast or bacteria).

10 **Dual-specific ligand** A ligand comprising a first immunoglobulin single variable domain and a second immunoglobulin single variable domain as herein defined, wherein the variable regions are capable of binding to two different antigens or two epitopes on the same antigen which are not normally bound by a monospecific immunoglobulin. For example, the two epitopes may be on the same hapten, but are not the same epitope or sufficiently adjacent to be bound by a monospecific ligand. The dual specific ligands according to the invention are composed of variable domains which have different specificities, and do not contain mutually complementary variable domain pairs which 15 have the same specificity.

20 **Antigen** A molecule that is bound by a ligand according to the present invention. Typically, antigens are bound by antibody ligands and are capable of raising an antibody response *in vivo*. It may be a polypeptide, protein, nucleic acid or other molecule. Generally, the dual specific ligands according to the invention are selected for target specificity against a particular antigen. In the case of conventional antibodies and fragments thereof, the antibody binding site defined by the variable loops (L1, L2, L3 and H1, H2, H3) is capable of binding to the antigen.

25 **Epitope** A unit of structure conventionally bound by an immunoglobulin V_H/V_L pair. Epitopes define the minimum binding site for an antibody, and thus represent the target of specificity of an antibody. In the case of a single domain antibody, an epitope represents the unit of structure bound by a variable domain in isolation.

30 **Generic ligand** A ligand that binds to all members of a repertoire. Generally, not bound through the antigen binding site as defined above. Non-limiting examples include protein A, protein L and protein G.

5 **Selecting** Derived by screening, or derived by a Darwinian selection process, in which binding interactions are made between a domain and the antigen or epitope or between an antibody and an antigen or epitope. Thus a first variable domain may be selected for binding to an antigen or epitope in the presence or in the absence of a complementary variable domain.

10 **Universal framework** A single antibody framework sequence corresponding to the regions of an antibody conserved in sequence as defined by Kabat ("Sequences of Proteins of Immunological Interest", US Department of Health and Human Services) or corresponding to the human germline immunoglobulin repertoire or structure as defined by Chothia and Lesk, (1987) *J. Mol. Biol.* 196:910-917. The invention provides for the use of a single framework, or a set of such frameworks, which has been found to permit the derivation of virtually any binding specificity though variation in the hypervariable regions alone.

15 **Half-life** The time taken for the serum concentration of the ligand to reduce by 50%, *in vivo*, for example due to degradation of the ligand and/or clearance or sequestration of the ligand by natural mechanisms. The ligands of the invention are stabilised *in vivo* and their half-life increased by binding to molecules which resist degradation and/or clearance 20 or sequestration. Typically, such molecules are naturally occurring proteins which themselves have a long half-life *in vivo*. The half-life of a ligand is increased if its functional activity persists, *in vivo*, for a longer period than a similar ligand which is not specific for the half-life increasing molecule. Thus, a ligand specific for HSA and a target molecule is compared with the same ligand wherein the specificity for HSA is not 25 present, that it does not bind HSA but binds another molecule. For example, it may bind a second epitope on the target molecule. Typically, the half life is increased by 10%, 20%, 30%, 40%, 50% or more. Increases in the range of 2x, 3x, 4x, 5x, 10x, 20x, 30x, 40x, 50x or more of the half life are possible. Alternatively, or in addition, increases in 30 the range of up to 30x, 40x, 50x, 60x, 70x, 80x, 90x, 100x, 150x of the half life are possible.

Homogeneous immunoassay An immunoassay in which analyte is detected without need for a step of separating bound and un-bound reagents.

Substantially identical (or "substantially homologous") A first amino acid or nucleotide sequence that contains a sufficient number of identical or equivalent (e.g., with a similar side chain, e.g., conserved amino acid substitutions) amino acid residues or 5 nucleotides to a second amino acid or nucleotide sequence such that the first and second amino acid or nucleotide sequences have similar activities. In the case of antibodies, the second antibody has the same binding specificity and has at least 50% of the affinity of the same.

10 As used herein, the terms "low stringency," "medium stringency," "high stringency," or "very high stringency conditions" describe conditions for nucleic acid hybridization and washing. Guidance for performing hybridization reactions can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6, which is incorporated herein by reference in its entirety. Aqueous and nonaqueous methods are 15 described in that reference and either can be used. Specific hybridization conditions referred to herein are as follows: (1) low stringency hybridization conditions in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by two washes in 0.2X SSC, 0.1% SDS at least at 50°C (the temperature of the washes can be increased to 55°C for low stringency conditions); (2) medium stringency hybridization conditions in 6X 20 SSC at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 60°C; (3) high stringency hybridization conditions in 6X SSC at about 45°C, followed by one or more washes in 0.2X SSC, 0.1% SDS at 65°C; and preferably (4) very high stringency hybridization conditions are 0.5M sodium phosphate, 7% SDS at 65°C, followed by one 25 or more washes at 0.2X SSC, 1% SDS at 65°C. Very high stringency conditions (4) are the preferred conditions and the ones that should be used unless otherwise specified.

Detailed Description of the Invention

30 Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art (e.g., in cell culture, molecular genetics, nucleic acid chemistry, hybridisation techniques and biochemistry). Standard techniques are used for molecular, genetic and biochemical methods (see

generally, Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, 2d ed. (1989) Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. and Ausubel *et al.*, Short Protocols in Molecular Biology (1999) 4th Ed, John Wiley & Sons, Inc. which are incorporated herein by reference) and chemical methods.

5

Preparation of immunoglobulin based multi-specific ligands

Dual specific ligands according to the invention, whether open or closed in conformation according to the desired configuration of the invention, may be prepared according to 10 previously established techniques, used in the field of antibody engineering, for the preparation of scFv, "phage" antibodies and other engineered antibody molecules. Techniques for the preparation of antibodies, and in particular bispecific antibodies, are for example described in the following reviews and the references cited therein: Winter & Milstein, (1991) *Nature* 349:293-299; Plueckthun (1992) *Immunological Reviews* 130:151-188; Wright *et al.*, (1992) *Crit. Rev. Immunol.* 12:125-168; Holliger, P. & 15 Winter, G. (1993) *Curr. Op. Biotechnol.* 4, 446-449; Carter, *et al.* (1995) *J. Hematother.* 4, 463-470; Chester, K.A. & Hawkins, R.E. (1995) *Trends Biotechnol.* 13, 294-300; Hoogenboom, H.R. (1997) *Nature Biotechnol.* 15, 125-126; Fearon, D. (1997) *Nature Biotechnol.* 15, 618-619; Plückthun, A. & Pack, P. (1997) *Immunotechnology* 3, 83-105; 20 Carter, P. & Merchant, A.M. (1997) *Curr. Opin. Biotechnol.* 8, 449-454; Holliger, P. & Winter, G. (1997) *Cancer Immunol. Immunother.* 45,128-130.

The invention provides for the selection of variable domains against two different antigens or epitopes, and subsequent combination of the variable domains.

25

The techniques employed for selection of the variable domains employ libraries and selection procedures which are known in the art. Natural libraries (Marks *et al.* (1991) *J. Mol. Biol.*, 222: 581; Vaughan *et al.* (1996) *Nature Biotech.*, 14: 309) which use rearranged V genes harvested from human B cells are well known to those skilled in the art. Synthetic libraries (Hoogenboom & Winter (1992) *J. Mol. Biol.*, 227: 381; Barbas *et al.* (1992) *Proc. Natl. Acad. Sci. USA*, 89: 4457; Nissim *et al.* (1994) *EMBO J.*, 13: 692; Griffiths *et al.* (1994) *EMBO J.*, 13: 3245; De Kruif *et al.* (1995) *J. Mol. Biol.*, 248: 97) 30 are prepared by cloning immunoglobulin V genes, usually using PCR. Errors in the PCR

process can lead to a high degree of randomisation. V_H and/or V_L libraries may be selected against target antigens or epitopes separately, in which case single domain binding is directly selected for, or together.

5 A preferred method for making a dual specific ligand according to the present invention comprises using a selection system in which a repertoire of variable domains is selected for binding to a first antigen or epitope and a repertoire of variable domains is selected for binding to a second antigen or epitope. The selected variable first and second variable domains are then combined and the dual-specific ligand selected for binding to both first
10 and second antigen or epitope. Closed conformation ligands are selected for binding both first and second antigen or epitope in isolation but not simultaneously.

A. Library vector systems

15 A variety of selection systems are known in the art which are suitable for use in the present invention. Examples of such systems are described below.

Bacteriophage lambda expression systems may be screened directly as bacteriophage plaques or as colonies of lysogens, both as previously described (Huse *et al.* (1989) 20 *Science*, **246**: 1275; Caton and Koprowski (1990) *Proc. Natl. Acad. Sci. U.S.A.*, **87**; Mullinax *et al.* (1990) *Proc. Natl. Acad. Sci. U.S.A.*, **87**: 8095; Persson *et al.* (1991) *Proc. Natl. Acad. Sci. U.S.A.*, **88**: 2432) and are of use in the invention. Whilst such expression systems can be used to screen up to 10^6 different members of a library, they are not really suited to screening of larger numbers (greater than 10^6 members).

25 Of particular use in the construction of libraries are selection display systems, which enable a nucleic acid to be linked to the polypeptide it expresses. As used herein, a selection display system is a system that permits the selection, by suitable display means, of the individual members of the library by binding the generic and/or target ligands.

30 Selection protocols for isolating desired members of large libraries are known in the art, as typified by phage display techniques. Such systems, in which diverse peptide sequences are displayed on the surface of filamentous bacteriophage (Scott and Smith

(1990) *Science*, 249: 386), have proven useful for creating libraries of antibody fragments (and the nucleotide sequences that encode them) for the *in vitro* selection and amplification of specific antibody fragments that bind a target antigen (McCafferty *et al.*, WO 92/01047). The nucleotide sequences encoding the V_H and V_L regions are linked to gene fragments which encode leader signals that direct them to the periplasmic space of *E. coli* and as a result the resultant antibody fragments are displayed on the surface of the bacteriophage, typically as fusions to bacteriophage coat proteins (e.g., pIII or pVIII). Alternatively, antibody fragments are displayed externally on lambda phage capsids (phagebodies). An advantage of phage-based display systems is that, because they are biological systems, selected library members can be amplified simply by growing the phage containing the selected library member in bacterial cells. Furthermore, since the nucleotide sequence that encode the polypeptide library member is contained on a phage or phagemid vector, sequencing, expression and subsequent genetic manipulation is relatively straightforward.

15 Methods for the construction of bacteriophage antibody display libraries and lambda phage expression libraries are well known in the art (McCafferty *et al.* (1990) *Nature*, 348: 552; Kang *et al.* (1991) *Proc. Natl. Acad. Sci. U.S.A.*, 88: 4363; Clackson *et al.* (1991) *Nature*, 352: 624; Lowman *et al.* (1991) *Biochemistry*, 30: 10832; Burton *et al.* (1991) *Proc. Natl. Acad. Sci U.S.A.*, 88: 10134; Hoogenboom *et al.* (1991) *Nucleic Acids Res.*, 19: 4133; Chang *et al.* (1991) *J. Immunol.*, 147: 3610; Breitling *et al.* (1991) *Gene*, 104: 147; Marks *et al.* (1991) *supra*; Barbas *et al.* (1992) *supra*; Hawkins and Winter (1992) *J. Immunol.*, 22: 867; Marks *et al.*, 1992, *J. Biol. Chem.*, 267: 16007; Lerner *et al.* (1992) *Science*, 258: 1313, incorporated herein by reference).

25 One particularly advantageous approach has been the use of scFv phage-libraries (Huston *et al.*, 1988, *Proc. Natl. Acad. Sci U.S.A.*, 85: 5879-5883; Chaudhary *et al.* (1990) *Proc. Natl. Acad. Sci U.S.A.*, 87: 1066-1070; McCafferty *et al.* (1990) *supra*; Clackson *et al.* (1991) *Nature*, 352: 624; Marks *et al.* (1991) *J. Mol. Biol.*, 222: 581; Chiswell *et al.* (1992) *Trends Biotech.*, 10: 80; Marks *et al.* (1992) *J. Biol. Chem.*, 267). Various embodiments of scFv libraries displayed on bacteriophage coat proteins have been described. Refinements of phage display approaches are also known, for example as

described in WO96/06213 and WO92/01047 (Medical Research Council *et al.*) and WO97/08320 (Morphosys), which are incorporated herein by reference.

Other systems for generating libraries of polypeptides involve the use of cell-free enzymatic machinery for the *in vitro* synthesis of the library members. In one method, 5 RNA molecules are selected by alternate rounds of selection against a target ligand and PCR amplification (Tuerk and Gold (1990) *Science*, 249: 505; Ellington and Szostak (1990) *Nature*, 346: 818). A similar technique may be used to identify DNA sequences which bind a predetermined human transcription factor (Thiesen and Bach (1990) *Nucleic 10 Acids Res.*, 18: 3203; Beaudry and Joyce (1992) *Science*, 257: 635; WO92/05258 and WO92/14843). In a similar way, *in vitro* translation can be used to synthesise polypeptides as a method for generating large libraries. These methods which generally comprise stabilised polysome complexes, are described further in WO88/08453, 15 WO90/05785, WO90/07003, WO91/02076, WO91/05058, and WO92/02536. Alternative display systems which are not phage-based, such as those disclosed in WO95/22625 and WO95/11922 (Affymax) use the polysomes to display polypeptides for selection.

A still further category of techniques involves the selection of repertoires in artificial compartments, which allow the linkage of a gene with its gene product. For example, a 20 selection system in which nucleic acids encoding desirable gene products may be selected in microcapsules formed by water-in-oil emulsions is described in WO99/02671, WO00/40712 and Tawfik & Griffiths (1998) *Nature Biotechnol* 16(7), 652-6. Genetic elements encoding a gene product having a desired activity are compartmentalised into microcapsules and then transcribed and/or translated to produce their respective gene 25 products (RNA or protein) within the microcapsules. Genetic elements which produce gene product having desired activity are subsequently sorted. This approach selects gene products of interest by detecting the desired activity by a variety of means.

30 B. Library Construction.

Libraries intended for selection, may be constructed using techniques known in the art, for example as set forth above, or may be purchased from commercial sources. Libraries

which are useful in the present invention are described, for example, in WO99/20749. Once a vector system is chosen and one or more nucleic acid sequences encoding polypeptides of interest are cloned into the library vector, one may generate diversity within the cloned molecules by undertaking mutagenesis prior to expression; 5 alternatively, the encoded proteins may be expressed and selected, as described above, before mutagenesis and additional rounds of selection are performed. Mutagenesis of nucleic acid sequences encoding structurally optimised polypeptides is carried out by standard molecular methods. Of particular use is the polymerase chain reaction, or PCR, (Mullis and Faloona (1987) *Methods Enzymol.*, 155: 335, herein incorporated by 10 reference). PCR, which uses multiple cycles of DNA replication catalysed by a thermostable, DNA-dependent DNA polymerase to amplify the target sequence of interest, is well known in the art. The construction of various antibody libraries has been discussed in Winter *et al.* (1994) *Ann. Rev. Immunology* 12, 433-55, and references cited therein.

15

PCR is performed using template DNA (at least 1fg; more usefully, 1-1000 ng) and at least 25 pmol of oligonucleotide primers; it may be advantageous to use a larger amount of primer when the primer pool is heavily heterogeneous, as each sequence is represented by only a small fraction of the molecules of the pool, and amounts become limiting in the 20 later amplification cycles. A typical reaction mixture includes: 2 μ l of DNA, 25 pmol of oligonucleotide primer, 2.5 μ l of 10X PCR buffer 1 (Perkin-Elmer, Foster City, CA), 0.4 μ l of 1.25 μ M dNTP, 0.15 μ l (or 2.5 units) of Taq DNA polymerase (Perkin Elmer, Foster City, CA) and deionized water to a total volume of 25 μ l. Mineral oil is overlaid and the PCR is performed using a programmable thermal cycler. The length and 25 temperature of each step of a PCR cycle, as well as the number of cycles, is adjusted in accordance to the stringency requirements in effect. Annealing temperature and timing are determined both by the efficiency with which a primer is expected to anneal to a template and the degree of mismatch that is to be tolerated; obviously, when nucleic acid molecules are simultaneously amplified and mutagenised, mismatch is required, at least 30 in the first round of synthesis. The ability to optimise the stringency of primer annealing conditions is well within the knowledge of one of moderate skill in the art. An annealing temperature of between 30 °C and 72 °C is used. Initial denaturation of the template molecules normally occurs at between 92°C and 99°C for 4 minutes, followed by 20-40

cycles consisting of denaturation (94-99°C for 15 seconds to 1 minute), annealing (temperature determined as discussed above; 1-2 minutes), and extension (72°C for 1-5 minutes, depending on the length of the amplified product). Final extension is generally for 4 minutes at 72°C, and may be followed by an indefinite (0-24 hour) step at 4°C.

5

C. Combining single variable domains

Domains useful in the invention, once selected, may be combined by a variety of methods known in the art, including covalent and non-covalent methods.

10

Preferred methods include the use of polypeptide linkers, as described, for example, in connection with scFv molecules (Bird *et al.*, (1988) *Science* 242:423-426). Discussion of suitable linkers is provided in *Bird et al.* *Science* 242, 423-426; *Hudson et al.*, *Journal Immunol Methods* 231 (1999) 177-189; *Hudson et al.*, *Proc Nat Acad Sci USA* 85, 5879-5883. Linkers are preferably flexible, allowing the two single domains to interact. One linker example is a (Gly₄ Ser)_n linker, where n=1 to 8, eg, 2, 3, 4, 5 or 7. The linkers used in diabodies, which are less flexible, may also be employed (Holliger *et al.*, (1993) *PNAS (USA)* 90:6444-6448).

20

In one embodiment, the linker employed is not an immunoglobulin hinge region.

25

Variable domains may be combined using methods other than linkers. For example, the use of disulphide bridges, provided through naturally-occurring or engineered cysteine residues, may be exploited to stabilise V_H-V_H,V_L-V_L or V_H-V_L dimers (Reiter *et al.*, (1994) *Protein Eng.* 7:697-704) or by remodelling the interface between the variable domains to improve the "fit" and thus the stability of interaction (Ridgeway *et al.*, (1996) *Protein Eng.* 7:617-621; Zhu *et al.*, (1997) *Protein Science* 6:781-788).

30

Other techniques for joining or stabilising variable domains of immunoglobulins, and in particular antibody V_H domains, may be employed as appropriate.

In accordance with the present invention, dual specific ligands can be in "closed" conformations in solution. A "closed" configuration is that in which the two domains (for

example V_H and V_L) are present in associated form, such as that of an associated V_H - V_L pair which forms an antibody binding site. For example, scFv may be in a closed conformation, depending on the arrangement of the linker used to link the V_H and V_L domains. If this is sufficiently flexible to allow the domains to associate, or rigidly holds 5 them in the associated position, it is likely that the domains will adopt a closed conformation.

Similarly, V_H domain pairs and V_L domain pairs may exist in a closed conformation. Generally, this will be a function of close association of the domains, such as by a rigid 10 linker, in the ligand molecule. Ligands in a closed conformation will be unable to bind both the molecule which increases the half-life of the ligand and a second target molecule. Thus, the ligand will typically only bind the second target molecule on dissociation from the molecule which increases the half-life of the ligand.

15 Moreover, the construction of V_H/V_H , V_L/V_L or V_H/V_L dimers without linkers provides for competition between the domains.

Ligands according to the invention may moreover be in an open conformation. In such a conformation, the ligands will be able to simultaneously bind both the molecule which 20 increases the half-life of the ligand and the second target molecule. Typically, variable domains in an open configuration are (in the case of V_H - V_L pairs) held far enough apart for the domains not to interact and form an antibody binding site and not to compete for binding to their respective epitopes. In the case of V_H/V_H or V_L/V_L dimers, the domains are not forced together by rigid linkers. Naturally, such domain pairings will not compete 25 for antigen binding or form an antibody binding site.

Fab fragments and whole antibodies will exist primarily in the closed conformation, although it will be appreciated that open and closed dual specific ligands are likely to exist in a variety of equilibria under different circumstances. Binding of the ligand to a 30 target is likely to shift the balance of the equilibrium towards the open configuration. Thus, certain ligands according to the invention can exist in two conformations in solution, one of which (the open form) can bind two antigens or epitopes independently,

whilst the alternative conformation (the closed form) can only bind one antigen or epitope; antigens or epitopes thus compete for binding to the ligand in this conformation.

Although the open form of the dual specific ligand may thus exist in equilibrium with the closed form in solution, it is envisaged that the equilibrium will favour the closed form; moreover, the open form can be sequestered by target binding into a closed conformation. Preferably, therefore, certain dual specific ligands of the invention are present in an equilibrium between two (open and closed) conformations.

10 Dual specific ligands according to the invention may be modified in order to favour an open or closed conformation. For example, stabilisation of V_H - V_L interactions with disulphide bonds stabilises the closed conformation. Moreover, linkers used to join the domains, including V_H domain and V_L domain pairs, may be constructed such that the open form is favoured; for example, the linkers may sterically hinder the association of 15 the domains, such as by incorporation of large amino acid residues in opportune locations, or the designing of a suitable rigid structure which will keep the domains physically spaced apart.

20 **D. Characterisation of the dual-specific ligand.**

The binding of the dual-specific ligand to its specific antigens or epitopes can be tested by methods which will be familiar to those skilled in the art and include ELISA. In a preferred embodiment of the invention binding is tested using monoclonal phage ELISA.

25 Phage ELISA may be performed according to any suitable procedure: an exemplary protocol is set forth below.

Populations of phage produced at each round of selection can be screened for binding by 30 ELISA to the selected antigen or epitope, to identify "polyclonal" phage antibodies. Phage from single infected bacterial colonies from these populations can then be screened by ELISA to identify "monoclonal" phage antibodies. It is also desirable to screen soluble antibody fragments for binding to antigen or epitope, and this can also be undertaken by

ELISA using reagents, for example, against a C- or N-terminal tag (see for example Winter *et al.* (1994) Ann. Rev. Immunology 12, 433-55 and references cited therein.

5 The diversity of the selected phage monoclonal antibodies may also be assessed by gel electrophoresis of PCR products (Marks *et al.* 1991, *supra*; Nissim *et al.* 1994 *supra*), probing (Tomlinson *et al.*, 1992) J. Mol. Biol. 227, 776) or by sequencing of the vector DNA.

10 E. Structure of 'Dual-specific ligands'.

15 As described above, an antibody is herein defined as an antibody (for example IgG, IgM, IgA, IgA, IgE) or fragment (Fab, Fv, disulphide linked Fv, scFv, diabody) which comprises at least one heavy and a light chain variable domain, at least two heavy chain variable domains or at least two light chain variable domains. It may be at least partly derived from any species naturally producing an antibody, or created by recombinant DNA technology; whether isolated from serum, B-cells, hybridomas, transfectomas, yeast or bacteria).

20 In a preferred embodiment of the invention the dual-specific ligand comprises at least one single heavy chain variable domain of an antibody and one single light chain variable domain of an antibody, or two single heavy or light chain variable domains. For example, the ligand may comprise a V_H/V_L pair, a pair of V_H domains or a pair of V_L domains.

25 The first and the second variable domains of such a ligand may be on the same polypeptide chain. Alternatively they may be on separate polypeptide chains. In the case that they are on the same polypeptide chain they may be linked by a linker, which is preferentially a peptide sequence, as described above.

30 The first and second variable domains may be covalently or non-covalently associated. In the case that they are covalently associated, the covalent bonds may be disulphide bonds.

In the case that the variable domains are selected from V-gene repertoires selected for instance using phage display technology as herein described, then these variable domains

comprise a universal framework region, such that they may be recognised by a specific generic ligand as herein defined. The use of universal frameworks, generic ligands and the like is described in WO99/20749.

5 Where V-gene repertoires are used variation in polypeptide sequence is preferably located within the structural loops of the variable domains. The polypeptide sequences of either variable domain may be altered by DNA shuffling or by mutation in order to enhance the interaction of each variable domain with its complementary pair. DNA shuffling is known in the art and taught, for example, by Stemmer, 1994, *Nature* 370: 389-391 and

10 U.S. Patent No. 6,297,053, both of which are incorporated herein by reference. Other methods of mutagenesis are well known to those of skill in the art.

15 In a preferred embodiment of the invention the 'dual-specific ligand' is a single chain Fv fragment. In an alternative embodiment of the invention, the 'dual-specific ligand' consists of a Fab format.

In a further aspect, the present invention provides nucleic acid encoding at least a 'dual-specific ligand' as herein defined.

20 One skilled in the art will appreciate that, depending on the aspect of the invention, both antigens or epitopes may bind simultaneously to the same antibody molecule. Alternatively, they may compete for binding to the same antibody molecule. For example, where both epitopes are bound simultaneously, both variable domains of a dual specific ligand are able to independently bind their target epitopes. Where the domains compete, the one variable domain is capable of binding its target, but not at the same time as the other variable domain binds its cognate target; or the first variable domain is capable of binding its target, but not at the same time as the second variable domain binds its cognate target.

25

30 The variable regions may be derived from antibodies directed against target antigens or epitopes. Alternatively they may be derived from a repertoire of single antibody domains such as those expressed on the surface of filamentous bacteriophage. Selection may be performed as described below.

In general, the nucleic acid molecules and vector constructs required for the performance of the present invention may be constructed and manipulated as set forth in standard laboratory manuals, such as Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor, USA.

The manipulation of nucleic acids useful in the present invention is typically carried out in recombinant vectors.

10 Thus in a further aspect, the present invention provides a vector comprising nucleic acid encoding at least a 'dual-specific ligand' as herein defined.

As used herein, vector refers to a discrete element that is used to introduce heterologous DNA into cells for the expression and/or replication thereof. Methods by which to select 15 or construct and, subsequently, use such vectors are well known to one of ordinary skill in the art. Numerous vectors are publicly available, including bacterial plasmids, bacteriophage, artificial chromosomes and episomal vectors. Such vectors may be used for simple cloning and mutagenesis; alternatively gene expression vector is employed. A vector of use according to the invention may be selected to accommodate a polypeptide 20 coding sequence of a desired size, typically from 0.25 kilobase (kb) to 40 kb or more in length. A suitable host cell is transformed with the vector after *in vitro* cloning manipulations. Each vector contains various functional components, which generally include a cloning (or "polylinker") site, an origin of replication and at least one selectable marker gene. If given vector is an expression vector, it additionally possesses one or more 25 of the following: enhancer element, promoter, transcription termination and signal sequences, each positioned in the vicinity of the cloning site, such that they are operatively linked to the gene encoding a ligand according to the invention.

Both cloning and expression vectors generally contain nucleic acid sequences that enable 30 the vector to replicate in one or more selected host cells. Typically in cloning vectors, this sequence is one that enables the vector to replicate independently of the host chromosomal DNA and includes origins of replication or autonomously replicating sequences. Such sequences are well known for a variety of bacteria, yeast and viruses.

The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2 micron plasmid origin is suitable for yeast, and various viral origins (e.g. SV 40, adenovirus) are useful for cloning vectors in mammalian cells. Generally, the origin of replication is not needed for mammalian expression vectors unless these are used in mammalian cells able to replicate high levels of DNA, such as COS cells.

Advantageously, a cloning or expression vector may contain a selection gene also referred to as selectable marker. This gene encodes a protein necessary for the survival or growth of transformed host cells grown in a selective culture medium. Host cells not transformed with the vector containing the selection gene will therefore not survive in the culture medium. Typical selection genes encode proteins that confer resistance to antibiotics and other toxins, e.g. ampicillin, neomycin, methotrexate or tetracycline, complement auxotrophic deficiencies, or supply critical nutrients not available in the growth media.

15

Since the replication of vectors encoding a ligand according to the present invention is most conveniently performed in *E. coli*, an *E. coli*-selectable marker, for example, the β -lactamase gene that confers resistance to the antibiotic ampicillin, is of use. These can be obtained from *E. coli* plasmids, such as pBR322 or a pUC plasmid such as pUC18 or pUC19.

20 Expression vectors usually contain a promoter that is recognised by the host organism and is operably linked to the coding sequence of interest. Such a promoter may be inducible or constitutive. The term "operably linked" refers to a juxtaposition wherein the components described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences.

25 30 Promoters suitable for use with prokaryotic hosts include, for example, the β -lactamase and lactose promoter systems, alkaline phosphatase, the tryptophan (trp) promoter system and hybrid promoters such as the tac promoter. Promoters for use in bacterial systems

will also generally contain a Shine-Delgarno sequence operably linked to the coding sequence.

The preferred vectors are expression vectors that enables the expression of a nucleotide sequence corresponding to a polypeptide library member. Thus, selection with the first and/or second antigen or epitope can be performed by separate propagation and expression of a single clone expressing the polypeptide library member or by use of any selection display system. As described above, the preferred selection display system is bacteriophage display. Thus, phage or phagemid vectors may be used, eg pIT1 or pIT2. Leader sequences useful in the invention include pelB, stII, ompA, phoA, bla and pelA. One example are phagemid vectors which have an *E. coli* origin of replication (for double stranded replication) and also a phage origin of replication (for production of single-stranded DNA). The manipulation and expression of such vectors is well known in the art (Hoogenboom and Winter (1992) *supra*; Nissim *et al.* (1994) *supra*). Briefly, the vector contains a β -lactamase gene to confer selectivity on the phagemid and a lac promoter upstream of a expression cassette that consists (N to C terminal) of a pelB leader sequence (which directs the expressed polypeptide to the periplasmic space), a multiple cloning site (for cloning the nucleotide version of the library member), optionally, one or more peptide tag (for detection), optionally, one or more TAG stop codon and the phage protein pIII. Thus, using various suppressor and non-suppressor strains of *E. coli* and with the addition of glucose, iso-propyl thio- β -D-galactoside (IPTG) or a helper phage, such as VCS M13, the vector is able to replicate as a plasmid with no expression, produce large quantities of the polypeptide library member only or produce phage, some of which contain at least one copy of the polypeptide-pIII fusion on their surface.

Construction of vectors encoding ligands according to the invention employs conventional ligation techniques. Isolated vectors or DNA fragments are cleaved, tailored, and religated in the form desired to generate the required vector. If desired, analysis to confirm that the correct sequences are present in the constructed vector can be performed in a known fashion. Suitable methods for constructing expression vectors, preparing *in vitro* transcripts, introducing DNA into host cells, and performing analyses for assessing expression and function are known to those skilled in the art. The presence

of a gene sequence in a sample is detected, or its amplification and/or expression quantified by conventional methods, such as Southern or Northern analysis, Western blotting, dot blotting of DNA, RNA or protein, *in situ* hybridisation, immunocytochemistry or sequence analysis of nucleic acid or protein molecules. Those 5 skilled in the art will readily envisage how these methods may be modified, if desired.

Structure of closed conformation multispecific ligands

According to one aspect of the second configuration of the invention present invention, 10 the two or more non-complementary epitope binding domains are linked so that they are in a closed conformation as herein defined. Advantageously, they may be further attached to a skeleton which may, as a alternative, or on addition to a linker described herein, facilitate the formation and/or maintenance of the closed conformation of the epitope binding sites with respect to one another.

15

(I) Skeletons

Skeletons may be based on immunoglobulin molecules or may be non-immunoglobulin in origin as set forth above. Preferred immunoglobulin skeletons as herein defined includes any one or more of those selected from the following: an immunoglobulin molecule 20 comprising at least (i) the CL (kappa or lambda subclass) domain of an antibody; or (ii) the CH1 domain of an antibody heavy chain; an immunoglobulin molecule comprising the CH1 and CH2 domains of an antibody heavy chain; an immunoglobulin molecule comprising the CH1, CH2 and CH3 domains of an antibody heavy chain; or any of the subset (ii) in conjunction with the CL (kappa or lambda subclass) domain of an antibody. 25 A hinge region domain may also be included.. Such combinations of domains may, for example, mimic natural antibodies, such as IgG or IgM, or fragments thereof, such as Fv, scFv, Fab or F(ab')₂ molecules. Those skilled in the art will be aware that this list is not intended to be exhaustive.

30 **(II) Protein scaffolds**

Each epitope binding domain comprises a protein scaffold and one or more CDRs which are involved in the specific interaction of the domain with one or more epitopes. Advantageously, an epitope binding domain according to the present invention comprises

three CDRs. Suitable protein scaffolds include any of those selected from the group consisting of the following: those based on immunoglobulin domains, those based on fibronectin, those based on affibodies, those based on CTLA4, those based on chaperones such as GroEL, those based on lipocalin and those based on the bacterial Fc receptors 5 SpA and SpD. Those skilled in the art will appreciate that this list is not intended to be exhaustive.

F: Scaffolds for use in Constructing Dual Specific Ligands

10 10 i. Selection of the main-chain conformation

The members of the immunoglobulin superfamily all share a similar fold for their polypeptide chain. For example, although antibodies are highly diverse in terms of their primary sequence, comparison of sequences and crystallographic structures has revealed that, contrary to expectation, five of the six antigen binding loops of antibodies (H1, H2, 15 L1, L2, L3) adopt a limited number of main-chain conformations, or canonical structures (Chothia and Lesk (1987) *J. Mol. Biol.*, 196: 901; Chothia *et al.* (1989) *Nature*, 342: 877). Analysis of loop lengths and key residues has therefore enabled prediction of the main-chain conformations of H1, H2, L1, L2 and L3 found in the majority of human antibodies (Chothia *et al.* (1992) *J. Mol. Biol.*, 227: 799; Tomlinson *et al.* (1995) *EMBO J.*, 14: 20 4628; Williams *et al.* (1996) *J. Mol. Biol.*, 264: 220). Although the H3 region is much more diverse in terms of sequence, length and structure (due to the use of D segments), it also forms a limited number of main-chain conformations for short loop lengths which depend on the length and the presence of particular residues, or types of residue, at key positions in the loop and the antibody framework (Martin *et al.* (1996) *J. Mol. Biol.*, 263: 25 800; Shirai *et al.* (1996) *FEBS Letters*, 399: 1).

The dual specific ligands of the present invention are advantageously assembled from libraries of domains, such as libraries of V_H domains and/or libraries of V_L domains. Moreover, the dual specific ligands of the invention may themselves be provided in the 30 form of libraries. In one aspect of the present invention, libraries of dual specific ligands and/or domains are designed in which certain loop lengths and key residues have been chosen to ensure that the main-chain conformation of the members is known. Advantageously, these are real conformations of immunoglobulin superfamily molecules

found in nature, to minimise the chances that they are non-functional, as discussed above. Germline V gene segments serve as one suitable basic framework for constructing antibody or T-cell receptor libraries; other sequences are also of use. Variations may occur at a low frequency, such that a small number of functional members may possess an 5 altered main-chain conformation, which does not affect its function.

Canonical structure theory is also of use to assess the number of different main-chain conformations encoded by ligands, to predict the main-chain conformation based on ligand sequences and to chose residues for diversification which do not affect the 10 canonical structure. It is known that, in the human V_{κ} domain, the L1 loop can adopt one of four canonical structures, the L2 loop has a single canonical structure and that 90% of human V_{κ} domains adopt one of four or five canonical structures for the L3 loop (Tomlinson *et al.* (1995) *supra*); thus, in the V_{κ} domain alone, different canonical structures can combine to create a range of different main-chain conformations. Given 15 that the V_{λ} domain encodes a different range of canonical structures for the L1, L2 and L3 loops and that V_{κ} and V_{λ} domains can pair with any V_H domain which can encode several canonical structures for the H1 and H2 loops, the number of canonical structure combinations observed for these five loops is very large. This implies that the generation 20 of diversity in the main-chain conformation may be essential for the production of a wide range of binding specificities. However, by constructing an antibody library based on a single known main-chain conformation it has been found, contrary to expectation, that diversity in the main-chain conformation is not required to generate sufficient diversity to target substantially all antigens. Even more surprisingly, the single main-chain conformation need not be a consensus structure - a single naturally occurring 25 conformation can be used as the basis for an entire library. Thus, in a preferred aspect, the dual-specific ligands of the invention possess a single known main-chain conformation.

The single main-chain conformation that is chosen is preferably commonplace among 30 molecules of the immunoglobulin superfamily type in question. A conformation is commonplace when a significant number of naturally occurring molecules are observed to adopt it. Accordingly, in a preferred aspect of the invention, the natural occurrence of the different main-chain conformations for each binding loop of an immunoglobulin domain are considered separately and then a naturally occurring variable domain is

chosen which possesses the desired combination of main-chain conformations for the different loops. If none is available, the nearest equivalent may be chosen. It is preferable that the desired combination of main-chain conformations for the different loops is created by selecting germline gene segments which encode the desired main-chain conformations. It is more preferable, that the selected germline gene segments are frequently expressed in nature, and most preferable that they are the most frequently expressed of all natural germline gene segments.

In designing dual specific ligands or libraries thereof the incidence of the different main-chain conformations for each of the six antigen binding loops may be considered separately. For H1, H2, L1, L2 and L3, a given conformation that is adopted by between 20% and 100% of the antigen binding loops of naturally occurring molecules is chosen. Typically, its observed incidence is above 35% (i.e. between 35% and 100%) and, ideally, above 50% or even above 65%. Since the vast majority of H3 loops do not have canonical structures, it is preferable to select a main-chain conformation which is commonplace among those loops which do display canonical structures. For each of the loops, the conformation which is observed most often in the natural repertoire is therefore selected. In human antibodies, the most popular canonical structures (CS) for each loop are as follows: H1 - CS 1 (79% of the expressed repertoire), H2 - CS 3 (46%), L1 - CS 2 of V_κ (39%), L2 - CS 1 (100%), L3 - CS 1 of V_κ (36%) (calculation assumes a κ:λ ratio of 70:30, Hood *et al.* (1967) *Cold Spring Harbor Symp. Quant. Biol.*, **48**: 133). For H3 loops that have canonical structures, a CDR3 length (Kabat *et al.* (1991) *Sequences of proteins of immunological interest*, U.S. Department of Health and Human Services) of seven residues with a salt-bridge from residue 94 to residue 101 appears to be the most common. There are at least 16 human antibody sequences in the EMBL data library with the required H3 length and key residues to form this conformation and at least two crystallographic structures in the protein data bank which can be used as a basis for antibody modelling (2cgr and 1tet). The most frequently expressed germline gene segments that this combination of canonical structures are the V_H segment 3-23 (DP-47), the J_H segment JH4b, the V_κ segment O2/O12 (DPK9) and the J_κ segment J_κ1. V_H segments DP45 and DP38 are also suitable. These segments can therefore be used in combination as a basis to construct a library with the desired single main-chain conformation.

Alternatively, instead of choosing the single main-chain conformation based on the natural occurrence of the different main-chain conformations for each of the binding loops in isolation, the natural occurrence of combinations of main-chain conformations is used as the basis for choosing the single main-chain conformation. In the case of antibodies, for example, the natural occurrence of canonical structure combinations for any two, three, four, five or for all six of the antigen binding loops can be determined. Here, it is preferable that the chosen conformation is commonplace in naturally occurring antibodies and most preferable that it observed most frequently in the natural repertoire. Thus, in human antibodies, for example, when natural combinations of the five antigen binding loops, H1, H2, L1, L2 and L3, are considered, the most frequent combination of canonical structures is determined and then combined with the most popular conformation for the H3 loop, as a basis for choosing the single main-chain conformation.

15 ii. Diversification of the canonical sequence

Having selected several known main-chain conformations or, preferably a single known main-chain conformation, dual specific ligands according to the invention or libraries for use in the invention can be constructed by varying the binding site of the molecule in order to generate a repertoire with structural and/or functional diversity. This means that variants are generated such that they possess sufficient diversity in their structure and/or in their function so that they are capable of providing a range of activities.

25 The desired diversity is typically generated by varying the selected molecule at one or more positions. The positions to be changed can be chosen at random or are preferably selected. The variation can then be achieved either by randomisation, during which the resident amino acid is replaced by any amino acid or analogue thereof, natural or synthetic, producing a very large number of variants or by replacing the resident amino acid with one or more of a defined subset of amino acids, producing a more limited 30 number of variants.

Various methods have been reported for introducing such diversity. Error-prone PCR (Hawkins *et al.* (1992) *J. Mol. Biol.*, 226: 889), chemical mutagenesis (Deng *et al.* (1994)

J. Biol. Chem., 269: 9533) or bacterial mutator strains (Low *et al.* (1996) *J. Mol. Biol.*, 260: 359) can be used to introduce random mutations into the genes that encode the molecule. Methods for mutating selected positions are also well known in the art and include the use of mismatched oligonucleotides or degenerate oligonucleotides, with or 5 without the use of PCR. For example, several synthetic antibody libraries have been created by targeting mutations to the antigen binding loops. The H3 region of a human tetanus toxoid-binding Fab has been randomised to create a range of new binding specificities (Barbas *et al.* (1992) *Proc. Natl. Acad. Sci. USA*, 89: 4457). Random or semi-random H3 and L3 regions have been appended to germline V gene segments to 10 produce large libraries with unmutated framework regions (Hoogenboom & Winter (1992) *J. Mol. Biol.*, 227: 381; Barbas *et al.* (1992) *Proc. Natl. Acad. Sci. USA*, 89: 4457; Nissim *et al.* (1994) *EMBO J.*, 13: 692; Griffiths *et al.* (1994) *EMBO J.*, 13: 3245; De Kruif *et al.* (1995) *J. Mol. Biol.*, 248: 97). Such diversification has been extended to include some or all of the other antigen binding loops (Crameri *et al.* (1996) *Nature Med.*, 15 2: 100; Riechmann *et al.* (1995) *Bio/Technology*, 13: 475; Morphosys, WO97/08320, supra).

Since loop randomisation has the potential to create approximately more than 10^{15} structures for H3 alone and a similarly large number of variants for the other five loops, it 20 is not feasible using current transformation technology or even by using cell free systems to produce a library representing all possible combinations. For example, in one of the largest libraries constructed to date, 6×10^{10} different antibodies, which is only a fraction of the potential diversity for a library of this design, were generated (Griffiths *et al.* (1994) supra).

25

In a preferred embodiment, only those residues which are directly involved in creating or modifying the desired function of the molecule are diversified. For many molecules, the function will be to bind a target and therefore diversity should be concentrated in the target binding site, while avoiding changing residues which are crucial to the overall 30 packing of the molecule or to maintaining the chosen main-chain conformation.

Diversification of the canonical sequence as it applies to antibody domains

In the case of antibody dual-specific ligands, the binding site for the target is most often the antigen binding site. Thus, in a highly preferred aspect, the invention provides libraries of or for the assembly of antibody dual-specific ligands in which only those residues in the antigen binding site are varied. These residues are extremely diverse in the human antibody repertoire and are known to make contacts in high-resolution antibody/antigen complexes. For example, in L2 it is known that positions 50 and 53 are diverse in naturally occurring antibodies and are observed to make contact with the antigen. In contrast, the conventional approach would have been to diversify all the residues in the corresponding Complementarity Determining Region (CDR1) as defined by Kabat *et al.* (1991, *supra*), some seven residues compared to the two diversified in the library for use according to the invention. This represents a significant improvement in terms of the functional diversity required to create a range of antigen binding specificities.

15 In nature, antibody diversity is the result of two processes: somatic recombination of germline V, D and J gene segments to create a naive primary repertoire (so called germline and junctional diversity) and somatic hypermutation of the resulting rearranged V genes. Analysis of human antibody sequences has shown that diversity in the primary repertoire is focused at the centre of the antigen binding site whereas somatic hypermutation spreads diversity to regions at the periphery of the antigen binding site that are highly conserved in the primary repertoire (see Tomlinson *et al.* (1996) *J. Mol. Biol.*, 20 256: 813). This complementarity has probably evolved as an efficient strategy for searching sequence space and, although apparently unique to antibodies, it can easily be applied to other polypeptide repertoires. The residues which are varied are a subset of those that form the binding site for the target. Different (including overlapping) subsets of residues in the target binding site are diversified at different stages during selection, if desired.

30 In the case of an antibody repertoire, an initial 'naive' repertoire is created where some, but not all, of the residues in the antigen binding site are diversified. As used herein in this context, the term "naive" refers to antibody molecules that have no pre-determined target. These molecules resemble those which are encoded by the immunoglobulin genes of an individual who has not undergone immune diversification, as is the case with fetal

and newborn individuals, whose immune systems have not yet been challenged by a wide variety of antigenic stimuli. This repertoire is then selected against a range of antigens or epitopes. If required, further diversity can then be introduced outside the region diversified in the initial repertoire. This matured repertoire can be selected for 5 modified function, specificity or affinity.

The invention provides two different naive repertoires of binding domains for the construction of dual specific ligands, or a naïve library of dual specific ligands, in which some or all of the residues in the antigen binding site are varied. The "primary" library 10 mimics the natural primary repertoire, with diversity restricted to residues at the centre of the antigen binding site that are diverse in the germline V gene segments (germline diversity) or diversified during the recombination process (junctional diversity). Those residues which are diversified include, but are not limited to, H50, H52, H52a, H53, H55, H56, H58, H95, H96, H97, H98, L50, L53, L91, L92, L93, L94 and L96. In the "somatic" 15 library, diversity is restricted to residues that are diversified during the recombination process (junctional diversity) or are highly somatically mutated). Those residues which are diversified include, but are not limited to: H31, H33, H35, H95, H96, H97, H98, L30, L31, L32, L34 and L96. All the residues listed above as suitable for diversification in these libraries are known to make contacts in one or more antibody-antigen complexes. 20 Since in both libraries, not all of the residues in the antigen binding site are varied, additional diversity is incorporated during selection by varying the remaining residues, if it is desired to do so. It shall be apparent to one skilled in the art that any subset of any of these residues (or additional residues which comprise the antigen binding site) can be used for the initial and/or subsequent diversification of the antigen binding site.

25 In the construction of libraries for use in the invention, diversification of chosen positions is typically achieved at the nucleic acid level, by altering the coding sequence which specifies the sequence of the polypeptide such that a number of possible amino acids (all 20 or a subset thereof) can be incorporated at that position. Using the IUPAC nomenclature, the most versatile codon is NNK, which encodes all amino acids as well as 30 the TAG stop codon. The NNK codon is preferably used in order to introduce the required diversity. Other codons which achieve the same ends are also of use, including

the NNN codon, which leads to the production of the additional stop codons TGA and TAA.

A feature of side-chain diversity in the antigen binding site of human antibodies is a 5 pronounced bias which favours certain amino acid residues. If the amino acid composition of the ten most diverse positions in each of the V_H , V_{κ} and V_{λ} regions are summed, more than 76% of the side-chain diversity comes from only seven different residues, these being, serine (24%), tyrosine (14%), asparagine (11%), glycine (9%), alanine (7%), aspartate (6%) and threonine (6%). This bias towards hydrophilic residues 10 and small residues which can provide main-chain flexibility probably reflects the evolution of surfaces which are predisposed to binding a wide range of antigens or epitopes and may help to explain the required promiscuity of antibodies in the primary repertoire.

15 Since it is preferable to mimic this distribution of amino acids, the distribution of amino acids at the positions to be varied preferably mimics that seen in the antigen binding site of antibodies. Such bias in the substitution of amino acids that permits selection of certain polypeptides (not just antibody polypeptides) against a range of target antigens is easily applied to any polypeptide repertoire. There are various methods for biasing the amino 20 acid distribution at the position to be varied (including the use of tri-nucleotide mutagenesis, see WO97/08320), of which the preferred method, due to ease of synthesis, is the use of conventional degenerate codons. By comparing the amino acid profile encoded by all combinations of degenerate codons (with single, double, triple and quadruple degeneracy in equal ratios at each position) with the natural amino acid use it is 25 possible to calculate the most representative codon. The codons (AGT)(AGC)T, (AGT)(AGC)C and (AGT)(AGC)(CT) - that is, DVT, DVC and DVY, respectively using IUPAC nomenclature - are those closest to the desired amino acid profile: they encode 22% serine and 11% tyrosine, asparagine, glycine, alanine, aspartate, threonine and cysteine. Preferably, therefore, libraries are constructed using either the DVT, DVC or 30 DVY codon at each of the diversified positions.

G: Antigens capable of increasing ligand half-life

The dual specific ligands according to the invention, in one configuration thereof, are capable of binding to one or more molecules which can increase the half-life of the ligand 5 *in vivo*. Typically, such molecules are polypeptides which occur naturally *in vivo* and which resist degradation or removal by endogenous mechanisms which remove unwanted material from the organism. For example, the molecule which increases the half-life of the organism may be selected from the following:

10 Proteins from the extracellular matrix; for example collagen, laminins, integrins and fibronectin. Collagens are the major proteins of the extracellular matrix. About 15 types of collagen molecules are currently known, found in different parts of the body, eg type I collagen (accounting for 90% of body collagen) found in bone, skin, tendon, ligaments, cornea, internal organs or type II collagen found in cartilage, invertebral disc, notochord, 15 vitreous humour of the eye.

Proteins found in blood, including:

20 Plasma proteins such as fibrin, α -2 macroglobulin, serum albumin, fibrinogen A, fibrinogen B, serum amyloid protein A, heptaglobin, profilin, ubiquitin, uteroglobin and β -2-microglobulin;

Enzymes and inhibitors such as plasminogen, lysozyme, cystatin C, alpha-1-antitrypsin and pancreatic trypsin inhibitor. Plasminogen is the inactive precursor of the trypsin-like 25 serine protease plasmin. It is normally found circulating through the blood stream. When plasminogen becomes activated and is converted to plasmin, it unfolds a potent enzymatic domain that dissolves the fibrinogen fibers that entangle the blood cells in a blood clot. This is called fibrinolysis.

30 Immune system proteins, such as IgE, IgG, IgM.

Transport proteins such as retinol binding protein, α -1 microglobulin.

Defensins such as beta-defensin 1, Neutrophil defensins 1,2 and 3.

Proteins found at the blood brain barrier or in neural tissues, such as melanocortin receptor, myelin, ascorbate transporter.

5

Transferrin receptor specific ligand-neuropharmaceutical agent fusion proteins (see US5977307);

brain capillary endothelial cell receptor, transferrin, transferrin receptor, insulin, insulin-

10 like growth factor 1 (IGF 1) receptor, insulin-like growth factor 2 (IGF 2) receptor, insulin receptor.

Proteins localised to the kidney, such as polycystin, type IV collagen, organic anion transporter K1, Heymann's antigen.

15

Proteins localised to the liver, for example alcohol dehydrogenase, G250.

Blood coagulation factor X

α 1 antitrypsin

20 HNF 1 α

Proteins localised to the lung, such as secretory component (binds IgA).

Proteins localised to the Heart, for example HSP 27. This is associated with dilated

25 cardiomyopathy.

Proteins localised to the skin, for example keratin.

Bone specific proteins, such as bone morphogenic proteins (BMPs), which are a subset of 30 the transforming growth factor β superfamily that demonstrate osteogenic activity.

Examples include BMP-2, -4, -5, -6, -7 (also referred to as osteogenic protein (OP-1) and -8 (OP-2).

Tumour specific proteins, including human trophoblast antigen, herceptin receptor, oestrogen receptor, cathepsins eg cathepsin B (found in liver and spleen).

Disease-specific proteins, such as antigens expressed only on activated T-cells: including

5 LAG-3 (lymphocyte activation gene), osteoprotegerin ligand (OPGL) see *Nature* 402, 304-309; 1999, OX40 (a member of the TNF receptor family, expressed on activated T cells and the only costimulatory T cell molecule known to be specifically up-regulated in human T cell leukaemia virus type-I (HTLV-I)-producing cells.) See *J Immunol.* 2000 Jul 1;165(1):263-70; Metalloproteases (associated with arthritis/cancers), including

10 CG6512 Drosophila, human paraplegin, human FtsH, human AFG3L2, murine ftsH; angiogenic growth factors, including acidic fibroblast growth factor (FGF-1), basic fibroblast growth factor (FGF-2), Vascular endothelial growth factor / vascular permeability factor (VEGF/VPF), transforming growth factor-a (TGF a), tumor necrosis factor-alpha (TNF- α), angiogenin, interleukin-3 (IL-3), interleukin-8 (IL-8), platelet-derived endothelial growth factor (PD-ECGF), placental growth factor (PIGF), midkine platelet-derived growth factor-BB (PDGF), fractalkine.

15

Stress proteins (heat shock proteins)

HSPs are normally found intracellularly. When they are found extracellularly, it is an 20 indicator that a cell has died and spilled out its contents. This unprogrammed cell death (necrosis) only occurs when as a result of trauma, disease or injury and therefore *in vivo*, extracellular HSPs trigger a response from the immune system that will fight infection and disease. A dual specific which binds to extracellular HSP can be localised to a disease site.

25

Proteins involved in Fc transport

Brambell receptor (also known as FcRB)

This Fc receptor has two functions, both of which are potentially useful for delivery

The functions are

30 (1) The transport of IgG from mother to child across the placenta
(2) the protection of IgG from degradation thereby prolonging its serum half life of IgG. It is thought that the receptor recycles IgG from endosome.

See Holliger *et al*, Nat Biotechnol 1997 Jul;15(7):632-6.

Ligands according to the invention may be designed to be specific for the above targets without requiring any increase in or increasing half life *in vivo*. For example, ligands 5 according to the invention can be specific for targets selected from the foregoing which are tissue-specific, thereby enabling tissue-specific targeting of the dual specific ligand, or a dAb monomer that binds a tissue-specific therapeutically relevant target, irrespective of any increase in half-life, although this may result. Moreover, where the ligand or dAb monomer targets kidney or liver, this may redirect the ligand or dAb monomer to an 10 alternative clearance pathway *in vivo* (for example, the ligand may be directed away from liver clearance to kidney clearance).

H: Use of multispecific ligands according to the second configuration of the invention

15 Multispecific ligands according to the method of the second configuration of the present invention may be employed in *in vivo* therapeutic and prophylactic applications, *in vitro* and *in vivo* diagnostic applications, *in vitro* assay and reagent applications, and the like. For example antibody molecules may be used in antibody based assay techniques, such as 20 ELISA techniques, according to methods known to those skilled in the art.

As alluded to above, the multispecific ligands according to the invention are of use in diagnostic, prophylactic and therapeutic procedures. Multispecific antibodies according to the invention are of use diagnostically in Western analysis and *in situ* protein detection by 25 standard immunohistochemical procedures; for use in these applications, the ligands may be labelled in accordance with techniques known to the art. In addition, such antibody polypeptides may be used preparatively in affinity chromatography procedures, when complexed to a chromatographic support, such as a resin. All such techniques are well known to one of skill in the art.

30 Diagnostic uses of the closed conformation multispecific ligands according to the invention include homogenous assays for analytes which exploit the ability of closed conformation multispecific ligands to bind two targets in competition, such that two

targets cannot bind simultaneously (a closed conformation), or alternatively their ability to bind two targets simultaneously (an open conformation).

A true homogenous immunoassay format has been avidly sought by manufacturers of diagnostics and research assay systems used in drug discovery and development. The main diagnostics markets include human testing in hospitals, doctor's offices and clinics, commercial reference laboratories, blood banks, and the home, non-human diagnostics (for example food testing, water testing, environmental testing, bio-defence, and veterinary testing), and finally research (including drug development; basic research and academic research).

At present all these markets utilise immunoassay systems that are built around chemiluminescent, ELISA, fluorescence or in rare cases radio-immunoassay technologies. Each of these assay formats requires a separation step (separating bound from un-bound reagents). In some cases, several separation steps are required. Adding these additional steps adds reagents and automation, takes time, and affects the ultimate outcome of the assays. In human diagnostics, the separation step may be automated, which masks the problem, but does not remove it. The robotics, additional reagents, additional incubation times, and the like add considerable cost and complexity. In drug development, such as high throughput screening, where literally millions of samples are tested at once, with very low levels of test molecule, adding additional separation steps can eliminate the ability to perform a screen. However, avoiding the separation creates too much noise in the read out. Thus, there is a need for a true homogenous format that provides sensitivities at the range obtainable from present assay formats. Advantageously, an assay possesses fully quantitative read-outs with high sensitivity and a large dynamic range. Sensitivity is an important requirement, as is reducing the amount of sample required. Both of these features are features that a homogenous system offers. This is very important in point of care testing, and in drug development where samples are precious. Heterogenous systems, as currently available in the art, require large quantities of sample and expensive reagents

Applications for homogenous assays include cancer testing, where the biggest assay is that for Prostate Specific Antigen, used in screening men for prostate cancer. Other

applications include fertility testing, which provides a series of tests for women attempting to conceive including beta-hcg for pregnancy. Tests for infectious diseases, including hepatitis, HIV, rubella, and other viruses and microorganisms and sexually transmitted diseases. Tests are used by blood banks, especially tests for HIV, hepatitis A, 5 B, C, non A non B. Therapeutic drug monitoring tests include monitoring levels of prescribed drugs in patients for efficacy and to avoid toxicity, for example digoxin for arrhythmia, and phenobarbital levels in psychotic cases; theophylline for asthma. Diagnostic tests are moreover useful in abused drug testing, such as testing for cocaine, marijuana and the like. Metabolic tests are used for measuring thyroid function, anaemia 10 and other physiological disorders and functions.

15 The homogenous immunoassay format is moreover useful in the manufacture of standard clinical chemistry assays. The inclusion of immunoassays and chemistry assays on the same instrument is highly advantageous in diagnostic testing. Suitable chemical assays include tests for glucose, cholesterol, potassium, and the like.

20 A further major application for homogenous immunoassays is drug discovery and development: high throughput screening includes testing combinatorial chemistry libraries versus targets in ultra high volume. Signal is detected, and positive groups then split into smaller groups, and eventually tested in cells and then animals. Homogenous assays may be used in all these types of test. In drug development, especially animal studies and clinical trials heavy use of immunoassays is made. Homogenous assays greatly accelerate and simplify these procedures. Other Applications include food and 25 beverage testing: testing meat and other foods for E. coli, salmonella, etc; water testing, including testing at water plants for all types of contaminants including E. coli; and veterinary testing.

30 In a broad embodiment, the invention provides a binding assay comprising a detectable agent which is bound to a closed conformation multispecific ligand according to the invention, and whose detectable properties are altered by the binding of an analyte to said closed conformation multispecific ligand. Such an assay may be configured in several different ways, each exploiting the above properties of closed conformation multispecific ligands.

The assay relies on the direct or indirect displacement of an agent by the analyte, resulting in a change in the detectable properties of the agent. For example, where the agent is an enzyme which is capable of catalysing a reaction which has a detectable end-point, said 5 enzyme can be bound by the ligand such as to obstruct its active site, thereby inactivating the enzyme. The analyte, which is also bound by the closed conformation multispecific ligand, displaces the enzyme, rendering it active through freeing of the active site. The enzyme is then able to react with a substrate, to give rise to a detectable event. In an alternative embodiment, the ligand may bind the enzyme outside of the active site, 10 influencing the conformation of the enzyme and thus altering its activity. For example, the structure of the active site may be constrained by the binding of the ligand, or the binding of cofactors necessary for activity may be prevented.

The physical implementation of the assay may take any form known in the art. For 15 example, the closed conformation multispecific ligand/enzyme complex may be provided on a test strip; the substrate may be provided in a different region of the test strip, and a solvent containing the analyte allowed to migrate through the ligand/enzyme complex, displacing the enzyme, and carrying it to the substrate region to produce a signal. Alternatively, the ligand/enzyme complex may be provided on a test stick or other solid 20 phase, and dipped into an analyte/substrate solution, releasing enzyme into the solution in response to the presence of analyte.

Since each molecule of analyte potentially releases one enzyme molecule, the assay is quantitative, with the strength of the signal generated in a given time being dependent on 25 the concentration of analyte in the solution.

Further configurations using the analyte in a closed conformation are possible. For example, the closed conformation multispecific ligand may be configured to bind an enzyme in an allosteric site, thereby activating the enzyme. In such an embodiment, the 30 enzyme is active in the absence of analyte. Addition of the analyte displaces the enzyme and removes allosteric activation, thus inactivating the enzyme.

In the context of the above embodiments which employ enzyme activity as a measure of the analyte concentration, activation or inactivation of the enzyme refers to an increase or decrease in the activity of the enzyme, measured as the ability of the enzyme to catalyse a signal-generating reaction. For example, the enzyme may catalyse the conversion of an undetectable substrate to a detectable form thereof. For example, horseradish peroxidase is widely used in the art together with chromogenic or chemiluminescent substrates, which are available commercially. The level of increase or decrease of the activity of the enzyme may be between 10% and 100%, such as 20%, 30%, 40%, 50%, 60%, 70%, 80% or 90%; in the case of an increase in activity, the increase may be more than 100%, i.e. 200%, 300%, 500% or more, or may not be measurable as a percentage if the baseline activity of the inhibited enzyme is undetectable.

In a further configuration, the closed conformation multispecific ligand may bind the substrate of an enzyme/substrate pair, rather than the enzyme. The substrate is therefore unavailable to the enzyme until released from the closed conformation multispecific ligand through binding of the analyte. The implementations for this configuration are as for the configurations which bind enzyme.

Moreover, the assay may be configured to bind a fluorescent molecule, such as a fluorescein or another fluorophore, in a conformation such that the fluorescence is quenched on binding to the ligand. In this case, binding of the analyte to the ligand will displace the fluorescent molecule, thus producing a signal. Alternatives to fluorescent molecules which are useful in the present invention include luminescent agents, such as luciferin/luciferase, and chromogenic agents, including agents commonly used in immunoassays such as HRP.

Therapeutic and prophylactic uses of multispecific ligands prepared according to the invention involve the administration of ligands according to the invention to a recipient mammal, such as a human. Multi-specificity can allow antibodies to bind to multimeric antigen with great avidity. Multispecific ligands can allow the cross-linking of two antigens, for example in recruiting cytotoxic T-cells to mediate the killing of tumour cell lines.

Substantially pure ligands or binding proteins thereof, for example dAb monomers, of at least 90 to 95% homogeneity are preferred for administration to a mammal, and 98 to 99% or more homogeneity is most preferred for pharmaceutical uses, especially when the mammal is a human. Once purified, partially or to homogeneity as desired, the ligands 5 may be used diagnostically or therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings and the like (Lefkovite and Pernis, (1979 and 1981) *Immunological Methods*, Volumes I and II, Academic Press, NY).

10 The ligands or binding proteins thereof, for example dAb monomers, of the present invention will typically find use in preventing, suppressing or treating inflammatory states, allergic hypersensitivity, cancer, bacterial or viral infection, and autoimmune disorders (which include, but are not limited to, Type I diabetes, asthma, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, Crohn's disease and 15 myasthenia gravis).

In the instant application, the term "prevention" involves administration of the protective composition prior to the induction of the disease. "Suppression" refers to administration of the composition after an inductive event, but prior to the clinical appearance of the 20 disease. "Treatment" involves administration of the protective composition after disease symptoms become manifest.

Animal model systems which can be used to screen the effectiveness of the antibodies or binding proteins thereof in protecting against or treating the disease are available. 25 Methods for the testing of systemic lupus erythematosus (SLE) in susceptible mice are known in the art (Knight *et al.* (1978) *J. Exp. Med.*, 147: 1653; Reinersten *et al.* (1978) *New Eng. J. Med.*, 299: 515). Myasthenia Gravis (MG) is tested in SJL/J female mice by inducing the disease with soluble AchR protein from another species (Lindstrom *et al.* (1988) *Adv. Immunol.*, 42: 233). Arthritis is induced in a susceptible strain of mice by 30 injection of Type II collagen (Stuart *et al.* (1984) *Ann. Rev. Immunol.*, 42: 233). A model by which adjuvant arthritis is induced in susceptible rats by injection of mycobacterial heat shock protein has been described (Van Eden *et al.* (1988) *Nature*, 331: 171). Thyroiditis is induced in mice by administration of thyroglobulin as described (Maron *et*

al. (1980) *J. Exp. Med.*, 152: 1115). Insulin dependent diabetes mellitus (IDDM) occurs naturally or can be induced in certain strains of mice such as those described by Kanasawa *et al.* (1984) *Diabetologia*, 27: 113. EAE in mouse and rat serves as a model for MS in human. In this model, the demyelinating disease is induced by administration 5 of myelin basic protein (see Paterson (1986) *Textbook of Immunopathology*, Mischer *et al.*, eds., Grune and Stratton, New York, pp. 179-213; McFarlin *et al.* (1973) *Science*, 179: 478: and Satoh *et al.* (1987) *J. Immunol.*, 138: 179).

Generally, the present ligands will be utilised in purified form together with 10 pharmacologically appropriate carriers. Typically, these carriers include aqueous or alcoholic/aqueous solutions, emulsions or suspensions, any including saline and/or buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride and lactated Ringer's. Suitable physiologically-acceptable adjuvants, if necessary to keep a polypeptide complex in suspension, may be chosen from 15 thickeners such as carboxymethylcellulose, polyvinylpyrrolidone, gelatin and alginates.

Intravenous vehicles include fluid and nutrient replenishers and electrolyte replenishers, such as those based on Ringer's dextrose. Preservatives and other additives, such as 20 antimicrobials, antioxidants, chelating agents and inert gases, may also be present (Mack (1982) *Remington's Pharmaceutical Sciences*, 16th Edition).

The ligands of the present invention may be used as separately administered compositions or in conjunction with other agents. These can include various immunotherapeutic drugs, such as cyclosporine, methotrexate, adriamycin or cisplatin, and immunotoxins. 25 Pharmaceutical compositions can include "cocktails" of various cytotoxic or other agents in conjunction with the ligands of the present invention, or even combinations of ligands according to the present invention having different specificities, such as ligands selected using different target antigens or epitopes, whether or not they are pooled prior to administration.

30 The route of administration of pharmaceutical compositions according to the invention may be any of those commonly known to those of ordinary skill in the art. For therapy, including without limitation immunotherapy, the selected ligands thereof of the invention

can be administered to any patient in accordance with standard techniques. The administration can be by any appropriate mode, including parenterally, intravenously, intramuscularly, intraperitoneally, transdermally, *via* the pulmonary route, or also, appropriately, by direct infusion with a catheter. The dosage and frequency of 5 administration will depend on the age, sex and condition of the patient, concurrent administration of other drugs, counterindications and other parameters to be taken into account by the clinician.

10 The ligands of this invention can be lyophilised for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immunoglobulins and art-known lyophilisation and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilisation and reconstitution can lead to varying degrees of antibody activity loss (e.g. with conventional immunoglobulins, IgM antibodies tend to have greater activity loss than IgG antibodies) 15 and that use levels may have to be adjusted upward to compensate.

20 The compositions containing the present ligands or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In certain therapeutic applications, an adequate amount to accomplish at least partial inhibition, suppression, modulation, killing, or some other measurable parameter, of a population of selected cells is defined as a "therapeutically-effective dose". Amounts needed to achieve this dosage will depend upon the severity of the disease and the general state of the patient's own immune system, but generally range from 0.005 to 5.0 mg of ligand, e.g. antibody, receptor (e.g. a T-cell 25 receptor) or binding protein thereof *per* kilogram of body weight, with doses of 0.05 to 2.0 mg/kg/dose being more commonly used. For prophylactic applications, compositions containing the present ligands or cocktails thereof may also be administered in similar or slightly lower dosages.

30 Treatment performed using the compositions described herein is considered "effective" if one or more symptoms is reduced (e.g., by at least 10% or at least one point on a clinical assessment scale), relative to such symptoms present before treatment, or relative to such symptoms in an individual (human or model animal) not treated with such composition. Symptoms will obviously vary depending upon the disease or disorder targeted, but can

be measured by an ordinarily skilled clinician or technician. Such symptoms can be measured, for example, by monitoring the level of one or more biochemical indicators of the disease or disorder (e.g., levels of an enzyme or metabolite correlated with the disease, affected cell numbers, etc.), by monitoring physical manifestations (e.g., 5 inflammation, tumor size, etc.), or by an accepted clinical assessment scale, for example, the Expanded Disability Status Scale (for multiple sclerosis), the Irvine Inflammatory Bowel Disease Questionnaire (32 point assessment evaluates quality of life with respect to bowel function, systemic symptoms, social function and emotional status - score ranges from 32 to 224, with higher scores indicating a better quality of life), the Quality of Life 10 Rheumatoid Arthritis Scale, or other accepted clinical assessment scale as known in the field. A sustained (e.g., one day or more, preferably longer) reduction in disease or disorder symptoms by at least 10% or by one or more points on a given clinical scale is indicative of "effective" treatment. Similarly, prophylaxis performed using a composition as described herein is "effective" if the onset or severity of one or more symptoms is 15 delayed, reduced or abolished relative to such symptoms in a similar individual (human or animal model) not treated with the composition.

A composition containing a ligand or cocktail thereof according to the present invention may be utilised in prophylactic and therapeutic settings to aid in the alteration, 20 inactivation, killing or removal of a select target cell population in a mammal. In addition, the selected repertoires of polypeptides described herein may be used extracorporeally or *in vitro* selectively to kill, deplete or otherwise effectively remove a target cell population from a heterogeneous collection of cells. Blood from a mammal may be combined extracorporeally with the ligands, e.g. antibodies, cell-surface receptors or binding 25 proteins thereof whereby the undesired cells are killed or otherwise removed from the blood for return to the mammal in accordance with standard techniques.

I: Use of half-life enhanced dual-specific ligands according to the invention

30

Dual-specific ligands according to the method of the present invention may be employed in *in vivo* therapeutic and prophylactic applications, *in vivo* diagnostic applications and the like.

Therapeutic and prophylactic uses of dual-specific ligands prepared according to the invention involve the administration of ligands according to the invention to a recipient mammal, such as a human. Dual specific antibodies according to the invention comprise 5 at least one specificity for a half-life enhancing molecule; one or more further specificities may be directed against target molecules. For example, a dual-specific IgG may be specific for four epitopes, one of which is on a half-life enhancing molecule. Dual-specificity can allow antibodies to bind to multimeric antigen with great avidity. Dual-specific antibodies can allow the cross-linking of two antigens, for example in recruiting 10 cytotoxic T-cells to mediate the killing of tumour cell lines.

Substantially pure ligands or binding proteins thereof, such as dAb monomers, of at least 90 to 95% homogeneity are preferred for administration to a mammal, and 98 to 99% or more homogeneity is most preferred for pharmaceutical uses, especially when the 15 mammal is a human. Once purified, partially or to homogeneity as desired, the ligands may be used diagnostically or therapeutically (including extracorporeally) or in developing and performing assay procedures, immunofluorescent stainings and the like (Lefkovite and Pernis, (1979 and 1981) Immunological Methods, Volumes I and II, Academic Press, NY).

20

The ligands of the present invention will typically find use in preventing, suppressing or treating inflammatory states, allergic hypersensitivity, cancer, bacterial or viral infection, and autoimmune disorders (which include, but are not limited to, Type I diabetes, multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, Crohn's disease 25 and myasthenia gravis).

In the instant application, the term "prevention" involves administration of the protective composition prior to the induction of the disease. "Suppression" refers to administration of the composition after an inductive event, but prior to the clinical appearance of the 30 disease. "Treatment" involves administration of the protective composition after disease symptoms become manifest.

Animal model systems which can be used to screen the effectiveness of the dual specific ligands in protecting against or treating the disease are available. Methods for the testing of systemic lupus erythematosus (SLE) in susceptible mice are known in the art (Knight *et al.* (1978) *J. Exp. Med.*, **147**: 1653; Reinersten *et al.* (1978) *New Eng. J. Med.*, **299**: 5 515). Myasthenia Gravis (MG) is tested in SJL/J female mice by inducing the disease with soluble AchR protein from another species (Lindstrom *et al.* (1988) *Adv. Immunol.*, **42**: 233). Arthritis is induced in a susceptible strain of mice by injection of Type II collagen (Stuart *et al.* (1984) *Ann. Rev. Immunol.*, **42**: 233). A model by which adjuvant 10 arthritis is induced in susceptible rats by injection of mycobacterial heat shock protein has been described (Van Eden *et al.* (1988) *Nature*, **331**: 171). Thyroiditis is induced in mice by administration of thyroglobulin as described (Maron *et al.* (1980) *J. Exp. Med.*, **152**: 1115). Insulin dependent diabetes mellitus (IDDM) occurs naturally or can be induced in certain strains of mice such as those described by Kanasawa *et al.* (1984) *Diabetologia*, **27**: 113. EAE in mouse and rat serves as a model for MS in human. In this model, the 15 demyelinating disease is induced by administration of myelin basic protein (see Paterson (1986) *Textbook of Immunopathology*, Mischer *et al.*, eds., Grune and Stratton, New York, pp. 179-213; McFarlin *et al.* (1973) *Science*, **179**: 478: and Satoh *et al.* (1987) *J. Immunol.*, **138**: 179).

20 Dual specific ligands according to the invention and dAb monomers able to bind to extracellular targets involved in endocytosis (e.g. Clathrin) enable dual specific ligands to be endocytosed, enabling another specificity able to bind to an intracellular target to be delivered to an intracellular environment. This strategy requires a dual specific ligand with physical properties that enable it to remain functional inside the cell. Alternatively, 25 if the final destination intracellular compartment is oxidising, a well folding ligand may not need to be disulphide free.

Generally, the present dual specific ligands will be utilised in purified form together with pharmacologically appropriate carriers. Typically, these carriers include aqueous or 30 alcoholic/aqueous solutions, emulsions or suspensions, any including saline and/or buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride and lactated Ringer's. Suitable physiologically-acceptable

adjuvants, if necessary to keep a polypeptide complex in suspension, may be chosen from thickeners such as carboxymethylcellulose, polyvinylpyrrolidone, gelatin and alginates.

5 Intravenous vehicles include fluid and nutrient replenishers and electrolyte replenishers, such as those based on Ringer's dextrose. Preservatives and other additives, such as antimicrobials, antioxidants, chelating agents and inert gases, may also be present (Mack (1982) *Remington's Pharmaceutical Sciences*, 16th Edition).

10 The ligands of the present invention may be used as separately administered compositions or in conjunction with other agents. These can include various immunotherapeutic drugs, such as cyclosporine, methotrexate, adriamycin or cisplatin, and immunotoxins. Pharmaceutical compositions can include "cocktails" of various cytotoxic or other agents in conjunction with the ligands of the present invention.

15 The route of administration of pharmaceutical compositions according to the invention may be any of those commonly known to those of ordinary skill in the art. For therapy, including without limitation immunotherapy, the ligands of the invention can be administered to any patient in accordance with standard techniques. The administration 20 can be by any appropriate mode, including parenterally, intravenously, intramuscularly, intraperitoneally, transdermally, via the pulmonary route, or also, appropriately, by direct infusion with a catheter. The dosage and frequency of administration will depend on the age, sex and condition of the patient, concurrent administration of other drugs, counterindications and other parameters to be taken into account by the clinician.

25 The ligands of the invention can be lyophilised for storage and reconstituted in a suitable carrier prior to use. This technique has been shown to be effective with conventional immunoglobulins and art-known lyophilisation and reconstitution techniques can be employed. It will be appreciated by those skilled in the art that lyophilisation and 30 reconstitution can lead to varying degrees of antibody activity loss (e.g. with conventional immunoglobulins, IgM antibodies tend to have greater activity loss than IgG antibodies) and that use levels may have to be adjusted upward to compensate.

The compositions containing the present ligands or a cocktail thereof can be administered for prophylactic and/or therapeutic treatments. In certain therapeutic applications, an adequate amount to accomplish at least partial inhibition, suppression, modulation, killing, or some other measurable parameter, of a population of selected cells 5 is defined as a "therapeutically-effective dose". Amounts needed to achieve this dosage will depend upon the severity of the disease and the general state of the patient's own immune system, but generally range from 0.005 to 5.0 mg of ligand *per* kilogram of body weight, with doses of 0.05 to 2.0 mg/kg/dose being more commonly used. For prophylactic applications, compositions containing the present ligands or cocktails thereof 10 may also be administered in similar or slightly lower dosages.

A composition containing a ligand according to the present invention may be utilised in prophylactic and therapeutic settings to aid in the alteration, inactivation, killing or removal of a select target cell population in a mammal.

15 In addition, the selected repertoires of polypeptides described herein may be used extracorporeally or *in vitro* selectively to kill, deplete or otherwise effectively remove a target cell population from a heterogeneous collection of cells. Blood from a mammal may be combined extracorporeally with the ligands, e.g. antibodies, cell- surface 20 receptors or binding proteins thereof whereby the undesired cells are killed or otherwise removed from the blood for return to the mammal in accordance with standard techniques.

The invention is further described, for the purposes of illustration only, in the following 25 examples. As used herein, for the purposes of dAb nomenclature, human TNF α is referred to as TAR1 and human TNF α receptor 1 (p55 receptor) is referred to as TAR2.

Example 1. Selection of a dual specific scFv antibody (K8) directed against human 30 serum albumin (HSA) and β -galactosidase (β -gal)

This example explains a method for making a dual specific antibody directed against β -gal and HSA in which a repertoire of V κ variable domains linked to a germline (dummy)

V_H domain is selected for binding to β -gal and a repertoire of V_H variable domains linked to a germline (dummy) V_K domain is selected for binding to HSA. The selected variable V_H HSA and V_K β -gal domains are then combined and the antibodies selected for binding to β -gal and HSA. HSA is a half-life increasing protein found in human blood.

5

Four human phage antibody libraries were used in this experiment.

Library 1	Germline V_K /DVT V_H	8.46×10^7
Library 2	Germline V_K /NNK V_H	9.64×10^7
10 Library 3	Germline V_H /DVT V_K	1.47×10^8
Library 4	Germline V_H /NNK V_K	1.45×10^8

15 All libraries are based on a single human framework for V_H (V3-23/DP47 and J_H4b) and V_K (O12/O2/DPK9 and J_K1) with side chain diversity incorporated in complementarity determining regions (CDR2 and CDR3).

20 Library 1 and Library 2 contain a dummy V_K sequence, whereas the sequence of V_H is diversified at positions H50, H52, H52a, H53, H55, H56, H58, H95, H96, H97 and H98 (DVT or NNK encoded, respectively) (Figure 1). Library 3 and Library 4 contain a dummy V_H sequence, whereas the sequence of V_K is diversified at positions L50, L53, L91, L92, L93, L94 and L96 (DVT or NNK encoded, respectively) (Figure 1). The libraries are in phagemid pIT2/ScFv format (Figure 2) and have been preselected for binding to generic ligands, Protein A and Protein L, so that the majority of clones in the unselected libraries are functional. The sizes of the libraries shown above correspond to the sizes after preselection. Library 1 and Library 2 were mixed prior to selections on antigen to yield a single V_H /dummy V_K library and Library 3 and Library 4 were mixed to form a single V_K /dummy V_H library.

30 Three rounds of selections were performed on β -gal using V_K /dummy V_H library and three rounds of selections were performed on HSA using V_H /dummy V_K library. In the

case of β -gal the phage titres went up from 1.1×10^6 in the first round to 2.0×10^8 in the third round. In the case of HSA the phage titres went up from 2×10^4 in the first round to 1.4×10^9 in the third round. The selections were performed as described by Griffith *et al.*, (1993), except that KM13 helper phage (which contains a pIII protein with a protease 5 cleavage site between the D2 and D3 domains) was used and phage were eluted with 1 mg/ml trypsin in PBS. The addition of trypsin cleaves the pIII proteins derived from the helper phage (but not those from the phagemid) and elutes bound scFv-phage fusions by cleavage in the c-myc tag (Figure 2), thereby providing a further enrichment for phages expressing functional scFvs and a corresponding reduction in background (Kristensen & 10 Winter, *Folding & Design* 3: 321-328, Jul 9, 1998). Selections were performed using immunotubes coated with either HSA or β -gal at 100 μ g/ml concentration.

To check for binding, 24 colonies from the third round of each selection were screened by 15 monoclonal phage ELISA. Phage particles were produced as described by Harrison *et al.*, *Methods Enzymol.* 1996;267:83-109. 96-well ELISA plates were coated with 100 μ l of HSA or β -gal at 10 μ g/ml concentration in PBS overnight at 4°C. A standard ELISA protocol was followed (Hoogenboom *et al.*, 1991) using detection of bound phage with anti-M13-HRP conjugate. A selection of clones gave ELISA signals of greater than 1.0 with 50 μ l supernatant.

20 Next, DNA preps were made from V_H /dummy V_K library selected on HSA and from V_K /dummy V_H library selected on β -gal using the QIAprep Spin Miniprep kit (Qiagen). To access most of the diversity, DNA preps were made from each of the three rounds of 25 selections and then pulled together for each of the antigens. DNA preps were then digested with SalI/NotI overnight at 37°C. Following gel purification of the fragments, V_K chains from the V_K /dummy V_H library selected on β -gal were ligated in place of a dummy V_K chain of the V_H /dummy V_K library selected on HSA creating a library of 3.3 $\times 10^9$ clones.

30 This library was then either selected on HSA (first round) and β -gal (second round), HSA/ β -gal selection, or on β -gal (first round) and HSA (second round), β -gal/HSA

selection. Selections were performed as described above. In each case after the second round 48 clones were tested for binding to HSA and β -gal by the monoclonal phage ELISA (as described above) and by ELISA of the soluble scFv fragments. Soluble antibody fragments were produced as described by Harrison *et al.*, (1996), and standard 5 ELISA protocol was followed Hoogenboom *et al.* (1991) Nucleic Acids Res., 19: 4133, except that 2% Tween/PBS was used as a blocking buffer and bound scFvs were detected with Protein L-HRP. Three clones (E4, E5 and E8) from the HSA/ β -gal selection and two clones (K8 and K10) from the β -gal/HSA selection were able to bind both antigens. scFvs from these clones were PCR amplified and sequenced as described by Ignatovich *et al.*, 10 (1999) J Mol Biol 1999 Nov 26;294(2):457-65, using the primers LMB3 and pHENseq. Sequence analysis revealed that all clones were identical. Therefore, only one clone encoding a dual specific antibody (K8) was chosen for further work (Figure 3).

15 **Example 2. Characterisation of the binding properties of the K8 antibody.**

Firstly, the binding properties of the K8 antibody were characterised by the monoclonal phage ELISA. A 96-well plate was coated with 100 μ l of HSA and β -gal alongside with alkaline phosphatase (APS), bovine serum albumin (BSA), peanut agglutinin, lysozyme 20 and cytochrome c (to check for cross-reactivity) at 10 μ g/ml concentration in PBS overnight at 4°C. The phagemid from K8 clone was rescued with KM13 as described by Harrison *et al.*, (1996) and the supernatant (50 μ l) containing phage assayed directly. A standard ELISA protocol was followed (Hoogenboom *et al.*, 1991) using detection of bound phage with anti-M13-HRP conjugate. The dual specific K8 antibody was found to 25 bind to HSA and β -gal when displayed on the surface of the phage with absorbance signals greater than 1.0 (Figure 4). Strong binding to BSA was also observed (Figure 4). Since HSA and BSA are 76% homologous on the amino acid level, it is not surprising that K8 antibody recognised both of these structurally related proteins. No cross-reactivity with other proteins was detected (Figure 4).

30

Secondly, the binding properties of the K8 antibody were tested in a soluble scFv ELISA. Production of the soluble scFv fragment was induced by IPTG as described by Harrison *et al.*, (1996). To determine the expression levels of K8 scFv, the soluble antibody

fragments were purified from the supernatant of 50ml inductions using Protein A-Sepharose columns as described by Harlow and Lane, *Antibodies: a Laboratory Manual*, (1988) Cold Spring Harbor. OD₂₈₀ was then measured and the protein concentration calculated as described by Sambrook *et al.*, (1989). K8 scFv was produced in supernatant 5 at 19mg/l.

A soluble scFv ELISA was then performed using known concentrations of the K8 antibody fragment. A 96-well plate was coated with 100µl of HSA, BSA and β-gal at 10µg/ml and 100µl of Protein A at 1µg/ml concentration. 50µl of the serial dilutions of 10 the K8 scFv was applied and the bound antibody fragments were detected with Protein L-HRP. ELISA results confirmed the dual specific nature of the K8 antibody (Figure 5).

To confirm that binding to β-gal is determined by the V_K domain and binding to HSA/BSA by the V_H domain of the K8 scFv antibody, the V_K domain was cut out from 15 K8 scFv DNA by SalI/NotI digestion and ligated into a SalI/NotI digested pIT2 vector containing dummy V_H chain (Figures 1 and 2). Binding characteristics of the resulting clone K8V_K/dummy V_H were analysed by soluble scFv ELISA. Production of the soluble scFv fragments was induced by IPTG as described by Harrison *et al.*, (1996) and the supernatant (50µ) containing scFvs assayed directly. Soluble scFv ELISA was performed 20 as described in Example 1 and the bound scFvs were detected with Protein L-HRP. The ELISA results revealed that this clone was still able to bind β-gal, whereas binding to BSA was abolished (Figure 6).

Example 3. Selection of single V_H domain antibodies antigens A and B and single 25 V_K domain antibodies directed against antigens C and D.

This example describes a method for making single V_H domain antibodies directed against antigens A and B and single V_K domain antibodies directed against antigens C and D by selecting repertoires of virgin single antibody variable domains for binding to 30 these antigens in the absence of the complementary variable domains.

Selections and characterisation of the binding clones is performed as described previously (see Example 5, PCT/GB 02/003014). Four clones are chosen for further work:

5 VH1 – Anti A V_H

VH2 – Anti B V_H

VK1 – Anti C V_K

VK2 – Anti D V_K

10 The procedures described above in Examples 1-3 may be used, in a similar manner as that described, to produce dimer molecules comprising combinations of V_H domains (i.e., V_H-V_H ligands) and combinations of V_L domains (V_L-V_L ligands).

Example 4. Creation and characterisation of the dual specific ScFv antibodies

15 (VH1/VH2 directed against antigens A and B and VK1/VK2 directed against antigens C and D).

This example demonstrates that dual specific ScFv antibodies (VH1/VH2 directed against antigens A and B and VK1/VK2 directed against antigens C and D) could be created by 20 combining V_K and V_H single domains selected against respective antigens in a ScFv vector.

To create dual specific antibody VH1/VH2, VH1 single domain is excised from variable domain vector 1 (Figure 7) by *NcoI/XhoI* digestion and ligated into *NcoI/XhoI* digested 25 variable domain vector 2 (Figure 7) to create VH1/ variable domain vector 2. VH2 single domain is PCR amplified from variable domain vector 1 using primers to introduce *SaII* restriction site to the 5' end and *NotI* restriction site to the 3' end. The PCR product is then digested with *SaII/NotI* and ligated into *SaII/NotI* digested VH1/ variable domain vector 2 to create VH1/VH2/ variable domain vector 2.

30 VK1/VK2/ variable domain vector 2 is created in a similar way. The dual specific nature of the produced VH1/VH2 ScFv and VK1/VK2 ScFv is tested in a soluble ScFv ELISA

as described previously (see Example 6, PCT/GB 02/003014). Competition ELISA is performed as described previously (see Example 8, PCT/GB 02/003014).

Possible outcomes:

- 5 -VH1/VH2 ScFv is able to bind antigens A and B simultaneously
- VK1/VK2 ScFv is able to bind antigens C and D simultaneously
- VH1/VH2 ScFv binding is competitive (when bound to antigen A, VH1/VH2 ScFv cannot bind to antigen B)
- VK1/VK2 ScFv binding is competitive (when bound to antigen C, VK1/VK2 ScFv
- 10 cannot bind to antigen D)

Example 5. Construction of dual specific VH1/VH2 Fab and VK1/VK2 Fab and analysis of their binding properties.

- 15 To create VH1/VH2 Fab, VH1 single domain is ligated into *Nco*I/*Xho*I digested CH vector (Figure 8) to create VH1/CH and VH2 single domain is ligated into *Sal*II/*Not*I digested CK vector (Figure 9) to create VH2/CK. Plasmid DNA from VH1/CH and VH2/CK is used to co-transform competent *E. coli* cells as described previously (see Example 8, PCT/GB02/003014).

20 The clone containing VH1/CH and VH2/CK plasmids is then induced by IPTG to produce soluble VH1/VH2 Fab as described previously (see Example 8, PCT/GB 02/003014).

- 25 VK1/VK2 Fab is produced in a similar way.

Binding properties of the produced Fabs are tested by competition ELISA as described previously (see Example 8, PCT/GB 02/003014).

- 30 Possible outcomes:

- VH1/VH2 Fab is able to bind antigens A and B simultaneously
- VK1/VK2 Fab is able to bind antigens C and D simultaneously

- VH1/VH2 Fab binding is competitive (when bound to antigen A, VH1/VH2 Fab cannot bind to antigen B)
- VK1/VK2 Fab binding is competitive (when bound to antigen C, VK1/VK2 Fab cannot bind to antigen D)

5

Example 6

Chelating dAb Dimers

10 Summary

VH and VK homo-dimers are created in a dAb-linker-dAb format using flexible polypeptide linkers. Vectors were created in the dAb linker-dAb format containing glycine-serine linkers of different lengths 3U:(Gly₄Ser)₃, 5U:(Gly₄Ser)₅, 7U:(Gly₄Ser)₇. Dimer libraries were created using guiding dAbs upstream of the linker: TAR1-5 (VK), 15 TAR1-27(VK), TAR2-5(VH) or TAR2-6(VK) and a library of corresponding second dAbs after the linker. Using this method, novel dimeric dAbs were selected. The effect of dimerisation on antigen binding was determined by ELISA and BIACore studies and in cell neutralisation and receptor binding assays. Dimerisation of both TAR1-5 and TAR1-27 resulted in significant improvement in binding affinity and neutralisation levels.

20

1.0 Methods

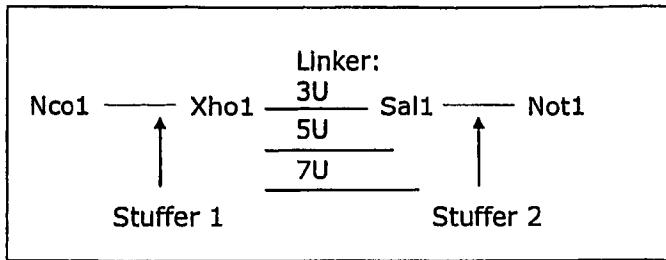
1.1 Library generation

1.1.1 Vectors

pEDA3U, pEDASU and pEDA7U vectors were designed to introduce different linker lengths compatible with the dAb-linker-dAb format. For pEDA3U, sense and anti-sense 25 73-base pair oligo linkers were annealed using a slow annealing program (95°C-5mins, 80°C-10mins, 70°C-15mins, 56°C-15mins, 42°C until use) in buffer containing 0.1M NaCl, 10mM Tris-HCl pH7.4 and cloned using the *Xho*1 and *Not*1 restriction sites. The linkers encompassed 3 (Gly₄Ser) units and a stuffer region housed between *Sall* and 30 *Not*1 cloning sites (scheme 1). In order to reduce the possibility of monomeric dAbs being selected for by phage display, the stuffer region was designed to include 3 stop codons, a *Sac*1 restriction site and a frame shift mutation to put the region out of frame when no second dAb was present. For pEDASU and 7U due to the length of the linkers

required, overlapping oligo-linkers were designed for each vector, annealed and elongated using Klenow. The fragment was then purified and digested using the appropriate enzymes before cloning using the *Xho1* and *Not1* restriction sites.

5



Scheme 1

1.1.2 Library preparation

The N-terminal V gene corresponding to the guiding dAb was cloned upstream of the linker using *Nco1* and *Xho1* restriction sites. VH genes have existing compatible sites, however cloning VK genes required the introduction of suitable restriction sites. This was achieved by using modifying PCR primers (VK-DLIBF: 5' cggccatggcgtaacggacat; VKXho1R: 5' atgtgcgctcgagcggttgattt 3') in 30 cycles of PCR amplification using a 2:1 mixture of SuperTaq (HTBiotechnology Ltd) and *pfu* turbo (Stratagene). This maintained the *Nco1* site at the 5' end while destroying the adjacent *Sal1* site and introduced the *Xho1* site at the 3' end. 5 guiding dAbs were cloned into each of the 3 dimer vectors: TAR1-5 (VK), TAR1-27(VK), TAR2-5(VH), TAR2-6(VK) and TAR2-7(VK). All constructs were verified by sequence analysis.

Having cloned the guiding dAbs upstream of the linker in each of the vectors (pEDA3U, 5U and 7U): TAR1-5 (VK), TAR1-27(VK), TAR2-5(VH) or TAR2-6(VK) a library of corresponding second dAbs were cloned after the linker. To achieve this, the complimentary dAb libraries were PCR amplified from phage recovered from round 1 selections of either a VK library against Human TNF α (at approximately 1×10^6 diversity after round 1) when TAR1-5 or TAR1-27 are the guiding dAbs, or a VH or VK library against human p55 TNF receptor (both at approximately 1×10^5 diversity after round 1) when TAR2-5 or TAR2-6 respectively are the guiding dAbs. For VK libraries PCR amplification was conducted using primers in 30 cycles of PCR amplification using a 2:1 mixture of SuperTaq and *pfu* turbo. VH libraries were PCR amplified using primers in

order to introduce a *SalI* restriction site at the 5' end of the gene. The dAb library PCRs were digested with the appropriate restriction enzymes, ligated into the corresponding vectors down stream of the linker, using *SalI/NotI* restriction sites and electroporated into freshly prepared competent TG1 cells.

5

The titres achieved for each library are as follows:

TAR1-5: pEDA3U = 4×10^8 , pEDA5U = 8×10^7 , pEDA7U = 1×10^8

TAR1-27: pEDA3U = 6.2×10^8 , pEDA5U = 1×10^8 , pEDA7U = 1×10^9

TAR2h-5: pEDA3U = 4×10^7 , pEDA5U = 2×10^8 , pEDA7U = 8×10^7

10 TAR2h-6: pEDA3U = 7.4×10^8 , pEDA5U = 1.2×10^8 , pEDA7U = 2.2×10^8

1.2 Selections

1.2.1 TNF α

Selections were conducted using human TNF α passively coated on immunotubes.

15 Briefly, Immunotubes are coated overnight with 1-4mls of the required antigen. The immunotubes were then washed 3 times with PBS and blocked with 2% milk powder in PBS for 1-2hrs and washed a further 3 times with PBS. The phage solution is diluted in 2% milk powder in PBS and incubated at room temperature for 2hrs. The tubes are then washed with PBS and the phage eluted with 1mg/ml trypsin-PBS. Three selection 20 strategies were investigated for the TAR1-5 dimer libraries. The first round selections were carried out in immunotubes using human TNF α coated at 1 μ g/ml or 20 μ g/ml with 20 washes in PBS 0.1% Tween. TG1 cells are infected with the eluted phage and the titres are determined (eg, Marks et al J Mol Biol. 1991 Dec 5;222(3):581-97, Richmann et al Biochemistry. 1993 Aug 31;32(34):8848-55).

25

The titres recovered were:

pEDA3U = 2.8×10^7 (1 μ g/ml TNF), 1.5×10^8 (20 μ g/ml TNF),

pEDA5U = 1.8×10^7 (1 μ g/ml TNF), 1.6×10^8 (20 μ g/ml TNF)

pEDA7U = 8×10^6 (1 μ g/ml TNF), 7×10^7 (20 μ g/ml TNF).

30

The second round selections were carried out using 3 different methods.

1. In immunotubes, 20 washes with overnight incubation followed by a further 10 washes.

2. In immunotubes, 20 washes followed by 1hr incubation at RT in wash buffer with (1 μ g/ml TNF α) and 10 further washes.

3. Selection on streptavidin beads using 33 pmoles biotinylated human TNF α (Henderikx *et al.*, 2002, *Selection of antibodies against biotinylated antigens*.

5 Antibody Phage Display : Methods and protocols, Ed. O'Brien and Atkin, Humana Press). Single clones from round 2 selections were picked into 96 well plates and crude supernatant preps were made in 2ml 96 well plate format.

	Round 1 Human TNF α immuno tube coating concentration	Round 2 selection method 1	Round 2 selection method 2	Round 2 selection method 3
pEDA3U	1 μ g/ml	1 x 10 ⁹	1.8 x 10 ⁹	2.4 x 10 ¹⁰
pEDA3U	20 μ g/ml	6 x 10 ⁹	1.8 x 10 ¹⁰	8.5 x 10 ¹⁰
pEDA5U	1 μ g/ml	9 x 10 ⁸	1.4 x 10 ⁹	2.8 x 10 ¹⁰
pEDA5U	20 μ g/ml	9.5 x 10 ⁹	8.5 x 10 ⁹	2.8 x 10 ¹⁰
pEDA7U	1 μ g/ml	7.8 x 10 ⁸	1.6 x 10 ⁸	4 x 10 ¹⁰
pEDA7U	20 μ g/ml	1 x 10 ¹⁰	8 x 10 ⁹	1.5 x 10 ¹⁰

10 For TAR1-27, selections were carried out as described previously with the following modifications. The first round selections were carried out in immunotubes using human TNF α coated at 1 μ g/ml or 20 μ g/ml with 20 washes in PBS 0.1%Tween. The second round selections were carried out in immunotubes using 20 washes with overnight incubation followed by a further 20 washes. Single clones from round 2 selections were 15 picked into 96 well plates and crude supernatant preps were made in 2ml 96 well plate format.

TAR1-27 titres are as follows:

	Human TNF α immunotube coating conc	Round 1	Round 2
pEDA3U	1 μ g/ml	4×10^9	6×10^9
pEDA3U	20 μ g/ml	5×10^9	4.4×10^{10}
pEDA5U	1 μ g/ml	1.5×10^9	1.9×10^{10}
pEDA5U	20 μ g/ml	3.4×10^9	3.5×10^{10}
pEDA7U	1 μ g/ml	2.6×10^9	5×10^9
pEDA7U	20 μ g/ml	7×10^9	1.4×10^{10}

1.2.2 TNF RECEPTOR 1 (p55 RECEPTOR; TAR2)

Selections were conducted as described previously for the TAR2h-5 libraries only. 3 rounds of selections were carried out in immunotubes using either 1 μ g/ml human p55 TNF receptor or 10 μ g/ml human p55 TNF receptor with 20 washes in PBS 0.1%Tween with overnight incubation followed by a further 20 washes. Single clones from round 2 and 3 selections were picked into 96 well plates and crude supernatant preps were made in 2ml 96 well plate format.

10 TAR2h-5 titres are as follows:

	Round 1 human p55 TNF receptor immunotube coating concentration	Round 1	Round 2	Round 3
pEDA3U	1 μ g/ml	2.4×10^6	1.2×10^7	1.9×10^9
pEDA3U	10 μ g/ml	3.1×10^7	7×10^7	1×10^9
pEDA5U	1 μ g/ml	2.5×10^6	1.1×10^7	5.7×10^8
pEDA5U	10 μ g/ml	3.7×10^7	2.3×10^8	2.9×10^9
pEDA7U	1 μ g/ml	1.3×10^6	1.3×10^7	1.4×10^9
pEDA7U	10 μ g/ml	1.6×10^7	1.9×10^7	3×10^{10}

1.3 Screening

Single clones from round 2 or 3 selections were picked from each of the 3U, 5U and 7U libraries from the different selections methods, where appropriate. Clones were grown in 2xTY with 100 μ g/ml ampicillin and 1% glucose overnight at 37°C. A 1/100 dilution of this culture was inoculated into 2mls of 2xTY with 100 μ g/ml ampicillin and 0.1% glucose in 2ml, 96 well plate format and grown at 37°C shaking until OD600 was

approximately 0.9. The culture was then induced with 1mM IPTG overnight at 30°C. The supernatants were clarified by centrifugation at 4000rpm for 15 mins in a sorval plate centrifuge. The supernatant preps the used for initial screening.

5 **1.3.1 ELISA**

Binding activity of dimeric recombinant proteins was compared to monomer by Protein A/L ELISA or by antigen ELISA. Briefly, a 96 well plate is coated with antigen or Protein A/L overnight at 4°C. The plate washed with 0.05% Tween-PBS, blocked for 2hrs with 2% Tween-PBS. The sample is added to the plate incubated for 1 hr at room 10 temperature. The plate is washed and incubated with the secondary reagent for 1hr at room temperature. The plate is washed and developed with TMB substrate. Protein A/L-HRP or India-HRP was used as a secondary reagent. For antigen ELISAs, the antigen concentrations used were 1 μ g/ml in PBS for Human TNF α and human THF receptor 1. Due to the presence of the guiding dAb in most cases dimers gave a positive ELISA 15 signal therefore off rate determination was examined by BIACore.

1.3.2 BIACore

BIACore analysis was conducted for TAR1-5 and TAR2h-5 clones. For screening, Human TNF α was coupled to a CM5 chip at high density (approximately 10000 RUs). 20 50 μ l of Human TNF α (50 μ g/ml) was coupled to the chip at 5 μ l/min in acetate buffer - pH5.5. Regeneration of the chip following analysis using the standard methods is not possible due to the instability of Human TNF α , therefore after each sample was analysed, the chip was washed for 10mins with buffer.

For TAR1-5, clones supernatants from the round 2 selection were screened by BIACore. 25 48 clones were screened from each of the 3U, 5U and 7U libraries obtained using the following selection methods:

R1: 1 μ g/ml human TNF α immunotube, R2 1 μ g/ml human TNF α immunotube, overnight wash.

30 R1: 20 μ g/ml human TNF α immunotube, R2 20 μ g/ml human TNF α immunotube, overnight wash.

R1: 1 μ g/ml human TNF α immunotube, R2 33 pmoles biotinylated human TNF α on beads.

R1: 20 μ g/ml human TNF α immunotube, R2 33 pmoles biotinylated human TNF α beads.

For screening, human p55 TNF receptor was coupled to a CM5 chip at high density (approximately 4000 RUs). 100 μ l of human p55 TNF receptor (10 μ g/ml) was coupled to the chip at 5 μ l/min in acetate buffer - pH5.5. Standard regeneration conditions were 5 examined (glycine pH2 or pH3) but in each case antigen was removed from the surface of the chip therefore as with TNF α , therefore after each sample was analysed, the chip was washed for 10mins with buffer.

For TAR2-5, clones supernatants from the round 2 selection were screened. 10 48 clones were screened from each of the 3U, 5U and 7U libraries, using the following selection methods:

R1: 1 μ g/ml human p55 TNF receptor immunotube, R2 1 μ g/ml human p55 TNF receptor immunotube, overnight wash.

R1: 10 μ g/ml human p55 TNF receptor immunotube, R2 10 μ g/ml human p55 TNF receptor immunotube, overnight wash.

15

1.3.3 Receptor and Cell Assays

The ability of the dimers to neutralise in the receptor assay was conducted as follows:

Receptor binding

20 Anti-TNF dAbs were tested for the ability to inhibit the binding of TNF to recombinant TNF receptor 1 (p55). Briefly, Maxisorp plates were incubated overnight with 30mg/ml anti-human Fc mouse monoclonal antibody (Zymed, San Francisco, USA). The wells were washed with phosphate buffered saline (PBS) containing 0.05% Tween-20 and then blocked with 1% BSA in PBS before being incubated with 100ng/ml TNF receptor 1 Fc 25 fusion protein (R&D Systems, Minneapolis, USA). Anti-TNF dAb was mixed with TNF which was added to the washed wells at a final concentration of 10ng/ml. TNF binding was detected with 0.2mg/ml biotinylated anti-TNF antibody (HyCult biotechnology, Uben, Netherlands) followed by 1 in 500 dilution of horse radish peroxidase labelled streptavidin (Amersham Biosciences, UK) and then incubation with TMB substrate (KPL, 30 Gaithersburg, USA). The reaction was stopped by the addition of HCl and the absorbance was read at 450nm. Anti-TNF dAb activity lead to a decrease in TNF binding and therefore a decrease in absorbance compared with the TNF only control.

L929 Cytotoxicity Assay

Anti-TNF dAbs were also tested for the ability to neutralise the cytotoxic activity of TNF on mouse L929 fibroblasts (Evans, T. (2000) Molecular Biotechnology 15, 243-248). Briefly, L929 cells plated in microtitre plates were incubated overnight with anti-TNF dAb, 100pg/ml TNF and 1mg/ml actinomycin D (Sigma, Poole, UK). Cell viability was measured by reading absorbance at 490nm following an incubation with [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium (Promega, Madison, USA). Anti-TNF dAb activity lead to a decrease in TNF cytotoxicity and therefore an increase in absorbance compared with the TNF only control.

10

In the initial screen, supernatants prepared for BIAcore analysis, described above, were also used in the receptor assay. Further analysis of selected dimers was also conducted in the receptor and cell assays using purified proteins.

15

HeLa IL-8 assay

Anti-TNFR1 or anti-TNF alpha dAbs were tested for the ability to neutralise the induction of IL-8 secretion by TNF in HeLa cells (method adapted from that of Akeson, L. et al (1996) Journal of Biological Chemistry 271, 30517-30523, describing the induction of IL-8 by IL-1 in HUVEC; here we look at induction by human TNF alpha and 20 we use HeLa cells instead of the HUVEC cell line). Briefly, HeLa cells plated in microtitre plates were incubated overnight with dAb and 300pg/ml TNF. Post incubation the supernatant was aspirated off the cells and IL-8 concentration measured via a sandwich ELISA (R&D Systems). Anti-TNFR1 dAb activity lead to a decrease in IL-8 secretion into the supernatant compared with the TNF only control.

25

The L929 assay is used throughout the following experiments; however, the use of the HeLa IL-8 assay is preferred to measure anti-TNF receptor 1 (p55) ligands; the presence of mouse p55 in the L929 assay poses certain limitations in its use.

30

1.4 Sequence analysis

Dimers that proved to have interesting properties in the BIAcore and the receptor assay screens were sequenced. Sequences are detailed in the sequence listing.

1.5 Formatting

1.5.1 TAR1-5-19 dimers

The TAR1-5 dimers that were shown to have good neutralisation properties were re-formatted and analysed in the cell and receptor assays. The TAR1-5 guiding dab was 5 substituted with the affinity matured clone TAR1-5-19. To achieve this TAR1-5 was cloned out of the individual dimer pair and substituted with TAR1-5-19 that had been amplified by PCR. In addition, TAR1-5-19 homodimers were also constructed in the 3U, 5U and 7U vectors. The N terminal copy of the gene was amplified by PCR and cloned as described above and the C-terminal gene fragment was cloned using existing *Sall* and 10 *Not1* restriction sites.

1.5.2 Mutagenesis

The amber stop codon present in dAb2, one of the C-terminal dAbs in the TAR1-5 dimer pairs was mutated to a glutamine by site-directed mutagenesis.

15

1.5.3 Fabs

The dimers containing TAR1-5 or TAR1-5-19 were re-formatted into Fab expression vectors. dAbs were cloned into expression vectors containing either the CK or CH genes using *Sfi1* and *Not1* restriction sites and verified by sequence analysis. The CK vector is 20 derived from a pUC based ampicillin resistant vector and the CH vector is derived from a pACYC chloramphenicol resistant vector. For Fab expression the dAb-CH and dAb-CK constructs were co-transformed into HB2151 cells and grown in 2xTY containing 0.1% glucose, 100 μ g/ml ampicillin and 10 μ g/ml chloramphenicol.

25 1.5.3 Hinge dimerisation

Dimerisation of dAbs via cystine bond formation was examined. A short sequence of amino acids EPKSGDKTHTCPPCP a modified form of the human IgGC1 hinge was engineered at the C terminal region on the dAb. An oligo linker encoding for this sequence was synthesised and annealed, as described previously. The linker was cloned 30 into the pEDA vector containing TAR1-5-19 using *Xhol* and *Not1* restriction sites. Dimerisation occurs *in situ* in the periplasm.

1.6 Expression and purification

1.6.1 Expression

Supernatants were prepared in the 2ml, 96-well plate format for the initial screening as described previously. Following the initial screening process selected dimers were analysed further. Dimer constructs were expressed in TOP10F' or HB2151 cells as supernatants. Briefly, an individual colony from a freshly streaked plate was grown overnight at 37°C in 2xTY with 100µg/ml ampicillin and 1% glucose. A 1/100 dilution of this culture was inoculated into 2xTY with 100µg/ml ampicillin and 0.1% glucose and grown at 37°C shaking until OD600 was approximately 0.9. The culture was then induced with 1mM IPTG overnight at 30°C. The cells were removed by centrifugation and the supernatant purified with protein A or L agarose.

Fab and cysteine hinge dimers were expressed as periplasmic proteins in HB2152 cells. A 1/100 dilution of an overnight culture was inoculated into 2xTY with 0.1% glucose and the appropriate antibiotics and grown at 30°C shaking until OD600 was approximately 0.9. The culture was then induced with 1mM IPTG for 3-4 hours at 25°C. The cells were harvested by centrifugation and the pellet resuspended in periplasmic preparation buffer (30mM Tris-HCl pH8.0, 1mM EDTA, 20% sucrose). Following centrifugation the supernatant was retained and the pellet resuspended in 5mM MgSO₄. The supernatant was harvested again by centrifugation, pooled and purified.

20 1.6.2 Protein A/L purification

Optimisation of the purification of dimer proteins from Protein L agarose (Affitech, Norway) or Protein A agarose (Sigma, UK) was examined. Protein was eluted by batch or by column elution using a peristaltic pump. Three buffers were examined 0.1M Phosphate-citrate buffer pH2.6, 0.2M Glycine pH2.5 and 0.1M Glycine pH2.5. The optimal condition was determined to be under peristaltic pump conditions using 0.1M Glycine pH2.5 over 10 column volumes. Purification from protein A was conducted peristaltic pump conditions using 0.1M Glycine pH2.5.

1.6.3 FPLC purification

30 Further purification was carried out by FPLC analysis on the AKTA Explorer 100 system (Amersham Biosciences Ltd). TAR1-5 and TAR1-5-19 dimers were fractionated by cation exchange chromatography (1ml Resource S – Amersham Biosciences Ltd) eluted with a 0-1M NaCl gradient in 50mM acetate buffer pH4. Hinge dimers were purified by

ion exchange (1ml Resource Q Amersham Biosciences Ltd) eluted with a 0-1M NaCl gradient in 25mMTris HCl pH 8.0. Fabs were purified by size exclusion chromatography using a superose 12 (Amersham Biosciences Ltd) column run at a flow rate of 0.5ml/min in PBS with 0.05% tween. Following purification samples were concentrated using 5 vivaspin 5K cut off concentrators (Vivascience Ltd).

2.0 Results

2.1 TAR1-5 dimers

6 x 96 clones were picked from the round 2 selection encompassing all the libraries and 10 selection conditions. Supernatant preps were made and assayed by antigen and Protein L ELISA, BIACore and in the receptor assays. In ELISAs, positive binding clones were identified from each selection method and were distributed between 3U, 5U and 7U libraries. However, as the guiding dAb is always present it was not possible to discriminate between high and low affinity binders by this method therefore BIACore 15 analysis was conducted.

BIACore analysis was conducted using the 2ml supernatants. BIACore analysis revealed that the dimer Koff rates were vastly improved compared to monomeric TAR1-5. Monomer Koff rate was in the range of 10^{-1} M compared with dimer Koff rates which - 20 were in the range of 10^{-3} - 10^{-4} M. 16 clones that appeared to have very slow off rates were selected, these came from the 3U, 5U and 7U libraries and were sequenced. In addition the supernatants were analysed for the ability to neutralise human TNF α in the receptor assay.

25 6 lead clones (d1-d6 below) that neutralised in these assays and have been sequenced. The results shows that out of the 6 clones obtained there are only 3 different second dAbs (dAb1, dAb2 and dAb3) however where the second dAb is found more than once they are linked with different length linkers.

30 **TAR1-5d1:** 3U linker 2nd dAb=dAb1 - 1 μ g/ml Ag immunotube overnight wash

TAR1-5d2: 3U linker 2nd dAb=dAb2 - 1 μ g/ml Ag immunotube overnight wash

TAR1-5d3: 5U linker 2nd dAb=dAb2 - 1 μ g/ml Ag immunotube overnight wash

TAR1-5d4: 5U linker 2nd dAb=dAb3 - 20 μ g/ml Ag immunotube overnight wash

TAR1-5d5: 5U linker 2nd dAb=dAb1 - 20 μ g/ml Ag immunotube overnight wash

TAR1-5d6: 7U linker 2nd dAb=dAb1- R1:1 μ g/ml Ag immunotube overnight wash,
R2:beads

5 The 6 lead clones were examined further. Protein was produced from the periplasm and supernatant, purified with protein L agarose and examined in the cell and receptor assays. The levels of neutralisation were variable (Table 1). The optimal conditions for protein preparation were determined. Protein produced from HB2151 cells as supernatants gave the highest yield (approximately 10mgs/L of culture). The supernatants were incubated
10 with protein L agarose for 2hrs at room temperature or overnight at 4°C. The beads were washed with PBS/NaCl and packed onto an FPLC column using a peristaltic pump. The beads were washed with 10 column volumes of PBS/NaCl and eluted with 0.1M glycine pH2.5. In general, dimeric protein is eluted after the monomer.

15 TAR1-5d1-6 dimers were purified by FPLC. Three species were obtained, by FPLC purification and were identified by SDS PAGE. One species corresponds to monomer and the other two species corresponds to dimers of different sizes. The larger of the two species is possibly due to the presence of C terminal tags. These proteins were examined in the receptor assay. The data presented in table 1 represents the optimum results
20 obtained from the two dimeric species (Figure 11)

The three second dAbs from the dimer pairs (ie, dAb1, dAb2 and dAb3) were cloned as monomers and examined by ELISA and in the cell and receptor assay. All three dAbs bind specifically to TNF by antigen ELISA and do not cross react with plastic or BSA. As
25 monomers, none of the dAbs neutralise in the cell or receptor assays.

2.1.2 TAR1-5-19 dimers

TAR1-5-19 was substituted for TAR1-5 in the 6 lead clones. Analysis of all TAR1-5-19 dimers in the cell and receptor assays was conducted using total protein (protein L purified only) unless otherwise stated (Table 2). TAR1-5-19d4 and TAR1-5-19d3 have the best ND₅₀ (~5nM) in the cell assay, this is consistent with the receptor assay results and is an improvement over TAR1-5-19 monomer (ND₅₀~30nM). Although purified TAR1-5 dimers give variable results in the receptor and cell assays TAR1-5-19 dimers

were more consistent. Variability was shown when using different elution buffers during the protein purification. Elution using 0.1M Phosphate-citrate buffer pH2.6 or 0.2M Glycine pH2.5 although removing all protein from the protein L agarose in most cases rendered it less functional.

5

TAR1-5-19d4 was expressed in the fermenter and purified on cation exchange FPLC to yield a completely pure dimer. As with TAR1-5d4 three species were obtained, by FPLC purification corresponding to monomer and two dimer species. This dimer was amino acid sequenced. TAR1-5-19 monomer and TAR1-5-19d4 were then examined in the receptor assay and the resulting IC₅₀ for monomer was 30nM and for dimer was 8nM. 10 The results of the receptor assay comparing TAR1-5-19 monomer, TAR1-5-19d4 and TAR1-5d4 is shown in figure 10.

15 TAR1-5-19 homodimers were made in the 3U, 5U and 7U vectors, expressed and purified on Protein L. The proteins were examined in the cell and receptor assays and the resulting IC₅₀s (for receptor assay) and ND₅₀s (for cell assay) were determined (table 3, figure 12).

2.2 Fabs

20 TAR1-5 and TAR1-5-19 dimers were also cloned into Fab format, expressed and purified on protein L agarose. Fabs were assessed in the receptor assays (Table 4). The results showed that for both TAR1-5-19 and TAR1-5 dimers the neutralisation levels were similar to the original Gly₄Ser linker dimers from which they were derived. A TAR1-5-19 Fab where TAR1-5-19 was displayed on both CH and CK was expressed, protein L purified and assessed in the receptor assay. The resulting IC₅₀ was approximately 1nM.

25

2.3 TAR1-27 dimers

3 x 96 clones were picked from the round 2 selection encompassing all the libraries and selection conditions. 2ml supernatant preps were made for analysis in ELISA and 30 bioassays. Antigen ELISA gave 71 positive clones. The receptor assay of crude supernatants yielded 42 clones with inhibitory properties (TNF binding 0-60%). In the majority of cases inhibitory properties correlated with a strong ELISA signal. 42 clones

were sequenced, 39 of these have unique second dAb sequences. The 12 dimers that gave the best inhibitory properties were analysed further.

The 12 neutralising clones were expressed as 200ml supernatant preps and purified on 5 protein L. These were assessed by protein L and antigen ELISA, BIAcore and in the receptor assay. Strong positive ELISA signals were obtained in all cases. BIAcore analysis revealed all clones to have fast on and off rates. The off rates were improved compared to monomeric TAR1-27, however the off rate of TAR1-27 dimers was faster (Koff is approximately in the range of 10^{-1} and 10^{-2} M) than the TAR1-5 dimers examined 10 previously (Koff is approximately in the range of 10^{-3} - 10^{-4} M). The stability of the purified dimers was questioned and therefore in order to improve stability, the addition on 15 5%glycerol, 0.5% Triton X100 or 0.5% NP40 (Sigma) was included in the purification of 2 TAR1-27 dimers (d2 and d16). Addition of NP40 or Triton X100™ improved the yield of purified product approximately 2 fold. Both dimers were assessed in the receptor assay. TAR1-27d2 gave IC50 of ~30nM under all purification conditions. TAR1-27d16 showed no neutralisation effect when purified without the use of stabilising agents but gave an IC50 of ~50nM when purified under stabilising conditions. No further analysis 20 was conducted.

2.4 TAR2-5 dimers

3 x 96 clones were picked from the second round selections encompassing all the libraries and selection conditions. 2ml supernatant preps were made for analysis. Protein A and antigen ELISAs were conducted for each plate. 30 interesting clones were identified as having good off-rates by BIAcore (Koff ranges between 10^{-2} – 10^{-3} M). The clones were 25 sequenced and 13 unique dimers were identified by sequence analysis.

Table 1: TAR1-5 dimers

Dimer	Cell type	Purification	Protein Fraction	Elution conditions	Receptor/Cell assay
TAR1-5d1	HB2151	Protein L + FPLC	small dimeric species	0.1M glycine pH2.5	RA~30nM
TAR1-5d2	HB2151	Protein L + FPLC	small dimeric species	0.1M glycine pH2.5	RA~50nM

		FPLC	species	pH2.5	M
TAR1-5d3	HB2151	Protein L + FPLC	large dimeric species	0.1M glycine pH2.5	RA~300 nM
TAR1-5d4	HB2151	Protein L + FPLC	small dimeric species	0.1M glycine pH2.5	RA~3n M
TAR1-5d5	HB2151	Protein L + FPLC	large dimeric species	0.1M glycine pH2.5	RA~200 nM
TAR1-5d6	HB2151	Protein L + FPLC	Large dimeric species	0.1M glycine pH2.5	RA~100 nM

*note dimer 2 and dimer 3 have the same second dAb (called dAb2), however have different linker lengths (d2 = (Gly₄Ser)₃, d3 = (Gly₄Ser)₃). dAb1 is the partner dAb to dimers 1, 5 and 6. dAb3 is the partner dAb to dimer4. None of the partner dAbs neutralise alone. FPLC purification is by cation exchange unless otherwise stated. The optimal dimeric species for each dimer obtained by FPLC was determined in these assays.

10 **Table 2: TAR1-5-19 dimers**

Dimer	Cell type	Purification	Protein Fraction	Elution conditions	Receptor/ Cell assay
TAR1-5-19 d1	TOP10F'	Protein L	Total protein	0.1M glycine pH 2.0	RA~15 nM
TAR1-5-19 d2 (no stop codon)	TOP10F'	Protein L	Total protein	0.1M glycine pH 2.0 + 0.05%NP40	RA~2n M
TAR1-5-19d3 (no stop codon)	TOP10F'	Protein L	Total protein	0.1M glycine pH 2.5 + 0.05%NP40	RA~8n M
TAR1-5-19d4	TOP10F'	Protein L + FPLC	Purified fraction	0.1M glycine pH2.0	RA~2-5nM CA~12 nM
TAR1-5-19d5	TOP10F'	Protein L	Total protein	0.1M glycine pH2.0 + NP40	RA~8n M CA~10 nM
TAR1-5-19 d6	TOP10F'	Protein L	Total protein	0.1M glycine pH 2.0	RA~10 nM

Table 3: TAR1-5-19 homodimers

15

Dimer	Cell type	Purification	Protein Fraction	Elution conditions	Receptor/ Cell assay

homodimer						nM CA~30 nM
TAR1-5-19 homodimer	5U	HB2151	Protein L	Total protein	0.1M glycine pH2.5	RA~2n M CA~3n M
TAR1-5-19 homodimer	7U	HB2151	Protein L	Total protein	0.1M glycine pH2.5	RA~10 nM CA~15 nM
TAR1-5-19 hinge	cys	HB2151	Protein L + FPLC	FPLC purified dimer fraction	0.1M glycine pH2.5	RA~2n M
TAR1-5-19CH/ TAR1-5-19 CK		HB2151	Protein	Total protein	0.1M glycine pH2.5	RA~1n M

Table 4: TAR1-5/TAR1-5-19 Fabs

Dimer	Cell type	Purification	Protein Fraction	Elution conditions	Receptor/Cell assay
TAR1-5CH/ dAb1 CK	HB2151	Protein L	Total protein	0.1M citrate pH2.6	RA~90 nM
TAR1-5CH/ dAb2 CK	HB2151	Protein L	Total protein	0.1M glycine pH2.5	RA~30 nM CA~60 nM
dAb3CH/ TAR1-5CK	HB2151	Protein L	Total protein	0.1M citrate pH2.6	RA~10 0nM
TAR1-5-19CH/ dAb1 CK	HB2151	Protein L	Total protein	0.1M glycine pH2.0	RA~6n M
dAb1 CH/ TAR1-5-19CK	HB2151	Protein L	0.1M glycine pH2.0	Myc/flag	RA~6n M
TAR1-5-19CH/ dAb2 CK	HB2151	Protein L	Total protein	0.1M glycine pH2.0	RA~8n M CA~12 nM
TAR1-5-19CH/ dAb3CK	HB2151	Protein L	Total protein	0.1M glycine pH2.0	RA~3n M

PCR construction of TAR1-5-19CYS dimer

See example 8 describing the dAb trimer. The trimer protocol gives rise to a mixture of monomer, dimer and trimer.

5 Expression and purification of TAR1-5-19CYS dimer

The dimer was purified from the supernatant of the culture by capture on Protein L agarose as outlined in the example 8.

Separation of TAR1-5-19CYS monomer from the TAR1-5-19CYS dimer

10 Prior to cation exchange separation, the mixed monomer/dimer sample was buffer exchanged into 50 mM sodium acetate buffer pH 4.0 using a PD-10 column (Amersham Pharmacia), following the manufacturer's guidelines. The sample was then applied to a 1mL Resource S cation exchange column (Amersham Pharmacia), which had been pre-equilibrated with 50 mM sodium acetate pH 4.0. The monomer and dimer were separated
15 using the following salt gradient in 50 mM sodium acetate pH 4.0:

150 to 200 mM sodium chloride over 15 column volumes
200 to 450 mM sodium chloride over 10 column volumes
450 to 1000 mM sodium chloride over 15 column volumes

20 Fractions containing dimer only were identified using SDS-PAGE and then pooled and the pH increased to 8 by the addition of 1/5 volume of 1M Tris pH 8.0.

***In vitro* functional binding assay: TNF receptor assay and cell assay**

25 The affinity of the dimer for human TNF α was determined using the TNF receptor and cell assay. IC50 in the receptor assay was approximately 0.3-0.8 nM; ND50 in the cell assay was approximately 3-8 nM.

Other possible TAR1-5-19CYS dimer formats

30

PEG dimers and custom synthetic maleimide dimers

Nektar (Shearwater) offer a range of bi-maleimide PEGs [mPEG2-(MAL)2 or mPEG-(MAL)2] which would allow the monomer to be formatted as a dimer, with a small linker

separating the dAbs and both being linked to a PEG ranging in size from 5 to 40 kDa. It has been shown that the 5kDa mPEG-(MAL)2 (ie, [TAR1-5-19]-Cys-maleimide-PEG x 2, wherein the maleimides are linked together in the dimer) has an affinity in the TNF receptor assay of ~ 1-3 nM. Also the dimer can also be produced using TMEA (Tris[2-maleimidoethyl]amine) (Pierce Biotechnology) or other bi-functional linkers.

It is also possible to produce the disulphide dimer using a chemical coupling procedure using 2,2'-dithiodipyridine (Sigma Aldrich) and the reduced monomer.

10 *Addition of a polypeptide linker or hinge to the C-terminus of the dAb.*

A small linker, either $(\text{Gly}_4\text{Ser})_n$ where $n=1$ to 10, eg, 1, 2, 3, 4, 5, 6 or 7, an immunoglobulin (eg, IgG hinge region or random peptide sequence (eg, selected from a library of random peptide sequences) can be engineered between the dAb and the terminal cysteine residue. This can then be used to make dimers as outlined above.

15

Example 8

dAb trimerisation

20

Summary

For dAb trimerisation, a free cysteine is required at the C-terminus of the protein. The cysteine residue, once reduced to give the free thiol, can then be used to specifically couple the protein to a trimeric maleimide molecule, for example TMEA (Tris[2-maleimidoethyl]amine).

PCR construction of TAR1-5-19CYS

The following oligonucleotides were used to specifically PCR TAR1-5-19 with a *Sal*I and

30

*Bam*HI sites for cloning and also to introduce a C-terminal cysteine residue:

*Sal*I

Trp Ser Ala Ser Thr Asp* Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Val

35

1 TGG AGC GCG TCG ACG GAC ATC CAG ATG ACC CAG TCT CCA TCC TCT CTG TCT GCA TCT GTA
ACC TCG CGC AGC TGC CTG TAG GTC TAC TGG GTC AGA GGT AGG AGA GAC AGA CGT AGA CAT

Gly Asp Arg Val Thr Ile Thr Cys Arg Ala Ser Gln Ser Ile Asp Ser Tyr Leu His Trp

61 GGA GAC CGT GTC ACC ATC ACT TGC CGG GCA AGT CAG AGC ATT GAT AGT TAT TTA CAT TGG
CCT CTG GCA CAG TGG TAG TGA ACG GCC CGT TCA GTC TCG TAA CTA TCA ATA AAT GTA ACC

5 Tyr Gln Gln Lys Pro Gly Lys Ala Pro Lys Leu Leu Ile Tyr Ser Ala Ser Glu Leu Gln
121 TAC CAG CAG AAA CCA GGG AAA GCC CCT AAG CTC CTG ATC TAT AGT GCA TCC GAG TTG CAA
ATG GTC GTC TTT GGT CCC TTT CGG GGA TTC GAG GAC TAG ATA TCA CGT AGG CTC AAC GTT

10 Ser Gly Val Pro Ser Arg Phe Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile
181 AGT GGG GTC CCA TCA CGT TTC AGT GGC AGT GGA TCT GGG ACA GAT TTC ACT CTC ACC ATC
TCA CCC CAG GGT AGT GCA AAG TCA CCG TCA CCT AGA CCC TGT CTA AAG TGA GAG TGG TAG

15 Ser Ser Leu Gln Pro Glu Asp Phe Ala Thr Tyr Tyr Cys Gln Gln Val Val Trp Arg Pro
241 AGC AGT CTG CAA CCT GAA GAT TTT GCT ACG TAC TAC TGT CAA CAG GTT GTG TGG CGT CCT
TCG TCA GAC GTT GGA CTT CTA AAA CGA TGC ATG ATG ACA GTT GTC CAA CAC ACC GCA GGA

BamHI

20 Phe Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg Cys *** *** Gly Ser Gly
301 TTT ACG TTC GGC CAA GGG ACC AAG GTG GAA ATC AAA CGG TGC TAA TAA GGA TCC GGC
AAA TGC AAG CCG GTT CCC TGG TTC CAC CTT TAG TTT GCC ACG ATT ATT CCT AGG CCG

(* start of TAR1-5-19CYS sequence)

Forward primer

25 5'-TGGAGCGCGTCGACGGACATCCAGATGACCCAGTCTCCA-3'

Reverse primer

30 5'-TTAGCAGCCGGATCCTTATTAGCACCGTTGATTCCAC-3'

The PCR reaction (50µL volume) was set up as follows: 200µM dNTPs, 0.4µM of each primer, 5 µL of 10x *Pfu*Turbo buffer (Stratagene), 100 ng of template plasmid (encoding TAR1-5-19), 1µL of *Pfu*Turbo enzyme (Stratagene) and the volume adjusted to 50µL using sterile water. The following PCR conditions were used: initial denaturing step 94 °C for 2 mins, then 25 cycles of 94 °C for 30 secs, 64 °C for 30 sec and 72 °C for 30 sec. A final extension step was also included of 72 °C for 5 mins. The PCR product was purified and digested with *Sal*I and *Bam*HI and ligated into the vector which had also been cut with the same restriction enzymes. Correct clones were verified by DNA sequencing.

Expression and purification of TAR1-5-19CYS

TAR1-5-19CYS vector was transformed into BL21 (DE3) pLysS chemically competent cells (Novagen) following the manufacturer's protocol. Cells carrying the dAb plasmid were selected for using 100 µg/mL carbenicillin and 37 µg/mL chloramphenicol. Cultures 5 were set up in 2L baffled flasks containing 500 mL of terrific broth (Sigma-Aldrich), 100 µg/mL carbenicillin and 37 µg/mL chloramphenicol. The cultures were grown at 30 °C at 200rpm to an O.D.600 of 1-1.5 and then induced with 1mM IPTG (isopropyl-beta-D-thiogalactopyranoside, from Melford Laboratories). The expression of the dAb was allowed to continue for 12-16 hrs at 30 °C. It was found that most of the dAb was present 10 in the culture media. Therefore, the cells were separated from the media by centrifugation (8,000xg for 30 mins), and the supernatant used to purify the dAb. Per litre of supernatant, 30 mL of Protein L agarose (Affitech) was added and the dAb allowed to batch bind with stirring for 2 hours. The resin was then allowed to settle under gravity for a further hour before the supernatant was siphoned off. The agarose was then packed into 15 a XK 50 column (Amersham Pharmacia) and was washed with 10 column volumes of PBS. The bound dAb was eluted with 100 mM glycine pH 2.0 and protein containing fractions were then neutralized by the addition of 1/5 volume of 1 M Tris pH 8.0. Per litre of culture supernatant 20 mg of pure protein was isolated, which contained a 50:50 ratio of monomer to dimer.

20

Trimerisation of TAR1-5-19CYS

2.5 ml of 100 µM TAR1-5-19CYS was reduce with 5 mM dithiothreitol and left at room temperature for 20 minutes. The sample was then buffer exchanged using a PD-10 column (Amersham Pharmacia). The column had been pre-equilibrated with 5 mM 25 EDTA, 50 mM sodium phosphate pH 6.5, and the sample applied and eluted following the manufactures guidelines. The sample was placed on ice until required. TMEA (Tris[2-maleimidioethyl]amine) was purchased from Pierce Biotechnology. A 20 mM stock solution of TMEA was made in 100% DMSO (dimethyl sulphoxide). It was found that a concentration of TMEA greater than 3:1 (molar ratio of dAb:TMEA) caused the rapid 30 precipitation and cross-linking of the protein. Also the rate of precipitation and cross-linking was greater as the pH increased. Therefore using 100 µM reduced TAR1-5-19CYS, 25 µM TMEA was added to trimerise the protein and the reaction allowed to proceed at room temperature for two hours. It was found that the addition of additives

such as glycerol or ethylene glycol to 20% (v/v), significantly reduced the precipitation of the trimer as the coupling reaction proceeded. After coupling, SDS-PAGE analysis showed the presence of monomer, dimer and trimer in solution.

5 **Purification of the trimeric TAR1-5-19CYS**

40 μ L of 40% glacial acetic acid was added per mL of the TMEA-TAR1-5-19cys reaction to reduce the pH to ~4. The sample was then applied to a 1mL Resource S cation exchange column (Amersham Pharmacia), which had been pre-equilibrated with 50 mM sodium acetate pH 4.0. The dimer and trimer were partially separated using a salt gradient of 340 to 450 mM Sodium chloride, 50 mM sodium acetate pH 4.0 over 30 column volumes. Fractions containing trimer only were identified using SDS-PAGE and then pooled and the pH increased to 8 by the addition of 1/5 volume of 1M Tris pH 8.0. To prevent precipitation of the trimer during concentration steps (using 5K cut off Viva spin concentrators; Vivascience), 10% glycerol was added to the sample.

15

***In vitro* functional binding assay: TNF receptor assay and cell assay**

The affinity of the trimer for human TNF α was determined using the TNF receptor and cell assay. IC50 in the receptor assay was 0.3nM; ND50 in the cell assay was in the range of 3 to 10nM (eg, 3nM).

20

Other possible TAR1-5-19CYS trimer formats

TAR1-5-19CYS may also be formatted into a trimer using the following reagents:

PEG trimers and custom synthetic maleimide trimers

25 Nektar (Shearwater) offer a range of multi arm PEGs, which can be chemically modified at the terminal end of the PEG. Therefore using a PEG trimer with a maleimide functional group at the end of each arm would allow the trimerisation of the dAb in a manner similar to that outlined above using TMEA. The PEG may also have the advantage in increasing the solubility of the trimer thus preventing the problem of aggregation. Thus, one could produce a dAb trimer in which each dAb has a C-terminal cysteine that is linked to a maleimide functional group, the maleimide functional groups being linked to a PEG trimer.

Addition of a polypeptide linker or hinge to the C-terminus of the dAb

A small linker, either $(\text{Gly}_4\text{Ser})_n$ where $n=1$ to 10 , eg, $1, 2, 3, 4, 5, 6$ or 7 , an immunoglobulin (eg, IgG hinge region or random peptide sequence (eg, selected from a library of random peptide sequences) could be engineered between the dAb and the terminal cysteine residue. When used to make multimers (eg, dimers or trimers), this again would introduce a greater degree of flexibility and distance between the individual monomers, which may improve the binding characteristics to the target, eg a multisubunit target such as human TNF α .

10 **Example 9.****Selection of a collection of single domain antibodies (dAbs) directed against human serum albumin (HSA) and mouse serum albumin (MSA).**

15 This example explains a method for making a single domain antibody (dAb) directed against serum albumin. Selection of dAbs against both mouse serum albumin (MSA) and human serum albumin (HSA) is described. Three human phage display antibody libraries were used in this experiment, each based on a single human framework for V_H (see Figure 13: sequence of dummy V_H based on V3-23/DP47 and JH4b) or V_K (see Figure 20 15: sequence of dummy V_K based on o12/o2/DPK9 and Jk1) with side chain diversity encoded by NNK codons incorporated in complementarity determining regions (CDR1, CDR2 and CDR3).

Library 1 (V_H):

25 Diversity at positions: H30, H31, H33, H35, H50, H52, H52a, H53, H55, H56, H58, H95, H97, H98.

Library size: 6.2×10^9

Library 2 (V_H):

30 Diversity at positions: H30, H31, H33, H35, H50, H52, H52a, H53, H55, H56, H58, H95, H97, H98, H99, H100, H100a, H100b.

Library size: 4.3×10^9

Library 3 (V κ):

Diversity at positions: L30, L31, L32, L34, L50, L53, L91, L92, L93, L94, L96

Library size: 2 x 10⁹

5 The V_H and V κ libraries have been preselected for binding to generic ligands protein A and protein L respectively so that the majority of clones in the unselected libraries are functional. The sizes of the libraries shown above correspond to the sizes after preselection.

10 Two rounds of selection were performed on serum albumin using each of the libraries separately. For each selection, antigen was coated on immunotube (nunc) in 4ml of PBS at a concentration of 100 μ g/ml. In the first round of selection, each of the three libraries was panned separately against HSA (Sigma) and MSA (Sigma). In the second round of selection, phage from each of the six first round selections was panned against (i) the same antigen again (eg 1st round MSA, 2nd round MSA) and (ii) against the reciprocal antigen (eg 1st round MSA, 2nd round HSA) resulting in a total of twelve 2nd round selections. In each case, after the second round of selection 48 clones were tested for binding to HSA and MSA. Soluble dAb fragments were produced as described for scFv fragments by Harrison *et al*, Methods Enzymol. 1996;267:83-109 and standard ELISA protocol was followed (Hoogenboom *et al*. (1991) Nucleic Acids Res., 19: 4133) except that 2% tween PBS was used as a blocking buffer and bound dAbs were detected with either protein L-HRP (Sigma) (for the V κ s) and protein A -HRP (Amersham Pharmacia Biotech) (for the V_Hs).

15

20

25 dAbs that gave a signal above background indicating binding to MSA, HSA or both were tested in ELISA insoluble form for binding to plastic alone but all were specific for serum albumin. Clones were then sequenced (see table below) revealing that 21 unique dAb sequences had been identified. The minimum similarity (at the amino acid level) between the V κ dAb clones selected was 86.25% ((69/80)x100; the result when all the diversified residues are different, eg clones 24 and 34). The minimum similarity between the V_H dAb clones selected was 94 % ((127/136)x100).

30

Next, the serum albumin binding dAbs were tested for their ability to capture biotinylated antigen from solution. ELISA protocol (as above) was followed except that ELISA plate was coated with 1 μ g/ml protein L (for the V κ clones) and 1 μ g/ml protein A (for the V H clones). Soluble dAb was captured from solution as in the protocol and detection was with biotinylated MSA or HSA and streptavidin HRP. The biotinylated MSA and HSA had been prepared according to the manufacturer's instructions, with the aim of achieving an average of 2 biotins per serum albumin molecule. Twenty four clones were identified that captured biotinylated MSA from solution in the ELISA. Two of these (clones 2 and 38 below) also captured biotinylated HSA. Next, the dAbs were tested for their ability to bind MSA coated on a CM5 biacore chip. Eight clones were found that bound MSA on the biacore.

dAb (all capture biotinylated MSA)	H or κ	CDR1	CDR2	CDR3	Binds MSA in biacore?	Captures biotinylated HSA?
V κ library 3 template						
(dummy)	K	XXXLX	XASXLQS	QQXXXXPXT		
2, 4, 7, 41,	K	SSYLN	RASPLQS	QQTYSVPPT	✓ all 4 bind	
38, 54	K	SSYLN	RASPLQS	QQTYRIPPT	✓ both bind	
46, 47, 52, 56	K	FKSLK	NASYLQS	QQVYWPVT		
13, 15	K	YYHLK	KASTLQS	QQVRKVPRT		
30, 35	K	RRYLK	QASVLQS	QQGLYPPIT		
19,	K	YNWLK	RASSLQS	QQNVVIPRT		
22,	K	LWHLR	HASLLQS	QQSAVYPKT		
23,	K	FRYLA	HASHLQS	QQRLLYPKT		
24,	K	FYHLA	PASKLQS	QQRARWPRT		
31,	K	IWHLN	RASRLQS	QQVARVPRT		
33,	K	YRYLR	KASSLQS	QQYVGYPRT		
34,	K	LKYLK	NASHLQS	QQTTYYPIT		
53,	K	LRYLR	KASWLQS	QQVLYYPQT		
11,	K	LRSLK	AASRLQS	QQVYWPAT	✓	
12,	K	FRHLK	AASRLQS	QQVALYPKT	✓	
17,	K	RKYLR	TASSLQS	QQNLFWPRT	✓	
18,	K	RRYLN	AASSLQS	QQMLFYPKT	✓	
16, 21	K	IKHLK	GASRLQS	QQGARWPQT	✓	
25, 26	K	YYHLK	KASTLQS	QQVRKVPRT	✓	
27,	K	YKHLK	NASHLQS	QQVGRYPKT	✓	
55,	K	FKSLK	NASYLQS	QQVYWPVT	✓	
V H library 1 (and 2) template						
(dummy)	H	XXYXXXX	XIXXXGXXTXYADSVKG	XXXX (XXXX) FDY		
8, 10	H	WVYQMD	SISAFGAKTLYADSVKG	LSGKFDY		
36,	H	WSYQMT	SISSFGSSTLYADSVKG	GRDHNYSLFDY		

In all cases the frameworks were identical to the frameworks in the corresponding dummy sequence, with diversity in the CDRs as indicated in the table above.

Of the eight clones that bound MSA on the biacore, two clones that are highly expressed
5 in *E. coli* (clones MSA16 and MSA26) were chosen for further study (see example 10).
Full nucleotide and amino acid sequences for MSA16 and 26 are given in figure 16.

Example 10.

10 **Determination of affinity and serum half-life in mouse of MSA binding dAbs MSA16 and MSA26.**

dAbs MSA16 and MSA26 were expressed in the periplasm of *E. coli* and purified using
batch absorbtion to protein L-agarose affinity resin (Affitech, Norway) followed by
15 elution with glycine at pH 2.2. The purified dAbs were then analysed by inhibition
biacore to determine K_d . Briefly, purified MSA16 and MSA26 were tested to determine
the concentration of dAb required to achieve 200RUs of response on a biacore CM5 chip
coated with a high density of MSA. Once the required concentrations of dAb had been
determined, MSA antigen at a range of concentrations around the expected K_d was
20 premixed with the dAb and incubated overnight. Binding to the MSA coated biacore chip
of dAb in each of the premixes was then measured at a high flow-rate of 30 μ l/minute.
The resulting curves were used to create Klotz plots, which gave an estimated K_d of
200nM for MSA16 and 70nM for MSA 26 (Figure 17 A & B).

25 Next, clones MSA16 and MSA26 were cloned into an expression vector with the HA tag
(nucleic acid sequence: TATCCTTATGATGTTCTGATTATGCA and amino acid
sequence: YPYDVPDYA) and 2-10 mg quantities were expressed in *E. coli* and purified
from the supernatant with protein L-agarose affinity resin (Affitech, Norway) and eluted
with glycine at pH2.2. Serum half life of the dAbs was determined in mouse. MSA26
30 and MSA16 were dosed as single i.v. injections at approx 1.5mg/kg into CD1 mice.
Analysis of serum levels was by goat anti-HA (Abcam, UK) capture and protein L-HRP
(invitrogen) detection ELISA which was blocked with 4% Marvel. Washing was with
0.05% tween PBS. Standard curves of known concentrations of dAb were set up in the

presence of 1xmouse serum to ensure comparability with the test samples. Modelling with a 2 compartment model showed MSA-26 had a $t_{1/2\alpha}$ of 0.16hr, a $t_{1/2\beta}$ of 14.5hr and an area under the curve (AUC) of 465hr.mg/ml (data not shown) and MSA-16 had a $t_{1/2\alpha}$ of 0.98hr, a $t_{1/2\beta}$ of 36.5hr and an AUC of 913hr.mg/ml (figure 18). Both anti-MSA 5 clones had considerably lengthened half life compared with HEL4 (an anti-hen egg white lysozyme dAb) which had a $t_{1/2\alpha}$ of 0.06hr, and a $t_{1/2\beta}$ of 0.34hr.

Example 11.

10 Creation of V_H - V_H and V_K - V_K dual specific Fab like fragments

This example describes a method for making V_H - V_H and V_K - V_K dual specifics as Fab like fragments. Before constructing each of the Fab like fragments described, dAbs that bind to targets of choice were first selected from dAb libraries similar to those described 15 in example 9. A V_H dAb, HEL4, that binds to hen egg lysozyme (Sigma) was isolated and a second V_H dAb (TAR2h-5) that binds to TNF α receptor (R and D systems) was also isolated. The sequences of these are given in the sequence listing. A V_K dAb that binds TNF α (TAR1-5-19) was isolated by selection and affinity maturation and the sequence is also set forth in the sequence listing. A second V_K dAb (MSA 26) described in example 9 20 whose sequence is in figure 17B was also used in these experiments.

DNA from expression vectors containing the four dAbs described above was digested with enzymes SalI and NotI to excise the DNA coding for the dAb. A band of the expected size (300-400bp) was purified by running the digest on an agarose gel and 25 excising the band, followed by gel purification using the Qiagen gel purification kit (Qiagen, UK). The DNA coding for the dAbs was then inserted into either the C_H or C_K vectors (Figs 8 and 9) as indicated in the table below.

dAb	Target antigen	dAb V_H or dAb V_K	Inserted into vector	tag (C terminal)	Antibiotic resistance
HEL4	Hen egg lysozyme	V_H	C_H	myc	Chloramphenicol
TAR2-5	TNF receptor	V_H	C_K	flag	Ampicillin
TAR1-5-19	TNF α	V_K	C_H	myc	Chloramphenicol
MSA 26	Mouse serum albumin	V_K	C_K	flag	Ampicillin

The $V_H C_H$ and $V_H C_K$ constructs were cotransformed into HB2151 cells. Separately, the $V_K C_H$ and $V_K C_K$ constructs were cotransformed into HB2151 cells. Cultures of each of the cotransformed cell lines were grown overnight (in 2xTy containing 5% glucose, 10 μ g/ml chloramphenicol and 100 μ g/ml ampicillin to maintain antibiotic selection for both C_H and C_K plasmids). The overnight cultures were used to inoculate fresh media (2xTy, 10 μ g/ml chloramphenicol and 100 μ g/ml ampicillin) and grown to OD 0.7-0.9 before induction by the addition of IPTG to express their C_H and C_K constructs. Expressed Fab like fragment was then purified from the periplasm by protein A purification (for the contrtransformed $V_H C_H$ and $V_H C_K$) and MSA affinity resin purification (for the contrtransformed $V_K C_H$ and $V_K C_K$).

V_H - V_H dual specific

Expression of the $V_H C_H$ and $V_H C_K$ dual specific was tested by running the protein on a gel. The gel was blotted and a band the expected size for the Fab fragment could be detected on the Western blot via both the myc tag and the flag tag, indicating that both the $V_H C_H$ and $V_H C_K$ parts of the Fab like fragment were present. Next, in order to determine whether the two halves of the dual specific were present in the same Fab-like fragment, an ELISA plate was coated overnight at 4°C with 100 μ l per well of hen egg lysozyme (HEL) at 3 mg/ml in sodium bicarbonate buffer. The plate was then blocked (as described in example 1) with 2% tween PBS followed by incubation with the $V_H C_H$ / $V_H C_K$ dual specific Fab like fragment. Detection of binding of the dual specific to the HEL was via the non cognate chain using 9e10 (a monoclonal antibody that binds the myc tag, Roche) and anti mouse IgG-HRP (Amersham Pharmacia Biotech). The signal for the $V_H C_H$ / $V_H C_K$ dual specific Fab like fragment was 0.154 compared to a background signal of 0.069 for the $V_H C_K$ chain expressed alone. This demonstrates that the Fab like fragment has binding specificity for target antigen.

V_K - V_K dual specific

After purifying the contrtransformed $V_K C_H$ and $V_K C_K$ dual specific Fab like fragment on an MSA affinity resin, the resulting protein was used to probe an ELISA plate coated with 1 μ g/ml TNF α and an ELISA plate coated with 10 μ g/ml MSA. As predicted, there was signal above background when detected with protein L-HRP on bot ELISA plates (data not shown). This indicated that the fraction of protein able to bind to MSA (and therefore

purified on the MSA affinity column) was also able to bind TNF α in a subsequent ELISA, confirming the dual specificity of the antibody fragment. This fraction of protein was then used for two subsequent experiments. Firstly, an ELISA plate coated with 1 μ g/ml TNF α was probed with dual specific V κ C_H and V κ C κ Fab like fragment and 5 also with a control TNF α binding dAb at a concentration calculated to give a similar signal on the ELISA. Both the dual specific and control dAb were used to probe the ELISA plate in the presence and in the absence of 2mg/ml MSA. The signal in the dual specific well was reduced by more than 50% but the signal in the dAb well was not reduced at all (see figure 19a). The same protein was also put into the receptor assay with 10 and without MSA and competition by MSA was also shown (see figure 19c). This demonstrates that binding of MSA to the dual specific is competitive with binding to TNF α .

15 **Example 12.**

Creation of a V κ - V κ dual specific cys bonded dual specific with specificity for mouse serum albumin and TNF α

20 This example describes a method for making a dual specific antibody fragment specific for both mouse serum albumin and TNF α by chemical coupling via a disulphide bond. Both MSA16 (from example 1) and TAR1-5-19 dAbs were recloned into a pET based vector with a C terminal cysteine and no tags. The two dAbs were expressed at 4-10 mg levels and purified from the supernatant using protein L-agarose affinity resin (Affitiech, 25 Norway). The cysteine tagged dAbs were then reduced with dithiothreitol. The TAR1-5-19 dAb was then coupled with dithiobipyridine to block reformation of disulphide bonds resulting in the formation of PEP 1-5-19 homodimers. The two different dAbs were then mixed at pH 6.5 to promote disulphide bond formation and the generation of TAR1-5-19, MSA16 cys bonded heterodimers. This method for producing conjugates of two unlike 30 proteins was originally described by King *et al.* (King TP, Li Y Kochoumian L Biochemistry. 1978 vol17:1499-506 Preparation of protein conjugates via intermolecular disulfide bond formation.) Heterodimers were separated from monomeric species by cation exchange. Separation was confirmed by the presence of a band of the expected

size on a SDS gel. The resulting heterodimeric species was tested in the TNF receptor assay and found to have an IC50 for neutralising TNF of approximately 18 nM. Next, the receptor assay was repeated with a constant concentration of heterodimer (18nM) and a dilution series of MSA and HSA. The presence of HSA at a range of concentrations (up to 2 mg/ml) did not cause a reduction in the ability of the dimer to inhibit TNF α . However, the addition of MSA caused a dose dependant reduction in the ability of the dimer to inhibit TNF α (figure 20). This demonstrates that MSA and TNF α compete for binding to the cys bonded TARI-5-19, MSA16 dimer.

10 **Data Summary**

A summary of data obtained in the experiments set forth in the foregoing examples is set forth in Annex 4.

15 All publications mentioned in the present specification, and references cited in said publications, are herein incorporated by reference. Various modifications and variations of the described methods and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such 20 specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Annex 1; polypeptides which enhance half-life *in vivo*.

Alpha-1 Glycoprotein (Orosomucoid) (AAG)

Alpha-1 Antichyromotrypsin (ACT)

5 Alpha-1 Antitrypsin (AAT)

Alpha-1 Microglobulin (Protein HC) (AIM)

Alpha-2 Macroglobulin (A2M)

Antithrombin III (AT III)

Apolipoprotein A-1 (Apo A-1)

10 Apolipoprotein B (Apo B)

Beta-2-microglobulin (B2M)

Ceruloplasmin (Cp)

Complement Component (C3)

Complement Component (C4)

15 C1 Esterase Inhibitor (C1 INH)

C-Reactive Protein (CRP)

Cystatin C (Cys C)

Ferritin (FER)

Fibrinogen (FIB)

20 Fibronectin (FN)

Haptoglobin (Hp)

Hemopexin (HPX)

Immunoglobulin A (IgA)

Immunoglobulin D (IgD)

25 Immunoglobulin E (IgE)

Immunoglobulin G (IgG)

Immunoglobulin M (IgM)

Immunoglobulin Light Chains (kappa/lambda)

Lipoprotein(a) [Lp(a)]

30 Mannose-bindign protein (MBP)

Myoglobin (Myo)

Plasminogen (PSM)

Prealbumin (Transthyretin) (PAL)

- Retinol-binding protein (RBP)
- Rheumatoid Factor (RF)
- Serum Amyloid A (SAA)
- Soluble Tranferrin Receptor (sTfR)
- 5 Transferrin (Tf)

Annex 2

Pairing	Therapeutic relevant references.
TNF ALPHA/TGF- β	<ul style="list-style-type: none"> TGF-β and TNF when injected into the ankle joint of collagen induced arthritis model significantly enhanced joint inflammation. In non-collagen challenged mice there was no effect.
TNF ALPHA/IL-1	<ul style="list-style-type: none"> TNF and IL-1 synergize in the pathology of uveitis. TNF and IL-1 synergize in the pathology of malaria (hypoglycaemia, NO). TNF and IL-1 synergize in the induction of polymorphonuclear (PMN) cells migration in inflammation. IL-1 and TNF synergize to induce PMN infiltration into the peritoneum. IL-1 and TNF synergize to induce the secretion of IL-1 by endothelial cells. Important in inflammation. IL-1 or TNF alone induced some cellular infiltration into knee synovium. IL-1 induced PMNs, TNF – monocytes. Together they induced a more severe infiltration due to increased PMNs. Circulating myocardial depressant substance (present in sepsis) is low levels of IL-1 and TNFacting synergistically.
TNF ALPHA/IL-2	<ul style="list-style-type: none"> Most relating to synergistic activation of killer T-cells.
TNF ALPHA/IL-3	<ul style="list-style-type: none"> Synergy of interleukin 3 and tumor necrosis factor alpha in stimulating clonal growth of acute myelogenous leukemia blasts is the result of induction of secondary hematopoietic cytokines by tumor necrosis factor alpha. Cancer Res. 1992 Apr 15;52(8):2197-201.
TNF ALPHA/IL-4	<ul style="list-style-type: none"> IL-4 and TNF synergize to induce VCAM expression on endothelial cells. Implied to have a role in asthma. Same for synovium – implicated in RA. TNF and IL-4 synergize to induce IL-6 expression in keratinocytes. Sustained elevated levels of VCAM-1 in cultured fibroblast-like synoviocytes can be achieved by TNF-alpha in combination with either IL-4 or IL-13 through increased mRNA stability. Am J Pathol. 1999 Apr;154(4):1149-58
TNF ALPHA/IL-5	<ul style="list-style-type: none"> Relationship between the tumor necrosis factor system and the serum interleukin-4, interleukin-5, interleukin-8, eosinophil cationic protein, and immunoglobulin E levels in the bronchial hyperreactivity of adults and their children. Allergy Asthma Proc. 2003 Mar-Apr;24(2):111-8.
TNF ALPHA/IL-6	<ul style="list-style-type: none"> TNF and IL-6 are potent growth factors for OH-2, a novel human myeloma cell line. Eur J Haematol. 1994 Jul;53(1):31-7.
TNF ALPHA/IL-8	<ul style="list-style-type: none"> TNF and IL-8 synergized with PMNs to activate platelets. Implicated in Acute Respiratory Distress Syndrome. See IL-5/TNF (asthma). Synergism between interleukin-8 and tumor necrosis factor-alpha for neutrophil-mediated platelet activation. Eur Cytokine Netw. 1994 Sep-Oct;5(5):455-60. (adult respiratory distress syndrome (ARDS))
TNF ALPHA/IL-9	
TNF ALPHA/IL-10	<ul style="list-style-type: none"> IL-10 induces and synergizes with TNF in the induction of HIV expression in chronically infected T-cells.
TNF ALPHA/IL-11	<ul style="list-style-type: none"> Cytokines synergistically induce osteoclast differentiation: support by immortalized or normal calvarial cells. Am J Physiol Cell Physiol. 2002 Sep;283(3):C679-87. (Bone loss)
TNF ALPHA/IL-12	

TNF ALPHA/IL-13	<ul style="list-style-type: none"> Sustained elevated levels of VCAM-1 in cultured fibroblast-like synoviocytes can be achieved by TNF-alpha in combination with either IL-4 or IL-13 through increased mRNA stability. <i>Am J Pathol.</i> 1999 Apr;154(4):1149-58. Interleukin-13 and tumour necrosis factor-alpha synergistically induce eotaxin production in human nasal fibroblasts. <i>Clin Exp Allergy.</i> 2000 Mar;30(3):348-55. Interleukin-13 and tumour necrosis factor-alpha synergistically induce eotaxin production in human nasal fibroblasts. <i>Clin Exp Allergy.</i> 2000 Mar;30(3):348-55 (allergic inflammation) Implications of serum TNF-beta and IL-13 in the treatment response of childhood nephrotic syndrome. <i>Cytokine.</i> 2003 Feb 7;21(3):155-9.
TNF ALPHA/IL-14	<ul style="list-style-type: none"> Effects of inhaled tumour necrosis factor alpha in subjects with mild asthma. <i>Thorax.</i> 2002 Sep;57(9):774-8.
TNF ALPHA/IL-15	<ul style="list-style-type: none"> Effects of inhaled tumour necrosis factor alpha in subjects with mild asthma. <i>Thorax.</i> 2002 Sep;57(9):774-8.
TNF ALPHA/IL-16	<ul style="list-style-type: none"> Tumor necrosis factor-alpha-induced synthesis of interleukin-16 in airway epithelial cells: priming for serotonin stimulation. <i>Am J Respir Cell Mol Biol.</i> 2003 Mar;28(3):354-62. (airway inflammation) Correlation of circulating interleukin 16 with proinflammatory cytokines in patients with rheumatoid arthritis. <i>Rheumatology (Oxford).</i> 2001 Apr;40(4):474-5. No abstract available. Interleukin 16 is up-regulated in Crohn's disease and participates in TNBS colitis in mice. <i>Gastroenterology.</i> 2000 Oct;119(4):972-82.
TNF ALPHA/IL-17	<ul style="list-style-type: none"> Inhibition of interleukin-17 prevents the development of arthritis in vaccinated mice challenged with <i>Borrelia burgdorferi</i>. <i>Infect Immun.</i> 2003 Jun;71(6):3437-42. Interleukin 17 synergises with tumour necrosis factor alpha to induce cartilage destruction <i>in vitro</i>. <i>Ann Rheum Dis.</i> 2002 Oct;61(10):870-6. A role of GM-CSF in the accumulation of neutrophils in the airways caused by IL-17 and TNF-alpha. <i>Eur Respir J.</i> 2003 Mar;21(3):387-93. (Airway inflammation) Abstract Interleukin-1, tumor necrosis factor alpha, and interleukin-17 synergistically up-regulate nitric oxide and prostaglandin E2 production in explants of human osteoarthritic knee menisci. <i>Arthritis Rheum.</i> 2001 Sep;44(9):2078-83.
TNF ALPHA/IL-18	<ul style="list-style-type: none"> Association of interleukin-18 expression with enhanced levels of both interleukin-1beta and tumor necrosis factor alpha in knee synovial tissue of patients with rheumatoid arthritis. <i>Arthritis Rheum.</i> 2003 Feb;48(2):339-47. Abstract Elevated levels of interleukin-18 and tumor necrosis factor-alpha in serum of patients with type 2 diabetes mellitus: relationship with diabetic nephropathy. <i>Metabolism.</i> 2003 May;52(5):605-8.
TNF ALPHA/IL-19	<ul style="list-style-type: none"> Abstract IL-19 induces production of IL-6 and TNF-alpha and results in cell apoptosis through TNF-alpha. <i>J Immunol.</i> 2002 Oct 15;169(8):4288-97.
TNF ALPHA/IL-20	<ul style="list-style-type: none"> Abstract Cytokines: IL-20 - a new effector in skin inflammation. <i>Curr Biol.</i> 2001 Jul 10;11(13):R531-4
TNF ALPHA/Complement	<ul style="list-style-type: none"> Inflammation and coagulation: implications for the septic patient. <i>Clin Infect Dis.</i> 2003 May 15;36(10):1259-65. Epub 2003 May 08. Review.
TNF ALPHA/IFN- γ	<ul style="list-style-type: none"> MHC induction in the brain. Synergize in anti-viral response/IFN-β induction. Neutrophil activation/ respiratory burst. Endothelial cell activation Toxicities noted when patients treated with TNF/IFN-γ as anti-viral therapy

	<ul style="list-style-type: none"> • Fractalkine expression by human astrocytes. • Many papers on inflammatory responses – i.e. LPS, also macrophage activation. • Anti-TNF and anti-IFN-γ synergize to protect mice from lethal endotoxemia.
TGF- β /IL-1	<ul style="list-style-type: none"> • Prostaglandin synthesis by osteoblasts • IL-6 production by intestinal epithelial cells (inflammation model) • Stimulates IL-11 and IL-6 in lung fibroblasts (inflammation model) • IL-6 and IL-8 production in the retina
TGF- β /IL-6	<ul style="list-style-type: none"> • Chondrocarcoma proliferation
IL-1/IL-2	<ul style="list-style-type: none"> • B-cell activation • LAK cell activation • T-cell activation • IL-1 synergy with IL-2 in the generation of lymphokine activated killer cells is mediated by TNF-alpha and beta (lymphotoxin). <i>Cytokine</i>. 1992 Nov;4(6):479-87.
IL-1/IL-3	
IL-1/IL-4	<ul style="list-style-type: none"> • B-cell activation • IL-4 induces IL-1 expression in endothelial cell activation.
IL-1/IL-5	
IL-1/IL-6	<ul style="list-style-type: none"> • B cell activation • T cell activation (can replace accessory cells) • IL-1 induces IL-6 expression • C3 and serum amyloid expression (acute phase response) • HIV expression • Cartilage collagen breakdown.
IL-1/IL-7	<ul style="list-style-type: none"> • IL-7 is requisite for IL-1-induced thymocyte proliferation. Involvement of IL-7 in the synergistic effects of granulocyte-macrophage colony-stimulating factor or tumor necrosis factor with IL-1. <i>J Immunol</i>. 1992 Jan 1;148(1):99-105.
IL-1/IL-8	
IL-1/IL-10	
IL-1/IL-11	<ul style="list-style-type: none"> • Cytokines synergistically induce osteoclast differentiation: support by immortalized or normal calvarial cells. <i>Am J Physiol Cell Physiol</i>. 2002 Sep;283(3):C679-87. (Bone loss)
IL-1/IL-16	<ul style="list-style-type: none"> • Correlation of circulating interleukin 16 with proinflammatory cytokines in patients with rheumatoid arthritis. <i>Rheumatology</i> (Oxford). 2001 Apr;40(4):474-5. No abstract available.
IL-1/IL-17	<ul style="list-style-type: none"> • Inhibition of interleukin-17 prevents the development of arthritis in vaccinated mice challenged with <i>Borrelia burgdorferi</i>. <i>Infect Immun</i>. 2003 Jun;71(6):3437-42. • Contribution of interleukin 17 to human cartilage degradation and synovial inflammation in osteoarthritis. <i>Osteoarthritis Cartilage</i>. 2002 Oct;10(10):799-807. • Abstract Interleukin-1, tumor necrosis factor alpha, and interleukin-17 synergistically up-regulate nitric oxide and prostaglandin E2 production in explants of human osteoarthritic knee menisci. <i>Arthritis Rheum</i>. 2001 Sep;44(9):2078-83.
IL-1/IL-18	<ul style="list-style-type: none"> • Association of interleukin-18 expression with enhanced levels of both interleukin-1beta and tumor necrosis factor alpha in knee synovial tissue of patients with rheumatoid arthritis. <i>Arthritis Rheum</i>. 2003 Feb;48(2):339-47.
IL-1/IFN- γ	
IL-2/IL-3	<ul style="list-style-type: none"> • T-cell proliferation • B cell proliferation

IL-2/IL-4	<ul style="list-style-type: none"> • B-cell proliferation • T-cell proliferation • (selectively inducing activation of CD8 and NK lymphocytes) IL-2R beta agonist P1-30 acts in synergy with IL-2, IL-4, IL-9, and IL-15: biological and molecular effects. <i>J Immunol.</i> 2000 Oct 15;165(8): 4312-8.
IL-2/IL-5	<ul style="list-style-type: none"> • B-cell proliferation/ Ig secretion • IL-5 induces IL-2 receptors on B-cells
IL-2/IL-6	<ul style="list-style-type: none"> • Development of cytotoxic T-cells
IL-2/IL-7	
IL-2/IL-9	<ul style="list-style-type: none"> • See IL-2/IL-4 (NK-cells)
IL-2/IL-10	<ul style="list-style-type: none"> • B-cell activation
IL-2/IL-12	<ul style="list-style-type: none"> • IL-12 synergizes with IL-2 to induce lymphokine-activated cytotoxicity and perforin and granzyme gene expression in fresh human NK cells. <i>Cell Immunol.</i> 1995 Oct 1;165(1):33-43. (T-cell activation)
IL-2/IL-15	<ul style="list-style-type: none"> • See IL-2/IL-4 (NK cells) • (T cell activation and proliferation) IL-15 and IL-2: a matter of life and death for T cells <i>in vivo</i>. <i>Nat Med.</i> 2001 Jan;7(1):114-8.
IL-2/IL-16	<ul style="list-style-type: none"> • Synergistic activation of CD4+ T cells by IL-16 and IL-2. <i>J Immunol.</i> 1998 Mar 1;160(5):2115-20.
IL-2/IL-17	<ul style="list-style-type: none"> • Evidence for the early involvement of interleukin 17 in human and experimental renal allograft rejection. <i>J Pathol.</i> 2002 Jul;197(3):322-32.
IL-2/IL-18	<ul style="list-style-type: none"> • Interleukin 18 (IL-18) in synergy with IL-2 induces lethal lung injury in mice: a potential role for cytokines, chemokines, and natural killer cells in the pathogenesis of interstitial pneumonia. <i>Blood.</i> 2002 Feb 15;99(4):1289-98.
IL-2/TGF- β	<ul style="list-style-type: none"> • Control of CD4 effector fate: transforming growth factor beta 1 and interleukin 2 synergize to prevent apoptosis and promote effector expansion. <i>J Exp Med.</i> 1995 Sep 1;182(3):699-709.
IL-2/IFN- γ	<ul style="list-style-type: none"> • Ig secretion by B-cells • IL-2 induces IFN-γ expression by T-cells
IL-2/IFN- α/β	<ul style="list-style-type: none"> • None
IL-3/IL-4	<ul style="list-style-type: none"> • Synergize in mast cell growth • Synergistic effects of IL-4 and either GM-CSF or IL-3 on the induction of CD23 expression by human monocytes: regulatory effects of IFN-alpha and IFN-gamma. <i>Cytokine.</i> 1994 Jul;6(4):407-13.
IL-3/IL-5	
IL-3/IL-6	
IL-3/IFN- γ	<ul style="list-style-type: none"> • IL-4 and IFN-gamma synergistically increase total polymeric IgA receptor levels in human intestinal epithelial cells. Role of protein tyrosine kinases. <i>J Immunol.</i> 1996 Jun 15;156(12):4807-14.
IL-3/GM-CSF	<ul style="list-style-type: none"> • Differential regulation of human eosinophil IL-3, IL-5, and GM-CSF receptor alpha-chain expression by cytokines: IL-3, IL-5, and GM-CSF down-regulate IL-5 receptor alpha expression with loss of IL-5 responsiveness, but up-regulate IL-3 receptor alpha expression. <i>J Immunol.</i> 2003 Jun 1;170(11):5359-66. (allergic inflammation)
IL-4/IL-2	<ul style="list-style-type: none"> • IL-4 synergistically enhances both IL-2- and IL-12-induced IFN-γ expression in murine NK cells. <i>Blood.</i> 2003 Mar 13 [Epub ahead of print]
IL-4/IL-5	<ul style="list-style-type: none"> • Enhanced mast cell histamine etc. secretion in response to IgE • A Th2-like cytokine response is involved in bullous pemphigoid. the role of IL-4 and IL-5 in the pathogenesis of the disease. <i>Int J Immunopathol Pharmacol.</i> 1999 May-Aug;12(2):55-61.
IL-4/IL-6	
IL-4/IL-10	
IL-4/IL-11	<ul style="list-style-type: none"> • Synergistic interactions between interleukin-11 and interleukin-4 in support of proliferation of primitive hematopoietic progenitors of mice. <i>Blood.</i>

	1991 Sep 15;78(6):1448-51.
IL-4/IL-12	<ul style="list-style-type: none"> • Synergistic effects of IL-4 and IL-18 on IL-12-dependent IFN-gamma production by dendritic cells. <i>J Immunol.</i> 2000 Jan 1;164(1):64-71. (increase Th1/Th2 differentiation) • IL-4 synergistically enhances both IL-2- and IL-12-induced IFN-$\{\gamma\}$ expression in murine NK cells. <i>Blood.</i> 2003 Mar 13 [Epub ahead of print]
IL-4/IL-13	<ul style="list-style-type: none"> • Abstract Interleukin-4 and interleukin-13 signaling connections maps. <i>Science.</i> 2003 Jun 6;300(5625):1527-8. (allergy, asthma) • Inhibition of the IL-4/IL-13 receptor system prevents allergic sensitization without affecting established allergy in a mouse model for allergic asthma. <i>J Allergy Clin Immunol.</i> 2003 Jun;111(6):1361-1369.
IL-4/IL-16	<ul style="list-style-type: none"> • (asthma) Interleukin (IL)-4/IL-9 and exogenous IL-16 induce IL-16 production by BEAS-2B cells, a bronchial epithelial cell line. <i>Cell Immunol.</i> 2001 Feb 1;207(2):75-80
IL-4/IL-17	<ul style="list-style-type: none"> • Interleukin (IL)-4 and IL-17 synergistically stimulate IL-6 secretion in human colonic myofibroblasts. <i>Int J Mol Med.</i> 2002 Nov;10(5):631-4. (Gut inflammation)
IL-4/IL-24	<ul style="list-style-type: none"> • IL-24 is expressed by rat and human macrophages. <i>Immunobiology.</i> 2002 Jul;205(3):321-34.
IL-4/IL-25	<ul style="list-style-type: none"> • Abstract New IL-17 family members promote Th1 or Th2 responses in the lung: in vivo function of the novel cytokine IL-25. <i>J Immunol.</i> 2002 Jul 1;169(1):443-53. (allergic inflammation) • Abstract Mast cells produce interleukin-25 upon Fc epsilon RI-mediated activation. <i>Blood.</i> 2003 May 1;101(9):3594-6. Epub 2003 Jan 02. (allergic inflammation)
IL-4/IFN- γ	<ul style="list-style-type: none"> • Abstract Interleukin 4 induces interleukin 6 production by endothelial cells: synergy with interferon-gamma. <i>Eur J Immunol.</i> 1991 Jan;21(1):97-101.
IL-4/SCF	<ul style="list-style-type: none"> • Regulation of human intestinal mast cells by stem cell factor and IL-4. <i>Immunol Rev.</i> 2001 Feb;179:57-60. Review.
IL-5/IL-3	<ul style="list-style-type: none"> • Differential regulation of human eosinophil IL-3, IL-5, and GM-CSF receptor alpha-chain expression by cytokines: IL-3, IL-5, and GM-CSF down-regulate IL-5 receptor alpha expression with loss of IL-5 responsiveness, but up-regulate IL-3 receptor alpha expression. <i>J Immunol.</i> 2003 Jun 1;170(11):5359-66. (Allergic inflammation see abstract)
IL-5/IL-6	
IL-5/IL-13	<ul style="list-style-type: none"> • Inhibition of allergic airways inflammation and airway hyperresponsiveness in mice by dexamethasone: role of eosinophils, IL-5, eotaxin, and IL-13. <i>J Allergy Clin Immunol.</i> 2003 May;111(5):1049-61.
IL-5/IL-17	<ul style="list-style-type: none"> • Interleukin-17 orchestrates the granulocyte influx into airways after allergen inhalation in a mouse model of allergic asthma. <i>Am J Respir Cell Mol Biol.</i> 2003 Jan;28(1):42-50.
IL-5/IL-25	<ul style="list-style-type: none"> • Abstract New IL-17 family members promote Th1 or Th2 responses in the lung: in vivo function of the novel cytokine IL-25. <i>J Immunol.</i> 2002 Jul 1;169(1):443-53. (allergic inflammation) • Abstract Mast cells produce interleukin-25 upon Fc epsilon RI-mediated activation. <i>Blood.</i> 2003 May 1;101(9):3594-6. Epub 2003 Jan 02. (allergic inflammation)
IL-5/IFN- γ	
IL-5/GM-CSF	<ul style="list-style-type: none"> • Differential regulation of human eosinophil IL-3, IL-5, and GM-CSF receptor alpha-chain expression by cytokines: IL-3, IL-5, and GM-CSF down-regulate IL-5 receptor alpha expression with loss of IL-5 responsiveness, but up-regulate IL-3 receptor alpha expression. <i>J Immunol.</i> 2003 Jun 1;170(11):5359-66. (Allergic inflammation)
IL-6/IL-10	
IL-6/IL-11	
IL-6/IL-16	<ul style="list-style-type: none"> • Interleukin-16 stimulates the expression and production of pro-

	inflammatory cytokines by human monocytes. <i>Immunology</i> . 2000 May;100(1):63-9.
IL-6/IL-17	<ul style="list-style-type: none"> Stimulation of airway mucin gene expression by interleukin (IL)-17 through IL-6 paracrine/autocrine loop. <i>J Biol Chem</i>. 2003 May 9;278(19):17036-43. Epub 2003 Mar 06. (airway inflammation, asthma)
IL-6/IL-19	<ul style="list-style-type: none"> Abstract IL-19 induces production of IL-6 and TNF-alpha and results in cell apoptosis through TNF-alpha. <i>J Immunol</i>. 2002 Oct 15;169(8):4288-97.
IL-6/IFN- γ	
IL-7/IL-2	<ul style="list-style-type: none"> Interleukin 7 worsens graft-versus-host disease. <i>Blood</i>. 2002 Oct 1;100(7):2642-9.
IL-7/IL-12	<ul style="list-style-type: none"> Synergistic effects of IL-7 and IL-12 on human T cell activation. <i>J Immunol</i>. 1995 May 15;154(10):5093-102.
IL-7/IL-15	<ul style="list-style-type: none"> Interleukin-7 and interleukin-15 regulate the expression of the bcl-2 and c-myb genes in cutaneous T-cell lymphoma cells. <i>Blood</i>. 2001 Nov 1;98(9):2778-83. (growth factor)
IL-8/IL-11	<ul style="list-style-type: none"> Abnormal production of interleukin (IL)-11 and IL-8 in polycythaemia vera. <i>Cytokine</i>. 2002 Nov 21;20(4):178-83.
IL-8/IL-17	<ul style="list-style-type: none"> The Role of IL-17 in Joint Destruction. <i>Drug News Perspect</i>. 2002 Jan;15(1):17-23. (arthritis) Abstract Interleukin-17 stimulates the expression of interleukin-8, growth-related oncogene-alpha, and granulocyte-colony-stimulating factor by human airway epithelial cells. <i>Am J Respir Cell Mol Biol</i>. 2002 Jun;26(6):748-53. (airway inflammation)
IL-8/GSF	<ul style="list-style-type: none"> Interleukin-8: an autocrine/paracrine growth factor for human hematopoietic progenitors acting in synergy with colony stimulating factor-1 to promote monocyte-macrophage growth and differentiation. <i>Exp Hematol</i>. 1999 Jan;27(1):28-36.
IL-8/VGEF	<ul style="list-style-type: none"> Intracavitary VEGF, bFGF, IL-8, IL-12 levels in primary and recurrent malignant glioma. <i>J Neurooncol</i>. 2003 May;62(3):297-303.
IL-9/IL-4	<ul style="list-style-type: none"> Anti-interleukin-9 antibody treatment inhibits airway inflammation and hyperreactivity in mouse asthma model. <i>Am J Respir Crit Care Med</i>. 2002 Aug 1;166(3):409-16.
IL-9/IL-5	<ul style="list-style-type: none"> Pulmonary overexpression of IL-9 induces Th2 cytokine expression, leading to immune pathology. <i>J Clin Invest</i>. 2002 Jan;109(1):29-39. Th2 cytokines and asthma. Interleukin-9 as a therapeutic target for asthma. <i>Respir Res</i>. 2001;2(2):80-4. Epub 2001 Feb 15. Review. Abstract Interleukin-9 enhances interleukin-5 receptor expression, differentiation, and survival of human eosinophils. <i>Blood</i>. 2000 Sep 15;96(6):2163-71 (asthma)
IL-9/IL-13	<ul style="list-style-type: none"> Anti-interleukin-9 antibody treatment inhibits airway inflammation and hyperreactivity in mouse asthma model. <i>Am J Respir Crit Care Med</i>. 2002 Aug 1;166(3):409-16. Direct effects of interleukin-13 on epithelial cells cause airway hyperreactivity and mucus overproduction in asthma. <i>Nat Med</i>. 2002 Aug;8(8):885-9.
IL-9/IL-16	<ul style="list-style-type: none"> See IL-4/IL-16
IL-10/IL-2	<ul style="list-style-type: none"> The interplay of interleukin-10 (IL-10) and interleukin-2 (IL-2) in humoral immune responses: IL-10 synergizes with IL-2 to enhance responses of human B lymphocytes in a mechanism which is different from upregulation of CD25 expression. <i>Cell Immunol</i>. 1994 Sep;157(2):478-88.
IL-10/IL-12	
IL-10/TGF- β	<ul style="list-style-type: none"> IL-10 and TGF-beta cooperate in the regulatory T cell response to mucosal allergens in normal immunity and specific immunotherapy. <i>Eur J Immunol</i>. 2003 May;33(5):1205-14.
IL-10/IFN- γ	

IL-11/IL-6	<ul style="list-style-type: none"> Interleukin-6 and interleukin-11 support human osteoclast formation by a RANKL-independent mechanism. <i>Bone</i>. 2003 Jan;32(1):1-7. (bone resorption in inflammation)
IL-11/IL-17	<ul style="list-style-type: none"> Polarized in vivo expression of IL-11 and IL-17 between acute and chronic skin lesions. <i>J Allergy Clin Immunol</i>. 2003 Apr;111(4):875-81. (allergic dermatitis) IL-17 promotes bone erosion in murine collagen-induced arthritis through loss of the receptor activator of NF-κB ligand/osteoprotegerin balance. <i>J Immunol</i>. 2003 Mar 1;170(5):2655-62.
IL-11/TGF- β	<ul style="list-style-type: none"> Polarized in vivo expression of IL-11 and IL-17 between acute and chronic skin lesions. <i>J Allergy Clin Immunol</i>. 2003 Apr;111(4):875-81. (allergic dermatitis)
IL-12/IL-13	<ul style="list-style-type: none"> Relationship of Interleukin-12 and Interleukin-13 imbalance with class-specific rheumatoid factors and anticardiolipin antibodies in systemic lupus erythematosus. <i>Clin Rheumatol</i>. 2003 May;22(2):107-11.
IL-12/IL-17	<ul style="list-style-type: none"> Upregulation of interleukin-12 and -17 in active inflammatory bowel disease. <i>Scand J Gastroenterol</i>. 2003 Feb;38(2):180-5.
IL-12/IL-18	<ul style="list-style-type: none"> Synergistic proliferation and activation of natural killer cells by interleukin 12 and interleukin 18. <i>Cytokine</i>. 1999 Nov;11(11):822-30. Inflammatory Liver Steatosis Caused by IL-12 and IL-18. <i>J Interferon Cytokine Res</i>. 2003 Mar;23(3):155-62.
IL-12/IL-23	<ul style="list-style-type: none"> Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. <i>Nature</i>. 2003 Feb 13;421(6924):744-8. Abstract A unique role for IL-23 in promoting cellular immunity. <i>J Leukoc Biol</i>. 2003 Jan;73(1):49-56. Review.
IL-12/IL-27	<ul style="list-style-type: none"> Abstract IL-27, a heterodimeric cytokine composed of EBI3 and p28 protein, induces proliferation of naive CD4(+) T cells. <i>Immunity</i>. 2002 Jun;16(6):779-90.
IL-12/IFN- γ	<ul style="list-style-type: none"> IL-12 induces IFN-γ expression by B and T-cells as part of immune stimulation.
IL-13/IL-5	<ul style="list-style-type: none"> See IL-5/IL-13
IL-13/IL-25	<ul style="list-style-type: none"> Abstract New IL-17 family members promote Th1 or Th2 responses in the lung: in vivo function of the novel cytokine IL-25. <i>J Immunol</i>. 2002 Jul 1;169(1):443-53. (allergic inflammation) Abstract Mast cells produce interleukin-25 upon Fcepsilon RI-mediated activation. <i>Blood</i>. 2003 May 1;101(9):3594-6. Epub 2003 Jan 02. (allergic inflammation)
IL-15/IL-13	<ul style="list-style-type: none"> Differential expression of interleukins (IL)-13 and IL-15 in ectopic and eutopic endometrium of women with endometriosis and normal fertile women. <i>Am J Reprod Immunol</i>. 2003 Feb;49(2):75-83.
IL-15/IL-16	<ul style="list-style-type: none"> IL-15 and IL-16 overexpression in cutaneous T-cell lymphomas: stage-dependent increase in mycosis fungoides progression. <i>Exp Dermatol</i>. 2000 Aug;9(4):248-51.
IL-15/IL-17	<ul style="list-style-type: none"> Abstract IL-17, produced by lymphocytes and neutrophils, is necessary for lipopolysaccharide-induced airway neutrophilia: IL-15 as a possible trigger. <i>J Immunol</i>. 2003 Feb 15;170(4):2106-12. (airway inflammation)
IL-15/IL-21	<ul style="list-style-type: none"> IL-21 in Synergy with IL-15 or IL-18 Enhances IFN-gamma Production in Human NK and T Cells. <i>J Immunol</i>. 2003 Jun 1;170(11):5464-9.
IL-17/IL-23	<ul style="list-style-type: none"> Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. <i>J Biol Chem</i>. 2003 Jan 17;278(3):1910-4. Epub 2002 Nov 03
IL-17/TGF- β	<ul style="list-style-type: none"> Polarized in vivo expression of IL-11 and IL-17 between acute and chronic skin lesions. <i>J Allergy Clin Immunol</i>. 2003 Apr;111(4):875-81. (allergic dermatitis)
IL-18/IL-12	<ul style="list-style-type: none"> Synergistic proliferation and activation of natural killer cells by interleukin

	<ul style="list-style-type: none">12 and interleukin 18. <i>Cytokine</i>. 1999 Nov;11(11):822-30.Abstract Inhibition of in vitro immunoglobulin production by IL-12 in murine chronic graft-vs.-host disease: synergism with IL-18. <i>Eur J Immunol</i>. 1998 Jun;28(6):2017-24.
IL-18/IL-21	<ul style="list-style-type: none">IL-21 in Synergy with IL-15 or IL-18 Enhances IFN-gamma Production in Human NK and T Cells. <i>J Immunol</i>. 2003 Jun 1;170(11):5464-9.
IL-18/TGF- β	<ul style="list-style-type: none">Interleukin 18 and transforming growth factor beta1 in the serum of patients with Graves' ophthalmopathy treated with corticosteroids. <i>Int Immunopharmacol</i>. 2003 Apr;3(4):549-52.
IL-18/IFN- γ	
Anti-TNF ALPHA/anti-CD4	<ul style="list-style-type: none">Synergistic therapeutic effect in DBA/1 arthritic mice.

Annex 3: Oncology combinations

Target	Disease	Pair with
CD89*	Use as cytotoxic cell recruiter	all
CD19	B cell lymphomas	HLA-DR CD5
HLA-DR	B cell lymphomas	CD89 CD19 CD5
CD38	Multiple myeloma	CD138 CD56 HLA-DR
CD138	Multiple myeloma	CD38 CD56 HLA-DR
CD138	Lung cancer	CD56 CEA
CD33	Acute myeloid lymphoma	CD34 HLA-DR
CD56	Lung cancer	CD138 CEA
CEA	Pan carcinoma	MET receptor
VEGF	Pan carcinoma	MET receptor
VEGF receptor	Pan carcinoma	MET receptor
IL-13	Asthma/pulmonary inflammation	IL-4 IL-5 Eotaxin(s) MDC TARC TNF α IL-9 EGFR CD40L IL-25 MCP-1 TGF β
IL-4	Asthma	IL-13 IL-5 Eotaxin(s) MDC TARC TNF α IL-9 EGFR CD40L IL-25 MCP-1 TGF β
Eotaxin	Asthma	IL-5 Eotaxin-2 Eotaxin-3
EGFR	cancer	HER2/neu HER3 HER4
HER2	cancer	HER3

		HER4
TNFR1	RA/Crohn's disease	IL-1R IL-6R IL-18R
TNF α	RA/Crohn's disease	IL-1 α / β IL-6 IL-18 ICAM-1 IL-15 IL-17
IL-1R	RA/Crohn's disease	IL-6R IL-18R
IL-18R	RA/Crohn's disease	IL-6R

Annex 4Data Summary

TARGET	dAb	Equilibrium dissociation constant (Kd = Koff/Kon)	Koff	IC50 for ligand assay	ND50 for cell based neutralism assay
TAR1	TAR1 monomers	300nM to 5pM (ie, 3×10^{-7} to 5×10^{-12}), preferably 50nM to 20pM	5×10^{-1} to 1×10^{-7}	500nM to 100pM	500nM to 50pM
	TAR1 dimers	As TAR1 monomer	As TAR1 monomer	As TAR1 monomer	As TAR1 monomer
	TAR1 trimers	As TAR1 monomer	As TAR1 monomer	As TAR1 monomer	As TAR1 monomer
TAR1-5					
TAR1-27					
TAR1-5-19 monomer		30nM			
TAR1-5-19 homodimer				With (Gly ₃ Ser) ₃ linker = 20nM With (Gly ₃ Ser) ₅ linker = 2nM With (Gly ₄ Ser) ₇ linker = 10nM <u>In Fab format = 1nM</u>	=30nM =3nM =1.5nM
TAR1-5-19 heterodimers				With (Gly ₄ Ser) _n linker TAR1-5-19 d2 = 2nM TAR1-5-19 d3 = 8nM TAR1-5-19 d4 = 2.5nM TAR1-5-19 d5 = 8nM	=12nM =10nM
				In Fab format TAR1-5-19CH d1CK = 6nM TAR1-5-19CK d1CH = 6nM TAR1-5-19CH d2CK = 8nM TAR1-5-19CH d3CK = 3nM	=12nM

TAR1-5 heterodimers	With (Gly ₄ Ser) _n linker TAR1-5d1 = 30nM TAR1-5d2 = 50nM TAR1-5d3 = 300nM <u>TAR1-5d4 = 3nM</u> TAR1-5d5 = 200nM TAR1-5d6 = 100nM	
	In Fab format <u>TAR1-5CH d2CK = 30nM</u> <u>TAR1-5CK d3CH = 100nM</u>	
		= 60nM
TAR1-5-19 homotrimer		0.3nM
TAR2	TAR2 monomers	As TAR1 monomer 500nM to 100pM
TAR2-10		500nM to 50pM
TAR2-5		
Serum Albumin	Anti-SA monomers	1nM to 500 μ M, preferably 100nM to 10 μ M In Dual Specific format, target affinity is 1 to 100,000 x affinity of SA dAb affinity, eg 100pM (target) and 10 μ M SA affinity.
	MSA-16	200nM
	MSA-26	70nM

Claims

1. A dual-specific ligand comprising a first immunoglobulin single variable domain having a binding specificity to a first epitope or antigen and a second complementary immunoglobulin single variable domain having a binding activity to a second epitope or antigen, wherein one or both of said antigens or epitopes acts to increase the half-life of the ligand *in vivo* and wherein said first and second domains lack mutually complementary domains which share the same specificity, provided that said dual specific ligand does not consist of an anti-HSA V_H domain and an anti- β galactosidase V_K domain.
2. A dual-specific ligand according to claim 1, comprising at least one single heavy chain variable domain of an antibody and one complementary single light chain variable domain of an antibody such that the two regions are capable of associating to form a complementary VH/VL pair.
3. A dual specific ligand according to claim 2 wherein the V_H and V_L are provided by an antibody scFv fragment.
4. A dual-specific ligand according to claim 2 wherein the V_H and V_L are provided by an antibody Fab region.
5. A four chain IgG immunoglobulin ligand comprising a dual specific ligand of claim 2.
6. A four chain IgG immunoglobulin ligand according to claim 5, wherein said IgG comprises two dual specific ligands, said dual specific ligands being identical in their variable domains.
7. A four chain IgG immunoglobulin ligand according to claim 5, wherein said IgG comprises two dual specific ligands, said dual specific ligands being different in their variable domains.
8. A ligand comprising a first immunoglobulin variable domain having a first antigen or epitope binding specificity and a second immunoglobulin variable domain having a second antigen or epitope binding specificity wherein one or both of said first and second variable domains bind to an antigen which increases the half-life of the ligand *in vivo*, and the variable domains are not complementary to one another.
9. A ligand according to claim 8 wherein the first and the second immunoglobulin variable domains are heavy chain variable domains (V_H).
10. A ligand according to claim 8 wherein the first and the second immunoglobulin variable domains are light chain variable domains (V_L).
11. A ligand according to claim any preceding claim, wherein the first and second epitopes bind independently, such that the dual specific ligand may simultaneously bind both the first and second epitopes or antigens.

12. A ligand according to claim 11, wherein the dual specific ligand comprises a first form and a second form in equilibrium in solution, wherein both epitopes or antigens bind to the first form independently but compete for binding to the second form.
13. A ligand according to any preceding claim wherein the variable regions are derived from immunoglobulins directed against said epitopes or antigens.
14. A ligand according to any preceding claim, wherein said first and second epitopes are present on separate antigens.
15. A ligand according to any one of claims 1 to 11, wherein said first and second epitopes are present on the same antigen.
16. A ligand according to any preceding claim comprising a variable domain that is derived from a repertoire of single antibody domains.
17. A ligand of claim 16 wherein said repertoire is displayed on the surface of filamentous bacteriophage and wherein the single antibody domains are selected by binding of the bacteriophage repertoire to antigen.
18. A ligand according to any preceding claim wherein the sequence of at least one variable domain is modified by mutation or DNA shuffling.
19. A dual-specific ligand according to any preceding claim wherein the variable regions are non-covalently associated.
20. A dual-specific ligand according to any one of claims 1 to 18 wherein the variable regions are covalently associated.
21. A dual-specific ligand according to claim 20 wherein the covalent association is mediated by disulphide bonds.
22. A dAb monomer ligand specific for TNF α , which dissociates from human TNF α with a dissociation constant (K_d) of 50nM to 20pM, and a K_{off} rate constant of 5×10^{-1} to 1×10^{-7} s $^{-1}$, as determined by surface plasmon resonance.
23. A dAb monomer ligand specific for TNF α according to claim 22, wherein the dAb is a V κ .
24. A dAb monomer ligand specific for TNF receptor 1 (p55), which dissociates from human TNF receptor 1 with a dissociation constant (K_d) of 50nM to 20pM, and a K_{off} rate constant of 5×10^{-1} to 1×10^{-7} s $^{-1}$, as determined by surface plasmon resonance.
25. A dAb monomer ligand according to claim 22 to claim 24, wherein the monomer neutralises human TNF α or TNF receptor 1 in a standard cell assay with an ND50 of 500nM to 50pM.

26. A dAb monomer ligand specific for TNF receptor 1 (p55), wherein the dAb antagonises the activity of the TNF receptor 1 in a standard cell assay with an ND₅₀ of ≤100nM, and at a concentration of ≤10μM the dAb agonises the activity of the TNF receptor 1 by ≤5% in the assay.
27. A dAb monomer ligand specific for serum albumin (SA) which dissociates from SA with a dissociation constant (K_d) of 1nM to 500μM, as determined by surface plasmon resonance.
28. A dAb monomer ligand according to claim 27, wherein the monomer binds SA in a standard ligand binding assay with an IC50 of 1nM to 500μM.
29. A dAb monomer ligand specific for TNF α , wherein the dAb comprises the amino acid sequence of TAR1-5-19 or a sequence that is at least 80% homologous thereto.
30. A dAb monomer ligand specific for TNF α , wherein the dAb comprises the amino acid sequence of TAR1-5 or a sequence that is at least 80% homologous thereto.
31. A dAb monomer ligand specific for TNF α , wherein the dAb comprises the amino acid sequence of TAR1-27 or a sequence that is at least 80% homologous thereto.
32. A dAb monomer ligand specific for TNF receptor 1, wherein the dAb comprises the amino acid sequence of TAR2-10 or a sequence that is at least 80% homologous thereto.
33. A dAb monomer ligand specific for TNF receptor 1, wherein the dAb comprises the amino acid sequence of TAR2-10 or a sequence that is at least 90% homologous thereto.
34. A dAb monomer ligand specific for TNF receptor 1, wherein the dAb comprises the amino acid sequence of TAR2-5 or a sequence that is at least 80% homologous thereto.
35. A dAb monomer ligand specific for TNF receptor 1, wherein the dAb comprises the amino acid sequence of TAR2-5 or a sequence that is at least 90% homologous thereto.
36. A dAb monomer ligand specific for SA, wherein the dAb comprises the amino acid sequence of MSA-16 or a sequence that is at least 80% homologous thereto.
37. A dAb monomer ligand specific for SA, wherein the dAb comprises the amino acid sequence of MSA-26 or a sequence that is at least 80% homologous thereto.

38. A dAb monomer according to any one of claims 29 to 37, wherein the TNF α , TNF receptor 1 or SA is in human form.
39. A dAb monomer further comprising a terminal Cys residue.
40. A dAb monomer according to any one of claims 29 to 38, further comprising a terminal Cys residue.
41. A dual specific ligand comprising at least one dAb monomer according to any one of claims 22 to 40.
42. A dual specific ligand according to claim 41, which is a dimer.
43. A dual specific ligand according to claim 42, wherein the dimer comprises anti-human TNF alpha dAb according to any one of claims 22, 23 and 28-30, and an anti-SA dAb according to any one of claims 26, 27 and 34-36.
44. A dual specific ligand according to claim 42, wherein the dimer is a homo- or hetero-dimer comprising first and second anti-human TNF alpha dAbs, each dAb being according to any one of claims 22, 23 and 28-30.
45. A dual specific ligand according to claim 41, which is a trimer.
46. A dual specific ligand according to claim 45, which is a homotrimer comprising three copies of an anti-human TNF alpha dAb according to any one of claims 22, 23 and 29-31.
47. A ligand according to any preceding claim, which comprises a universal framework.
48. A ligand according to claim 47, wherein the universal framework comprises a V_H framework selected from the group consisting of DP47, DP45 and DP38; and/or the V_L framework is DPK9.
49. A ligand according to any preceding claim which comprises a binding site for a generic ligand.
50. The ligand of claim 49, wherein the generic ligand binding site is selected from the group consisting of protein A, protein L and protein G.
51. A ligand according to any preceding claim, wherein the ligand comprises a variable domain having one or more framework regions comprising an amino acid sequence that is the same as the amino acid sequence of a corresponding framework region encoded by a human germline antibody gene segment, or the amino acid sequences of one or more of said framework regions collectively comprises up to 5 amino acid differences relative to the amino acid sequence of said corresponding framework region encoded by a human germline antibody gene segment.

52. A ligand according to any one of claims 1 to 51, wherein the ligand comprises a variable domain, wherein the amino acid sequences of FW1, FW2, FW3 and FW4 are the same as the amino acid sequences of corresponding framework regions encoded by a human germline antibody gene segment, or the amino acid sequences of FW1, FW2, FW3 and FW4 collectively contain up to 10 amino acid differences relative to the amino acid sequences of corresponding framework regions encoded by said human germline antibody gene segment.

53. The ligand according to claim 51 or claim 52, which comprises an antibody variable domain comprising FW1, FW2 and FW3 regions, and the amino acid sequence of said FW1, FW2 and FW3 are the same as the amino acid sequences of corresponding framework regions encoded by human germline antibody gene segments.

54. The ligand according to any one of claims 51 to 53, wherein said human germline antibody gene segment is selected from the group consisting of DP47, DP45, DP48 and DPK9.

55. A ligand according to any preceding claim, comprising a V_H domain that is not a Camelid immunoglobulin variable domain.

56. The ligand of Claim 55, comprising a V_H domain that does not contain one or more amino acids that are specific to Camelid immunoglobulin variable domains as compared to human V_H domains.

57. A method for producing a ligand comprising a first immunoglobulin single variable domain having a first binding specificity and a second single immunoglobulin single variable domain having a second binding specificity, one or both of the binding specificities being specific for a protein which increases the half-life of the ligand *in vivo*, the method comprising the steps of:

- (a) selecting a first variable domain by its ability to bind to a first epitope,
- (b) selecting a second variable region by its ability to bind to a second epitope,
- (c) combining the variable regions; and
- (d) selecting the ligand by its ability to bind to said first and second epitopes; wherein, when said variable domains are complementary, neither of said domains is a V_H domain specific for HSA.

58. A method according to claim 57 wherein said first variable domain is selected for binding to said first epitope in absence of a complementary variable domain.

59. A method according to claim 57 wherein said first variable domain is selected for binding to said first epitope in the presence of a third complementary variable domain in which said third variable domain is different from said second variable domain.

60. Nucleic acid encoding a dual-specific ligand according to any one of claims 1 to 56.

61. A nucleic acid according to claim 60 which is specific for TNF α , comprising the nucleic acid sequence of TAR1-5-19 or a sequence that is at least 70% homologous thereto.

62. A nucleic acid according to claim 60 which is specific for TNF α , comprising the nucleic acid sequence of TAR1-5 or a sequence that is at least 70% homologous thereto.

63. A nucleic acid according to claim 60 which is specific for TNF α , comprising the nucleic acid sequence of TAR1-27 or a sequence that is at least 70% homologous thereto.

64. A nucleic acid according to claim 60 which is specific for TNF receptor 1, comprising the nucleic acid sequence of TAR2-10 or a sequence that is at least 70% homologous thereto.

65. A nucleic acid according to claim 60 which is specific for TNF receptor 1, comprising the nucleic acid sequence of TAR2-10 or a sequence that is at least 80% homologous thereto.

66. A nucleic acid according to claim 60 which is specific for TNF receptor 1, comprising the nucleic acid sequence of TAR2h-5 or a sequence that is at least 70% homologous thereto.

67. A nucleic acid according to claim 60 which is specific for TNF receptor 1, comprising the nucleic acid sequence of TAR2h-5 or a sequence that is at least 80% homologous thereto.

68. A nucleic acid according to claim 60 which is specific for SA, comprising the nucleic acid sequence of MSA-16 or a sequence that is at least 70% homologous thereto.

69. A nucleic acid according to claim 60 which is specific for SA, comprising the nucleic acid sequence of MSA-26 or a sequence that is at least 70% homologous thereto.

70. A vector comprising nucleic acid according to any one of claims 60 to 69.

71. A vector according to claim 70, further comprising components necessary for the expression of a dual-specific ligand.

72. A host cell transfected with a vector according to claim 71.

73. A method for producing a closed conformation multi-specific ligand comprising a first single epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity, wherein the first and second binding specificities are capable of competing for epitope binding such that the closed

conformation multi-specific ligand may not bind both epitopes simultaneously, said method comprising the steps of:

- a) selecting a first epitope binding domain by its ability to bind to a first epitope,
- b) selecting a second epitope binding domain by its ability to bind to a second epitope,
- c) combining the epitope binding domains such that the domains are in a closed conformation; and
- d) selecting the closed conformation multispecific ligand by its ability to bind to said first second epitope and said second epitope, but not to both said first and second epitopes simultaneously.

74. A method according to claim 73 wherein the first and the second epitope binding domains are immunoglobulin variable heavy chain domains (V_H).

75. A method according to claim 73 wherein the first and the second immunoglobulin variable domains are immunoglobulin variable light chain domains (V_L).

76. A method according to any one of claims 73 to 75 wherein the immunoglobulin domains are derived from immunoglobulins directed against said epitopes.

77. A method according to any one of claims 73 to 76, wherein said first and second epitopes are present on separate antigens.

78. A method according to any one of claims 73 to 76, wherein said first and second epitopes are present on the same antigen.

79. A method according to any one of claims 73 to 78 wherein the variable domain is derived from a repertoire of single antibody domains.

80. A method of claim 79 wherein said repertoire is displayed on the surface of filamentous bacteriophage and wherein the single antibody domains are selected by binding of the bacteriophage repertoire to antigen.

81. A method of any one of claims 73 to 80 wherein the sequence of at least one immunoglobulin variable domain is modified by mutation or DNA shuffling.

82. A closed conformation multispecific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity wherein the first and second binding specificities are capable of competing for epitope binding such that the closed conformation multi-specific ligand cannot bind both epitopes simultaneously.

83. A closed conformation multispecific ligand according to claim 82, obtainable by a method according to any one of claims 73 to 80.

84. A closed conformation multispecific ligand according to claim 82 or claim 83, comprising more than one single heavy chain variable domain of an antibody or more than one light chain variable domain of an antibody.

85. A closed conformation multi-specific ligand according to claim 84 wherein the V_H and V_L are linked by a peptide linker.

86. A closed conformation multi-specific ligand according to claim 84 wherein the V_H or V_L are provided by an antibody Fab-like region.

87. A closed conformation multi-specific ligand according to any one of claims 82 to 84 wherein the variable regions are non-covalently associated.

88. A closed conformation multi-specific ligand according to any one of claims 82 to 84 wherein the variable regions are covalently associated.

89. A closed conformation multi-specific ligand according to claim 87 wherein the covalent association is mediated by disulphide bonds.

90. A closed conformation multi-specific ligand according to any of claims 82 to 89 which comprises a universal framework.

91. A closed conformation multi-specific ligand according to any of claims 82 to 90 which comprises a binding site for a generic ligand.

92. The closed conformation multi-specific ligand of claim 91, wherein the generic ligand binding site is selected from the group consisting of protein A, protein L and protein G.

93. A closed conformation ligand according to any of claims 82 to 92, wherein the ligand comprises a variable domain having one or more framework regions comprising an amino acid sequence that is the same as the amino acid sequence of a corresponding framework region encoded by a human germline antibody gene segment, or the amino acid sequences of one or more of said framework regions collectively comprises up to 5 amino acid differences relative to the amino acid sequence of said corresponding framework region encoded by a human germline antibody gene segment.

94. The closed conformation ligand according to claim 93, wherein the ligand comprises a variable domain wherein the amino acid sequences of FW1, FW2, FW3 and FW4 are the same as the amino acid sequences of corresponding framework regions encoded by a human germline antibody gene segment, or the amino acid sequences of FW1, FW2, FW3 and FW4 collectively contain up to 10 amino acid differences relative to the amino acid sequences of corresponding framework regions encoded by said human germline antibody gene segment.

95. The closed conformation ligand according to claim 93 or claim 94, which comprises an antibody variable domain comprising FW1, FW2 and FW3 regions,

and the amino acid sequence of said FW1, FW2 and FW3 are the same as the amino acid sequences of corresponding framework regions encoded by human germline antibody gene segments.

96. The closed conformation ligand according to any one of claims 92 to 95, wherein said human germline antibody gene segment is selected from the group consisting of DP47, DP45, DP48 and DPK9.

97. A closed conformation ligand according to any one of claims 92 to 96, comprising a V_H domain that is not a Camelid immunoglobulin variable domain.

98. The closed conformation ligand of Claim 97, wherein the V_H domain does not contain one or more amino acids that are specific to Camelid immunoglobulin variable domains as compared to human V_H domains.

99. A closed conformation multi-specific ligand according to any one of claims 82 to 98, wherein one specificity thereof is for an agent effective to increase the half life of the ligand.

100. A kit comprising a closed conformation multi-specific ligand according to any one of claims 82 to 99.

101. Nucleic acid encoding at least a closed conformation multispecific ligand according to any one of claims 82 to 99.

102. A vector comprising nucleic acid according to claim 101.

103. A vector according to claim 102, further comprising components necessary for the expression of a closed conformation multispecific ligand.

104. A host cell transfected with a vector according to claim 103.

105. A method for detecting the presence of a target molecule, comprising:
(a) providing a closed conformation multispecific ligand bound to an agent, said ligand being specific for the target molecule and the agent, wherein the agent which is bound by the ligand leads to the generation of a detectable signal on displacement from the ligand;
(b) exposing the closed conformation multispecific ligand to the target molecule; and
(c) detecting the signal generated as a result of the displacement of the agent.

106. A method according to claim 105, wherein the agent is an enzyme, which is inactive when bound by the closed conformation multispecific ligand.

107. A method according to claim 105, wherein the agent is the substrate for an enzyme

108. A method according to claim 107, wherein the agent is a fluorescent, luminescent or chromogenic molecule which is inactive or quenched when bound by the ligand.

109. A kit for performing a method according to any one of claims 105-108, comprising a closed conformation multispecific ligand capable of binding to a target molecule, and optionally an agent and buffers suitable therefor.

110. A homogenous immunoassay incorporating a method according to any one of claims 105-108.

111. A ligand according to any one of claims 1 to 56 for use in therapy.

112. A pharmaceutical composition comprising a ligand according to any one of claims 1 to 56, and a pharmaceutically acceptable excipient, carrier or diluent.

113. A method for preparing a chelating multimeric ligand comprising the steps of:

- (a) providing a vector comprising a nucleic acid sequence encoding a single binding domain specific for a first epitope on a target;
- (b) providing a vector encoding a repertoire comprising second binding domains specific for a second epitope on said target, which epitope can be the same or different to the first epitope, said second epitope being adjacent to said first epitope; and
- (c) expressing said first and second binding domains; and
- (d) isolating those combinations of first and second binding domains which combine together to produce a target-binding dimer.

114. A method according to claim 113, wherein the first and second binding domains are associated covalently through a linker.

115. A method according to claim 113, wherein the first and second binding domains are associated non-covalently.

116. A method according to claim 113, wherein the first and second binding domains are associated through natural association of the domains.

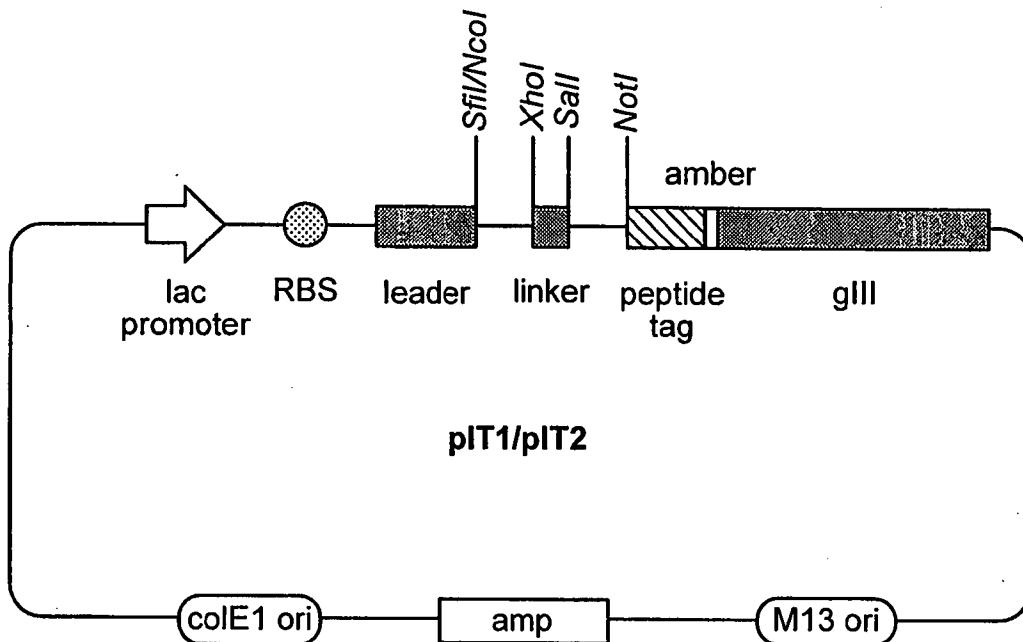
117. A method according to claim 116, wherein the binding domains comprise a V_H domain and a V_{κ} domain.

FIG. 1

H1 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F S GAG GTG CAG CTG TTG GAG TCT GGG GGA GGC TTG GTA CAG CCT GGG TCC CTG AGA CTC TCC TGT GCA GCC TCT GGA TTC ACC TTT AGC H30

H10 S Y A M S W V R Q A P G K G L E W V S A I H50 H52 a H80 H82 a b c H90 H100 H110 H113 H120 H130 H140 H150 H160 H170 H180 H190 H20 H21 H22 H23 H24 H25 H26 H27 H28 H29 H30 H31 H32 H33 H34 H35 H36 H37 H38 H39 H40 H41 H42 H43 H44 H45 H46 H47 H48 H49 H50 H51 H52 H53 H54 H55 H56 H57 H58 H59 H60 H61 H62 H63 H64 H65 H66 H67 H68 H69 H70 H71 H72 H73 H74 H75 H76 H77 H78 H79 H80 H81 H82 H83 H84 H85 H86 H87 H88 H89 H90 H91 H92 H93 H94 H95 H96 H97 H98 H99 H100 H101 H102 H103 H104 H105 H106 H107 H108 H109 H110 H111 H112 H113 H114 H115 H116 H117 H118 H119 H120 H121 H122 H123 H124 H125 H126 H127 H128 H129 H130 H131 H132 H133 H134 H135 H136 H137 H138 H139 H140 H141 H142 H143 H144 H145 H146 H147 H148 H149 H150 H151 H152 H153 H154 H155 H156 H157 H158 H159 H160 H161 H162 H163 H164 H165 H166 H167 H168 H169 H170 H171 H172 H173 H174 H175 H176 H177 H178 H179 H180 H181 H182 H183 H184 H185 H186 H187 H188 H189 H190 H191 H192 H193 H194 H195 H196 H197 H198 H199 H200 H201 H202 H203 H204 H205 H206 H207 H208 H209 H2010 H2011 H2012 H2013 H2014 H2015 H2016 H2017 H2018 H2019 H2020 H2021 H2022 H2023 H2024 H2025 H2026 H2027 H2028 H2029 H2030 H2031 H2032 H2033 H2034 H2035 H2036 H2037 H2038 H2039 H2040 H2041 H2042 H2043 H2044 H2045 H2046 H2047 H2048 H2049 H2050 H2051 H2052 H2053 H2054 H2055 H2056 H2057 H2058 H2059 H2060 H2061 H2062 H2063 H2064 H2065 H2066 H2067 H2068 H2069 H2070 H2071 H2072 H2073 H2074 H2075 H2076 H2077 H2078 H2079 H2080 H2081 H2082 H2083 H2084 H2085 H2086 H2087 H2088 H2089 H2090 H2091 H2092 H2093 H2094 H2095 H2096 H2097 H2098 H2099 H20100 H20101 H20102 H20103 H20104 H20105 H20106 H20107 H20108 H20109 H20110 H20111 H20112 H20113 H20114 H20115 H20116 H20117 H20118 H20119 H20120 H20121 H20122 H20123 H20124 H20125 H20126 H20127 H20128 H20129 H20130 H20131 H20132 H20133 H20134 H20135 H20136 H20137 H20138 H20139 H20140 H20141 H20142 H20143 H20144 H20145 H20146 H20147 H20148 H20149 H20150 H20151 H20152 H20153 H20154 H20155 H20156 H20157 H20158 H20159 H20160 H20161 H20162 H20163 H20164 H20165 H20166 H20167 H20168 H20169 H20170 H20171 H20172 H20173 H20174 H20175 H20176 H20177 H20178 H20179 H20180 H20181 H20182 H20183 H20184 H20185 H20186 H20187 H20188 H20189 H20190 H20191 H20192 H20193 H20194 H20195 H20196 H20197 H20198 H20199 H20200 H20201 H20202 H20203 H20204 H20205 H20206 H20207 H20208 H20209 H202010 H202011 H202012 H202013 H202014 H202015 H202016 H202017 H202018 H202019 H202020 H202021 H202022 H202023 H202024 H202025 H202026 H202027 H202028 H202029 H202030 H202031 H202032 H202033 H202034 H202035 H202036 H202037 H202038 H202039 H202040 H202041 H202042 H202043 H202044 H202045 H202046 H202047 H202048 H202049 H202050 H202051 H202052 H202053 H202054 H202055 H202056 H202057 H202058 H202059 H202060 H202061 H202062 H202063 H202064 H202065 H202066 H202067 H202068 H202069 H202070 H202071 H202072 H202073 H202074 H202075 H202076 H202077 H202078 H202079 H202080 H202081 H202082 H202083 H202084 H202085 H202086 H202087 H202088 H202089 H202090 H202091 H202092 H202093 H202094 H202095 H202096 H202097 H202098 H202099 H2020100 H2020101 H2020102 H2020103 H2020104 H2020105 H2020106 H2020107 H2020108 H2020109 H2020110 H2020111 H2020112 H2020113 H2020114 H2020115 H2020116 H2020117 H2020118 H2020119 H2020120 H2020121 H2020122 H2020123 H2020124 H2020125 H2020126 H2020127 H2020128 H2020129 H2020130 H2020131 H2020132 H2020133 H2020134 H2020135 H2020136 H2020137 H2020138 H2020139 H2020140 H2020141 H2020142 H2020143 H2020144 H2020145 H2020146 H2020147 H2020148 H2020149 H2020150 H2020151 H2020152 H2020153 H2020154 H2020155 H2020156 H2020157 H2020158 H2020159 H2020160 H2020161 H2020162 H2020163 H2020164 H2020165 H2020166 H2020167 H2020168 H2020169 H2020170 H2020171 H2020172 H2020173 H2020174 H2020175 H2020176 H2020177 H2020178 H2020179 H2020180 H2020181 H2020182 H2020183 H2020184 H2020185 H2020186 H2020187 H2020188 H2020189 H2020190 H2020191 H2020192 H2020193 H2020194 H2020195 H2020196 H2020197 H2020198 H2020199 H2020200 H2020201 H2020202 H2020203 H2020204 H2020205 H2020206 H2020207 H2020208 H2020209 H20202010 H20202011 H20202012 H20202013 H20202014 H20202015 H20202016 H20202017 H20202018 H20202019 H20202020 H20202021 H20202022 H20202023 H20202024 H20202025 H20202026 H20202027 H20202028 H20202029 H202020200 H202020201 H202020202 H202020203 H202020204 H202020205 H202020206 H202020207 H202020208 H202020209 H2020202010 H2020202011 H2020202012 H2020202013 H2020202014 H2020202015 H2020202016 H2020202017 H2020202018 H2020202019 H2020202020 H2020202021 H2020202022 H2020202023 H2020202024 H2020202025 H2020202026 H2020202027 H2020202028 H2020202029 H20202020200 H20202020201 H20202020202 H20202020203 H20202020204 H20202020205 H20202020206 H20202020207 H20202020208 H20202020209 H202020202010 H202020202011 H202020202012 H202020202013 H202020202014 H202020202015 H202020202016 H202020202017 H202020202018 H202020202019 H202020202020 H202020202021 H202020202022 H202020202023 H202020202024 H202020202025 H202020202026 H202020202027 H202020202028 H202020202029 H2020202020200 H2020202020201 H2020202020202 H2020202020203 H2020202020204 H2020202020205 H2020202020206 H2020202020207 H2020202020208 H2020202020209 H20202020202010 H20202020202011 H20202020202012 H20202020202013 H20202020202014 H20202020202015 H20202020202016 H20202020202017 H20202020202018 H20202020202019 H20202020202020 H20202020202021 H20202020202022 H20202020202023 H20202020202024 H20202020202025 H20202020202026 H20202020202027 H20202020202028 H20202020202029 H202020202020200 H202020202020201 H202020202020202 H202020202020203 H202020202020204 H202020202020205 H202020202020206 H202020202020207 H202020202020208 H202020202020209 H2020202020202010 H2020202020202011 H2020202020202012 H2020202020202013 H2020202020202014 H2020202020202015 H2020202020202016 H2020202020202017 H2020202020202018 H2020202020202019 H2020202020202020 H2020202020202021 H2020202020202022 H2020202020202023 H2020202020202024 H2020202020202025 H2020202020202026 H2020202020202027 H2020202020202028 H2020202020202029 H20202020202020200 H20202020202020201 H20202020202020202 H20202020202020203 H20202020202020204 H20202020202020205 H20202020202020206 H20202020202020207 H20202020202020208 H20202020202020209 H202020202020202010 H202020202020202011 H202020202020202012 H202020202020202013 H202020202020202014 H202020202020202015 H202020202020202016 H202020202020202017 H202020202020202018 H202020202020202019 H202020202020202020 H202020202020202021 H202020202020202022 H202020202020202023 H202020202020202024 H202020202020202025 H202020202020202026 H202020202020202027 H202020202020202028 H202020202020202029 H2020202020202020200 H2020202020202020201 H2020202020202020202 H2020202020202020203 H2020202020202020204 H2020202020202020205 H2020202020202020206 H2020202020202020207 H2020202020202020208 H2020202020202020209 H20202020202020202010 H20202020202020202011 H20202020202020202012 H20202020202020202013 H20202020202020202014 H20202020202020202015 H20202020202020202016 H20202020202020202017 H20202020202020202018 H20202020202020202019 H20202020202020202020 H20202020202020202021 H20202020202020202022 H20202020202020202023 H20202020202020202024 H20202020202020202025 H20202020202020202026 H20202020202020202027 H20202020202020202028 H20202020202020202029 H202020202020202020200 H202020202020202020201 H202020202020202020202 H202020202020202020203 H202020202020202020204 H202020202020202020205 H202020202020202020206 H202020202020202020207 H202020202020202020208 H202020202020202020209 H2020202020202020202010 H2020202020202020202011 H2020202020202020202012 H2020202020202020202013 H2020202020202020202014 H2020202020202020202015 H2020202020202020202016 H2020202020202020202017 H2020202020202020202018 H2020202020202020202019 H2020202020202020202020 H2020202020202020202021 H2020202020202020202022 H2020202020202020202023 H2020202020202020202024 H2020202020202020202025 H2020202020202020202026 H2020202020202020202027 H2020202020202020202028 H2020202020202020202029 H20202020202020202020200 H20202020202020202020201 H20202020202020202020202 H20202020202020202020203 H20202020202020202020204 H20202020202020202020205 H20202020202020202020206 H20202020202020202020207 H20202020202020202020208 H20202020202020202020209 H202020202020202020202010 H202020202020202020202011 H202020202020202020202012 H202020202020202020202013 H202020202020202020202014 H202020202020202020202015 H202020202020202020202016 H202020202020202020202017 H202020202020202020202018 H202020202020202020202019 H202020202020202020202020 H202020202020202020202021 H202020202020202020202022 H202020202020202020202023 H202020202020202020202024 H202020202020202020202025 H202020202020202020202026 H202020202020202020202027 H202020202020202020202028 H202020202020202020202029 H2020202020202020202020200 H2020202020202020202020201 H2020202020202020202020202 H2020202020202020202020203 H2020202020202020202020204 H2020202020202020202020205 H2020202020202020202020206 H2020202020202020202020207 H2020202020202020202020208 H2020202020202020202020209 H20202020202020202020202010 H20202020202020202020202011 H20202020202020202020202012 H20202020202020202020202013 H20202020202020202020202014 H20202020202020202020202015 H20202020202020202020202016 H20202020202020202020202017 H20202020202020202020202018 H20202020202020202020202019 H20202020202020202020202020 H20202020202020202020202021 H20202020202020202020202022 H20202020202020202020202023 H20202020202020202020202024 H20202020202020202020202025 H202020202020202020202026 H202020202020202020202027 H202020202020202020202028 H202020202020202020202029 H2020202020202020202020200 H2020202020202020202020201 H2020202020202020202020202 H2020202020202020202020203 H2020202020202020202020204 H2020202020202020202020205 H2020202020202020202020206 H2020202020202020202020207 H2020202020202020202020208 H2020202020202020202020209 H20202020202020202020202010 H20202020202020202020202011 H20202020202020202020202012 H20202020202020202020202013 H20202020202020202020202014 H20202020202020202020202015 H20202020202020202020202016 H20202020202020202020202017 H20202020202020202020202018 H20202020202020202020202019 H20202020202020202020202020 H20202020202020202020202021 H20202020202020202020202022 H20202020202020202020202023 H20202020202020202020202024 H20202020202020202020202025 H20202020202020202020202026 H20202020202020202020202027 H20202020202020202020202028 H20202020202020202020202029 H202020202020202020202020200 H202020202020202020202020201 H202020202020202020202020202 H202020202020202020202020203 H202020202020202020202020204 H202020202020202020202020205 H202020202020202020202020206 H202020202020202020202020207 H202020202020202020202020208 H202020202020202020202020209 H2020202020202020202020202010 H2020202020202020202020202011 H2020202020202020202020202012 H2020202020202020202020202013 H2020202020202020202020202014 H2020202020202020202020202015 H2020202020202020202020202016 H2020202020202020202020202017 H2020202020202020202020202018 H2020202020202020202020202019 H2020202020202020202020202020 H2020202020202020202020202021 H2020202020202020202020202022 H2020202020202020202020202023 H2020202020202020202020202024 H2020202020202020202020202025 H2020202020202020202020202026 H2020202020202020202020202027 H2020202020202020202020202028 H2020202020202020202020202029 H20202020202020202020202020200 H20202020202020202020202020201 H20202020202020202020202020202 H20202020202020202020202020203 H20202020202020202020202020204 H20202020202020202020202020205 H20202020202020202020202020206 H20202020202020202020202020207 H20202020202020202020202020208 H20202020202020202020202020209 H202020202020202020202020202010 H202020202020202020202020202011 H202020202020202020202020202012 H202020202020202020202020202013 H202020202020202020202020202014 H202020202020202020202020202015 H202020202020202020202020202016 H202020202020202020202020202017 H202020202020202020202020202018 H202020202020202020202020202019 H202020202020202020202020202020 H202020202020202020202020202021 H202020202020202020202020202022 H202020202020202020202020202023 H202020202020202020202020202024 H202020202020202020202020202025 H202020202020202020202020202026 H202020202020202020202020202027 H2

FIG. 2



RBS

CAGGAAACAGCTATGACCATGATTACGCCAAGCTTGCATGCAAATTCTATTCAGGAGACAGTCATA ATG AAA TAC CTA
 -----> M K Y L

TMB3

TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC GCG GCC CAG CCG GCC ATG GCC GAG GTG TTT
 L P T A A A G L L L L A A Q P A M A E V F

XbaI **linker**
 GAC TAC TGG GGC CAG GGA ACC CTG GTC ACC GTC TCG AGC GGT GGA GGC GGT TCA GGC GGA GGT
 D Y W G Q G T L V T V S S G G G G S G G G

HIS-tag
 CAT CAT CAT CAC CAT CAC GGG GCC GCA
 H H H H H H G A A
 (insertion in pIT2 only)

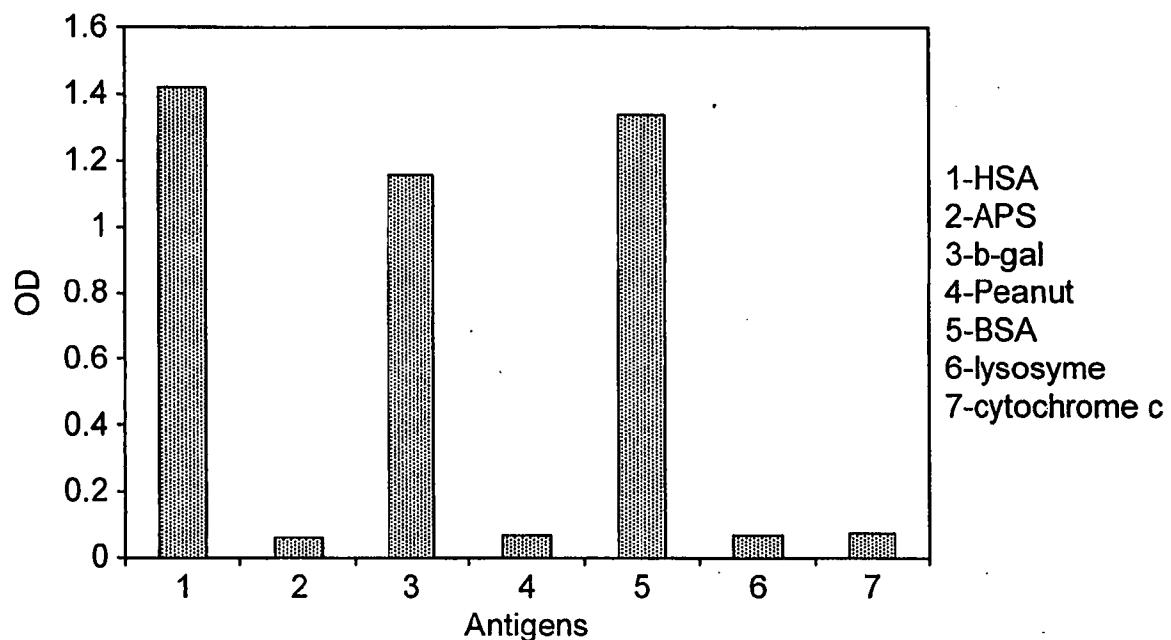
myc-tag *Gene III*
 ATC TCA GAA GAG GAT CTG AAT GGG GCC GCA TAG ACT GTT GAA AGT TGT TTA GCA AAA CCT CAT
 I S E E D L N G A A * T V E S C L A K P H
 <-----
 pHEN seq

3 / 17

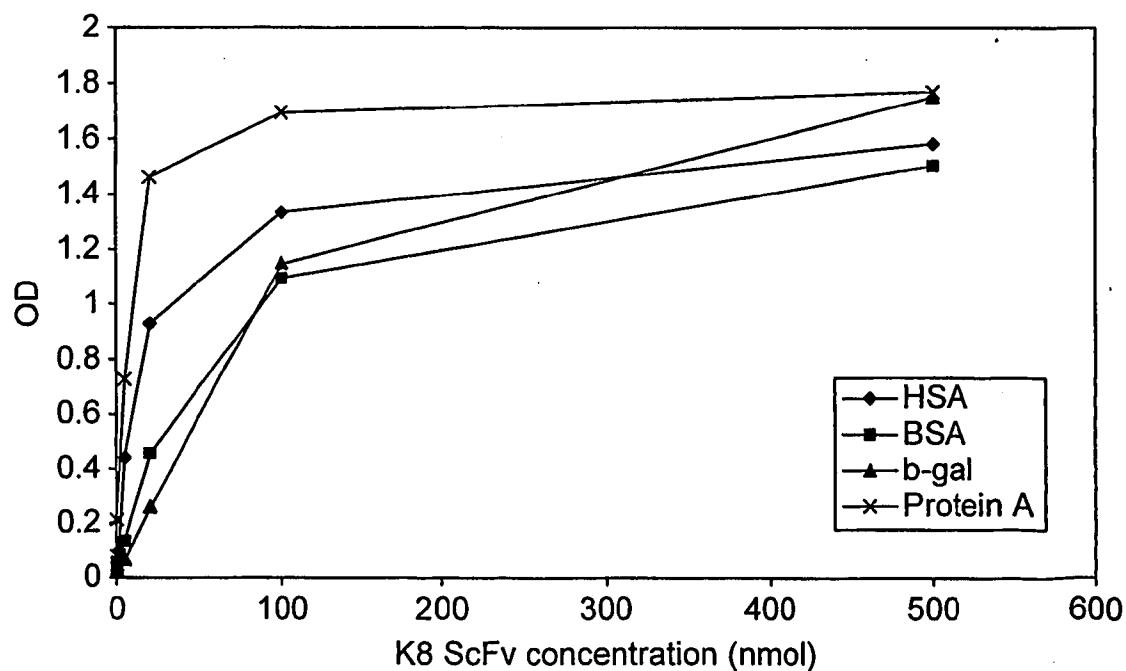
3
FIG

FIG. 4

Phage ELISA of a dual specific ScFv antibody K8

**FIG. 5**

Soluble ELISA of the Dual Specific ScFv Antibody K8



5 / 17

FIG. 6

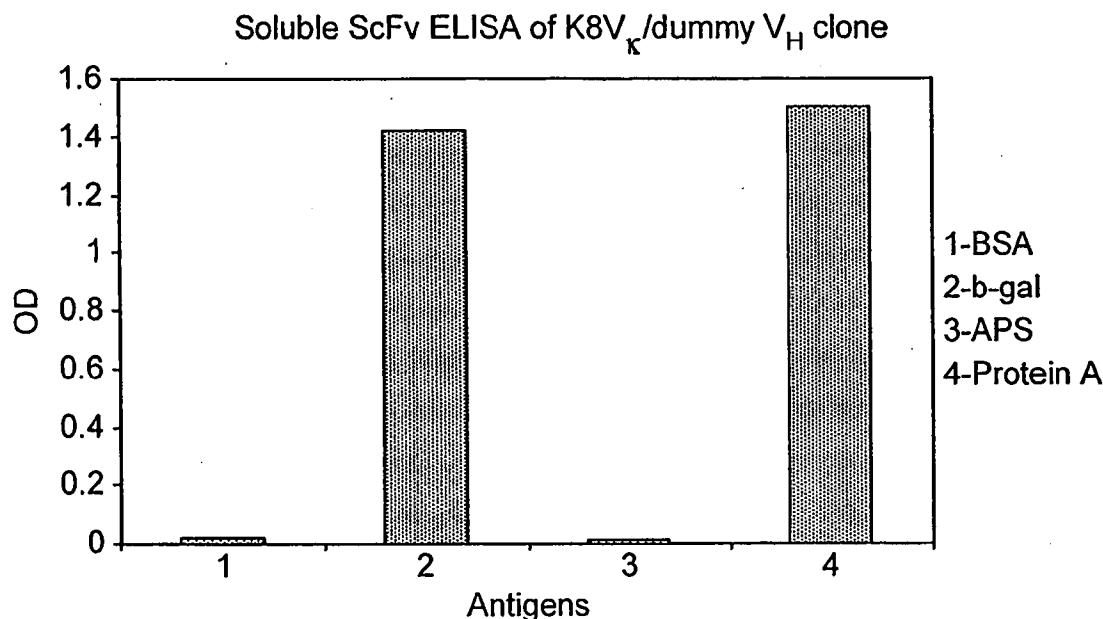


FIG. 7

RBS

CAGGAAACAGCTATGACCATGATTACGCCAAGCTTGCATGCAAATTCTATTTCAAGGAGACAGTCATA ATG AAA TAC CTA
-----> M K Y L

LMB3

SfiI NcoI

TTG CCT ACG GCA GCC GCT GGA TTG TTA CTC GCG GCC CAG CCG GCC ATG GCC GAG GTG TTT
L P T A A A G L L L A A Q P A M A E V F

XbaI linker

GAC TAC TGG GGC CAG GGA ACC CTG GTC ACC GTC TCG AGC GGT GGA GGC GGT TCA GGC GGA GGT
D Y W G Q G T L V T V S S G G G G S G G G

SalI NotI

GGC AGC GGC GGT GGC GGG TCG ACG GAC ATC CAG ATG ACC CAG GCG GCC GCA GAA CAA AAA CTC
G S G G G S T D I Q M T Q A A A E Q K L

link seq new

HIS-tag

CAT CAT CAT CAC CAT CAC GGG GCC GCA H H H H H G A A (insertion in V domain vector 2 only)

myc-tag Gene III

ATC TCA GAA GAG GAT CTG AAT GGG GCC GCA TAG ACT GTT GAA AGT TGT TTA GCA AAA CCT CAT
I S E E D L N G A A * T V E S C L A K P H

-----< pHEN seq

FIG. 8

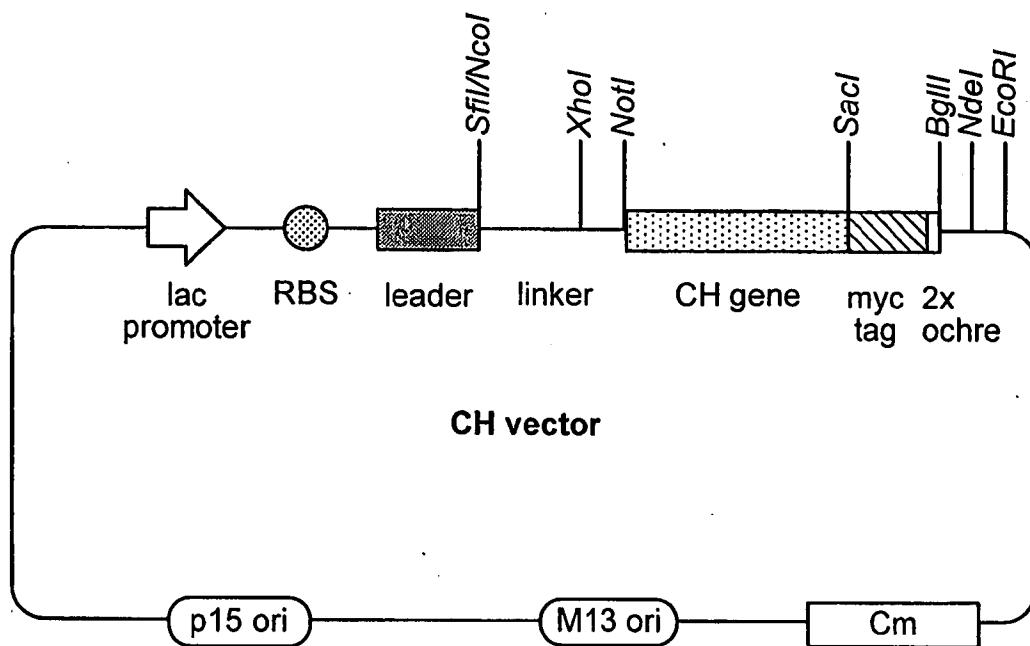


FIG. 9

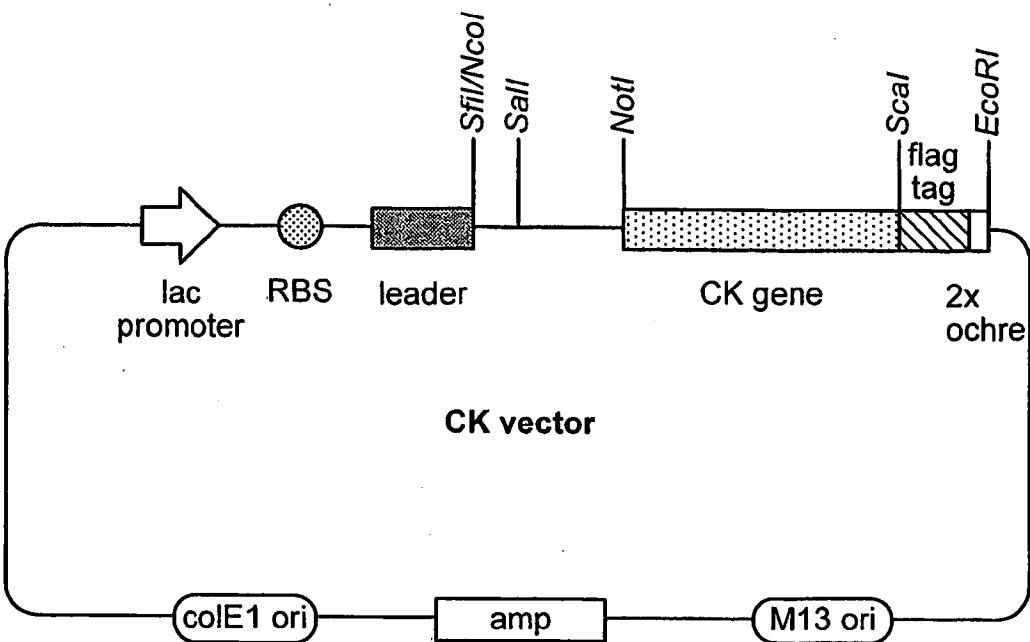


FIG. 10
TNF receptor assay

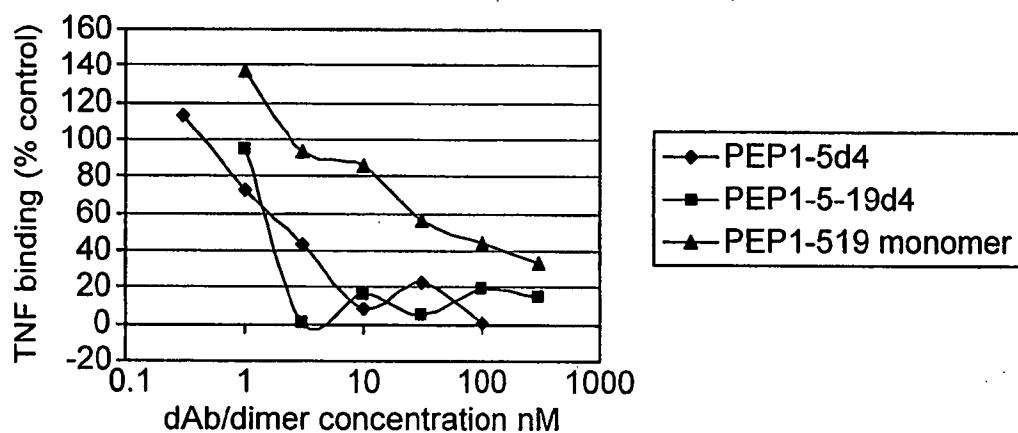


FIG. 11
TNF receptor assay

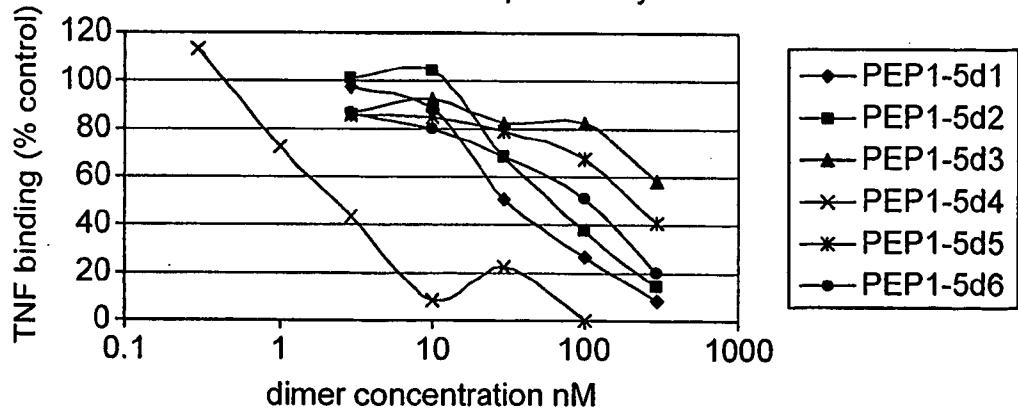


FIG. 12

TNF Receptor assay

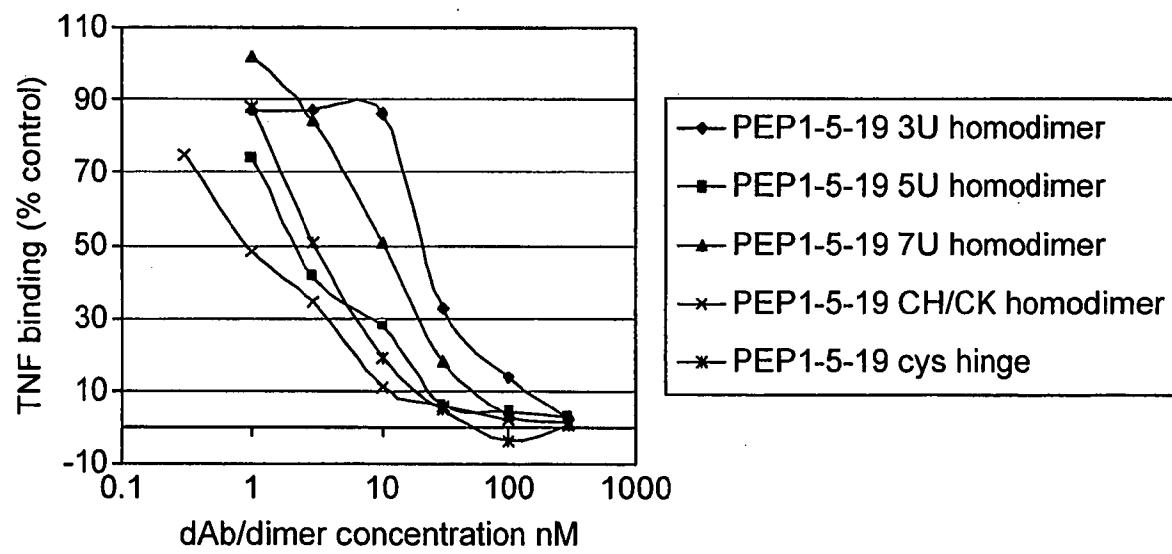


FIG. 13

Dummy V_H sequence for library 1

1 E V Q L L E S G G G L V Q P G G
 GAG GTG CAG CTG TTG GAG TCT GGG GGA GGC TTG GTA CAG CCT GGG GGG
 CTC CAC GTC GAC AAC CTC AGA CCC CCT CCG AAC CAT GTC GGA CCC CCC

49 S L R L S C A A S G F T F S S Y
 TCC CTG CGT CTC TCC TGT GCA GCC TCC GGA TTC ACC TTT AGC AGC TAT
 AGG GAC GCA GAG AGG ACA CGT CGG AGG CCT AAG TGG AAA TCG TCG ATA

97 A M S W V R Q A P G K G L E W V
 GCC ATG AGC TGG GTC CGC CAG GCT CCA GGG AAG GGT CTA GAG TGG GTC
 CGG TAC TCG ACC CAG GCG GTC CGA GGT CCC TTC CCA GAT CTC ACC CAG

145 S A I S G S G G S T Y Y A D S V
 TCA GCT ATT AGT GGT AGT GGT GGT AGC ACA TAC TAC GCA GAC TCC GTG
 AGT CGA TAA TCA CCA TCA CCA CCA TCG TGT ATG ATG CGT CTG AGG CAC

193 K G R F T I S R D N S K N T L Y
 AAG GGC CGG TTC ACC ATC TCC CGT GAC AAT TCC AAG AAC ACG CTG TAT
 TTC CCG GCC AAG TGG TAG AGG GCA CTG TTA AGG TTC TTG TGC GAC ATA

241 L Q M N S L R A E D T A V Y Y C
 CTG CAA ATG AAC AGC CTG CGT GCC GAG GAC ACC GCG GTA TAT TAC TGT
 GAC GTT TAC TTG TCG GAC GCA CGG CTC CTG TGG CGC CAT ATA ATG ACA

289 A K S Y G A F D Y W G Q G T L V
 GCG AAA AGT TAT GGT GCT TTT GAC TAC TGG GGC CAG GGA ACC CTG GTC
 CGC TTT TCA ATA CCA CGA AAA CTG ATG ACC CCG GTC CCT TGG GAC CAG

337 T V S S
 ACC GTC TCG AGC
 TGG CAG AGC TCG

FIG. 15

Dummy V_k sequence for library 3

1	D	I	Q	M	T	Q	S	P	S	S	L	S	A	S	V	G
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	GCA	TCT	GTA	GGA
	CTG	TAG	GTC	TAC	TGG	GTC	AGA	GGT	AGG	AGG	GAC	AGA	CGT	AGA	CAT	CCT
49	D	R	V	T	I	T	C	R	A	S	Q	S	I	<u>S</u>	<u>S</u>	<u>Y</u>
	GAC	CGT	GTC	ACC	ATC	ACT	TGC	CGG	GCA	AGT	CAG	AGC	ATT	<u>AGC</u>	<u>AGC</u>	<u>TAT</u>
	CTG	GCA	CAG	TGG	TAG	TGA	ACG	GCC	CGT	TCA	GTC	TCG	TAA	<u>TCG</u>	<u>TCG</u>	<u>ATA</u>
97	L	<u>N</u>	W	Y	Q	Q	K	P	G	K	A	P	K	L	L	I
	TTA	<u>AAT</u>	TGG	TAC	CAG	CAG	AAA	CCA	GGG	AAA	GCC	CCT	AAG	CTC	CTG	ATC
	AAT	<u>TTA</u>	ACC	ATG	GTC	GTC	TTT	GGT	CCC	TTT	CGG	GGA	TTC	GAG	GAC	TAG
145	Y	<u>A</u>	A	S	<u>S</u>	L	Q	S	G	V	P	S	R	F	S	G
	TAT	<u>GCT</u>	GCA	TCC	<u>AGT</u>	TTG	CAA	AGT	GGG	GTC	CCA	TCA	CGT	TTC	AGT	GGC
	ATA	<u>CGA</u>	CGT	AGG	<u>TCA</u>	AAC	GTT	TCA	CCC	CAG	GGT	AGT	GCA	AAG	TCA	CCG
193	S	G	S	G	T	D	F	T	L	T	I	S	S	L	Q	P
	AGT	GGA	TCT	GGG	ACA	GAT	TTC	ACT	CTC	ACC	ATC	AGC	AGT	CTG	CAA	CCT
	TCA	CCT	AGA	CCC	TGT	CTA	AAG	TGA	GAG	TGG	TAG	TCG	TCA	GAC	GTT	GGA
241	E	D	F	A	T	Y	Y	C	Q	Q	<u>S</u>	<u>Y</u>	<u>S</u>	<u>T</u>	P	<u>N</u>
	GAA	GAT	TTT	GCT	ACG	TAC	TAC	TGT	CAA	CAG	<u>AGT</u>	<u>TAC</u>	<u>AGT</u>	<u>ACC</u>	CCT	<u>AAT</u>
	CTT	CTA	AAA	CGA	TGC	ATG	ATG	ACA	GGT	GTC	<u>TCA</u>	<u>ATG</u>	<u>TCA</u>	<u>TGG</u>	GGA	<u>TTA</u>
289	T	F	G	Q	G	T	K	V	E	I	K	R				
	ACG	TTC	GGC	CAA	GGG	ACC	AAG	GTG	GAA	ATC	AAA	CGG				
	TGC	AAG	CCG	GTT	CCC	TGG	TTC	CAC	CTT	TAG	TTT	GCC				

FIG. 16

Nucleotide and amino acid sequence of anti MSA dAbs MSA 16 and MSA 26

A: MSA 16

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GAC ATC CAG ATG ACC CAG TCT CCA TCC TCC CTG TCT GCA TCT
D   I   Q   M   T   Q   S   P   S   S   L   S   A   S

GTA GGA GAC CGT GTC ACC ATC ACT TGC CGG GCA AGT CAG AGC
V   G   D   R   V   T   I   T   C   R   A   S   Q   S

ATT ATT AAG CAT TTA AAG TGG TAC CAG CAG AAA CCA GGG AAA
I   I   K   H   L   K   W   Y   Q   Q   K   P   G   K

GCC CCT AAG CTC CTG ATC TAT GGT GCA TCC CGG TTG CAA AGT
A   P   K   L   L   I   Y   G   A   S   R   L   Q   S

GGG GTC CCA TCA CGT TTC AGT GGC AGT GGA TCT GGG ACA GAT
G   V   P   S   R   F   S   G   S   G   S   G   T   D

TTC ACT CTC ACC ATC AGC AGT CTG CAA CCT GAA GAT TTT GCT
F   T   L   T   I   S   S   L   Q   P   E   D   F   A

ACG TAC TAC TGT CAA CAG GGG GCT CGG TGG CCT CAG ACG TTC
T   Y   Y   C   Q   Q   G   A   R   W   P   Q   T   F

GGC CAA GGG ACC AAG GTG GAA ATC AAA CGG
G   Q   G   T   K   V   E   I   K   R

```

B: MSA 26

```

GAC ATC CAG ATG ACC CAG TCT CCA TCC TCC CTG TCT GCA TCT
D   I   Q   M   T   Q   S   P   S   S   L   S   A   S

GTA GGA GAC CGT GTC ACC ATC ACT TGC CGG GCA AGT CAG AGC
V   G   D   R   V   T   I   T   C   R   A   S   Q   S

ATT TAT TAT CAT TTA AAG TGG TAC CAG CAG AAA CCA GGG AAA
I   Y   Y   H   L   K   W   Y   Q   Q   K   P   G   K

GCC CCT AAG CTC CTG ATC TAT AAG GCA TCC ACG TTG CAA AGT
A   P   K   L   L   I   Y   K   A   S   T   L   Q   S

GGG GTC CCA TCA CGT TTC AGT GGC AGT GGA TCT GGG ACA GAT
G   V   P   S   R   F   S   G   S   G   S   G   T   D

TTC ACT CTC ACC ATC AGC AGT CTG CAA CCT GAA GAT TTT GCT
F   T   L   T   I   S   S   L   Q   P   E   D   F   A

ACG TAC TAC TGT CAA CAG GTT CGG AAG GTG CCT CGG ACG TTC
T   Y   Y   C   Q   Q   V   R   K   V   P   R   T   F

GGC CAA GGG ACC AAG GTG GAA ATC AAA CGG
G   Q   G   T   K   V   E   I   K   R

```

FIG. 17

Inhibition biacore of MSA16

(a)

<u>0</u>	MSA competition 19. Fc=4-3 - 10	<u>10nM</u>	MSA competition 19. Fc=4-3 - 11	<u>50nM</u>	MSA competition 19. Fc=4-3 - 12
<u>100nM</u>	MSA competition 19. Fc=4-3 - 13	<u>200nM</u>	MSA competition 19. Fc=4-3 - 14	<u>300nM</u>	MSA competition 19. Fc=4-3 - 15
<u>400nM</u>	MSA competition 19. Fc=4-3 - 16	<u>600nM</u>	MSA competition 19. Fc=4-3 - 17	<u>800nM</u>	MSA competition 19. Fc=4-3 - 18

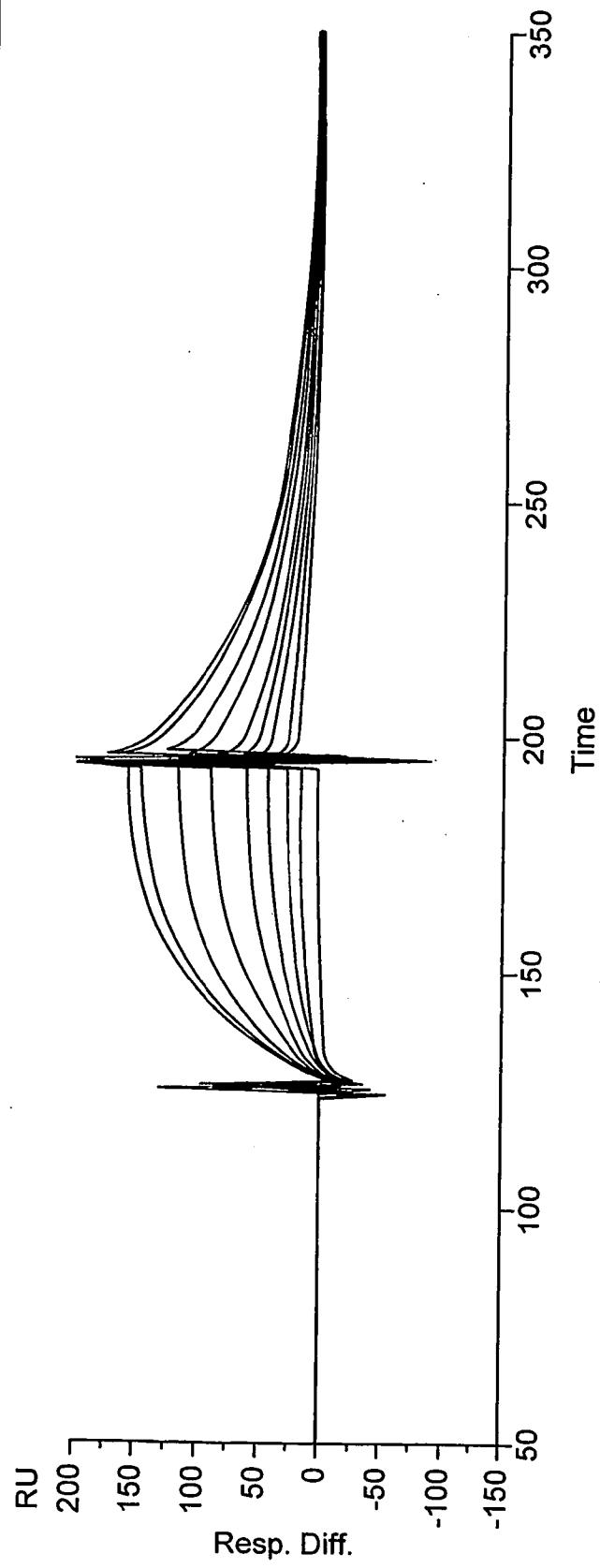


FIG. 17 CONT'D
(b) Inhibition biacore of MSA26

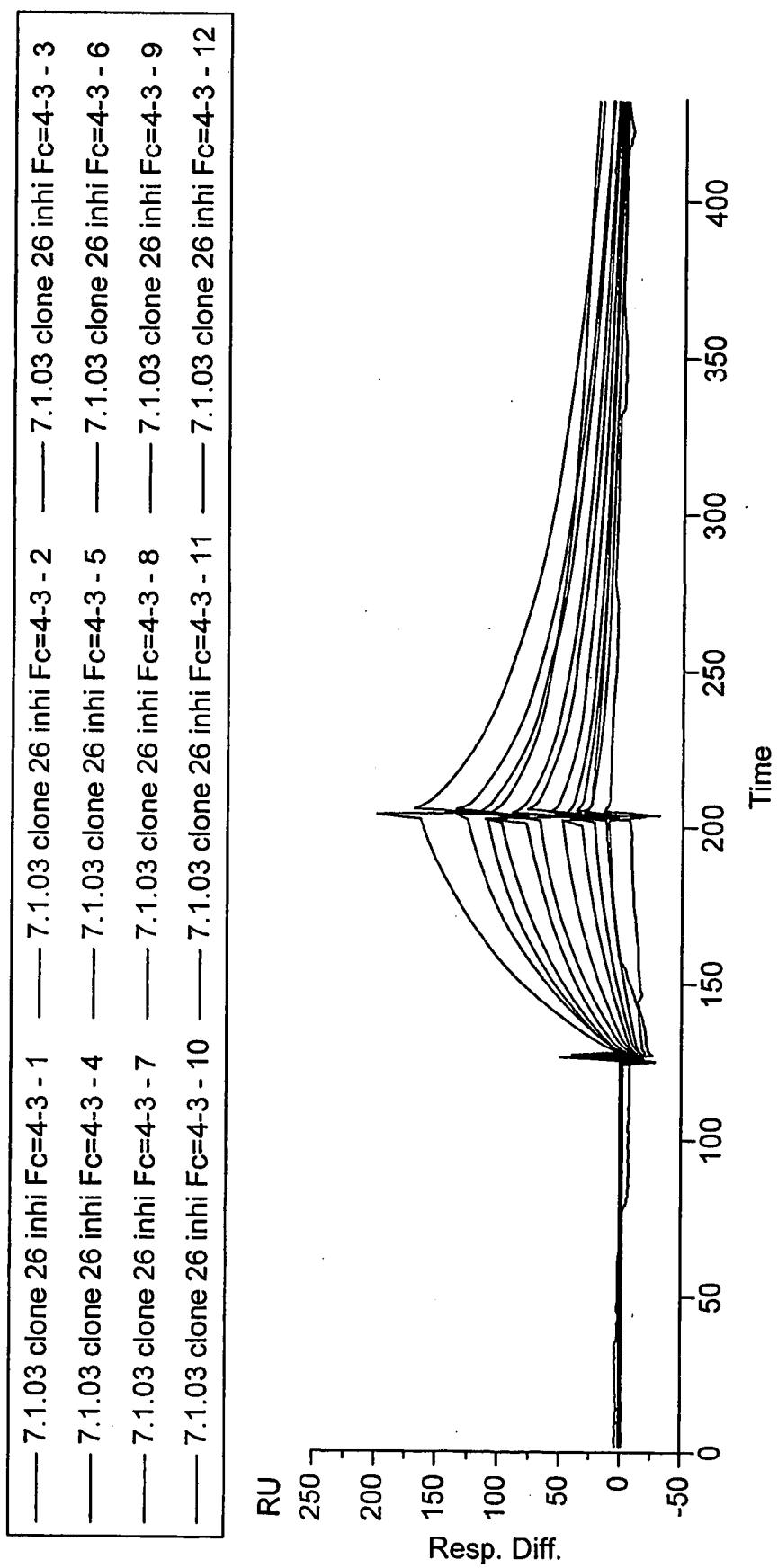
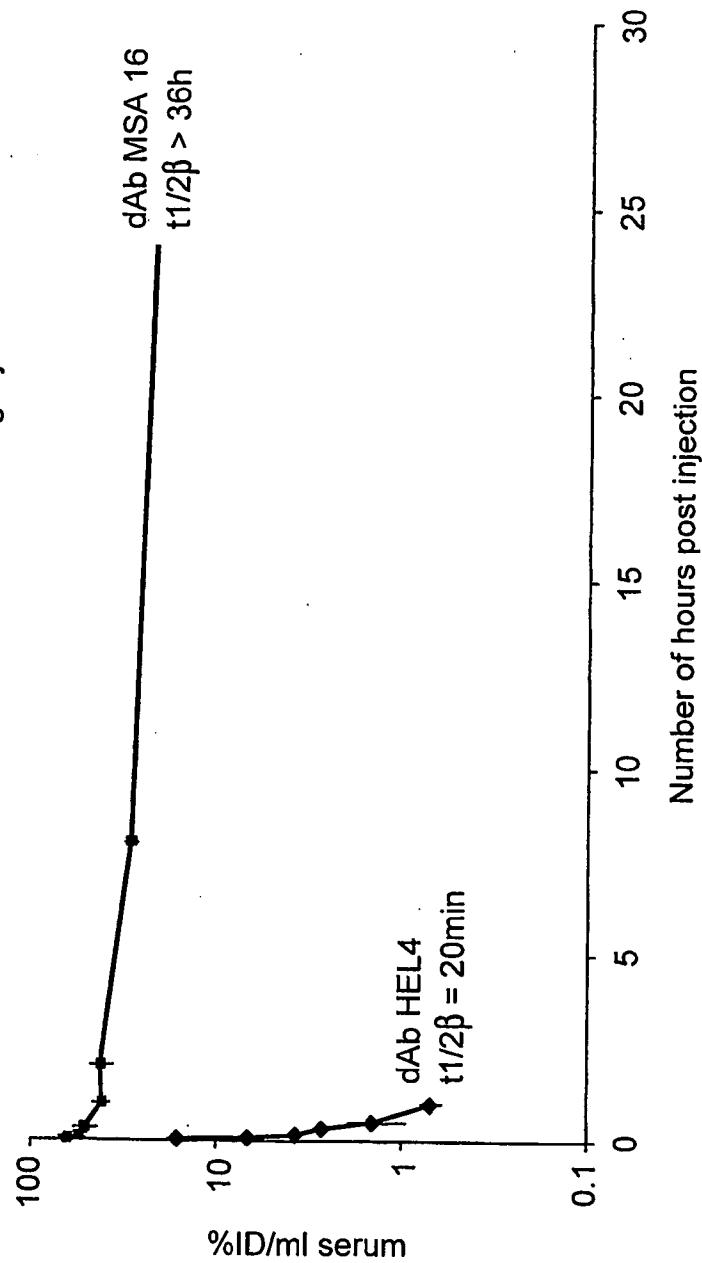


FIG. 18

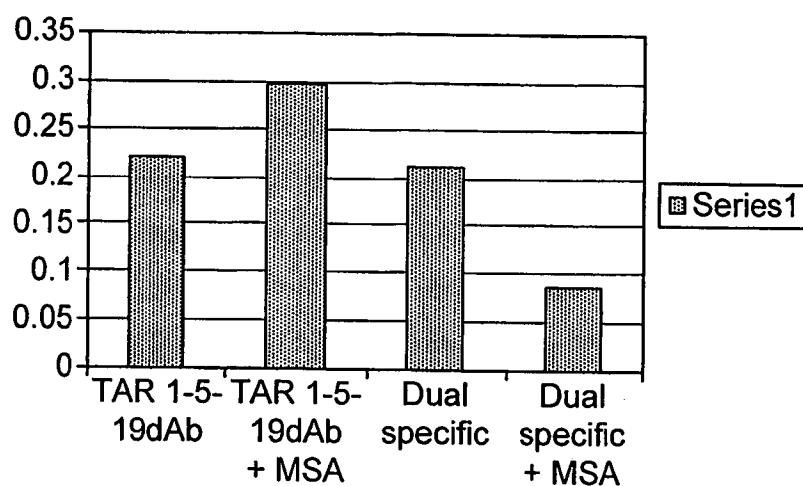
Serum levels of MSA16 following injection



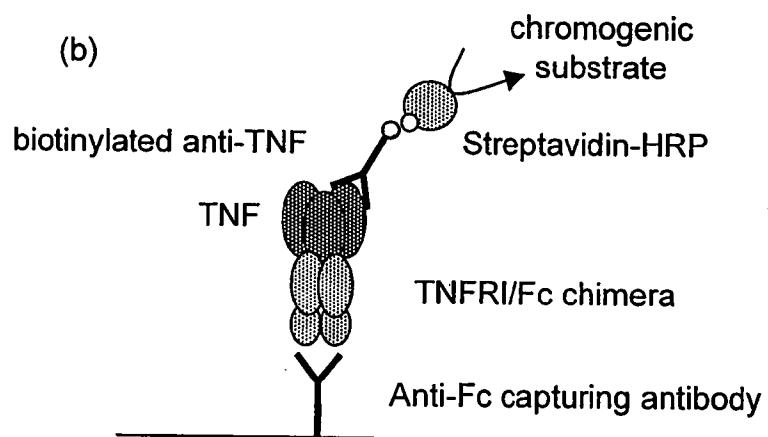
16 / 17

FIG. 19

(a)



(b)



(c)

TNF Receptor assay

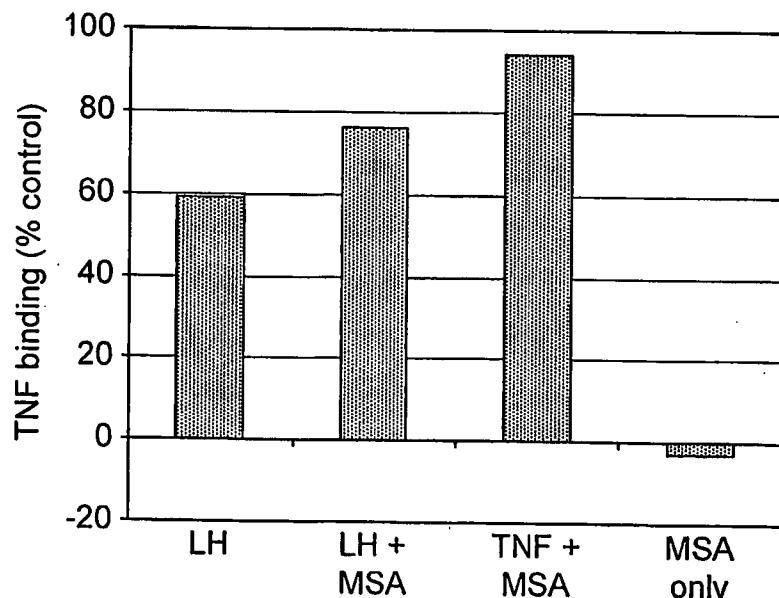
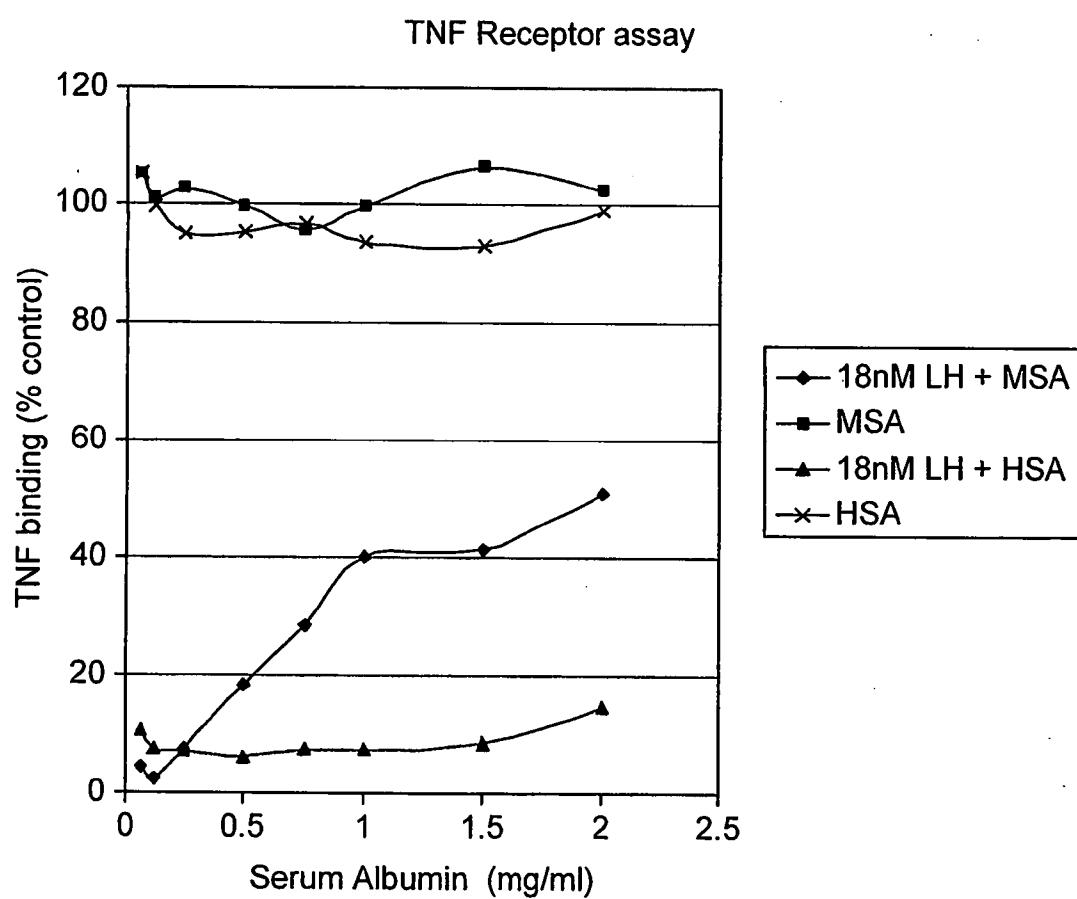


FIG. 20



SEQUENCE LISTING

5	E V Q L L E S G G G L V Q P G G S L R L S C A A S G F R I S D E D M .	GAGTCACGC TCTTCGACTC TGGGGAGCC TTGGTACAGC CTCGGGGTC CTCGGGATT TAGGATTGC GATGAGGATA
10	. G W V R Q A P G K G L E W V S S I Y G P S G S T Y X A D S V K G R .	TCGGCTGGT CGCCCTGGCTT CGCGGAAGG GCTCTAGAGTG GGTATCAAGC ATTATGGCC CTAGCGTAG CACATACGAC GCAGACTCCG TGAAGGGCG
15	. F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A S A L	GTCACCATC TCCCGGACA ATTCCAGAA CACGCTGTAT CTGCAATGA ACAGCTCG TGCGAGGAC ACCGCGGTAT ATTATGGCG GAGTCGCTTG
20	E P L S E P L G F W G Q G T L V T V S S	GAGCCGCTTT CGGAGCCCTT GGGCTTGGG GGTCAGGGAA CCCTGGTAC CGTCTCGAGC
25	E D F D Y W G Q G T L V T V S S	GAGGATTGG ACTACTGGG CCAGGGAAACC CTGGTCACCG TCTCGAGC
30	TAR1-5-19	D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S V K E F L W .
35	GGTGTGACCA GCAGAAACCA GGGAAAGCCC CTAAAGCTCTT GATCTATG GCATCCATT TGCAAAAGTG GGTCCCATCA CGTTCACTGTC GCGATGGATC	
40	GGGACCAAGG TGGAAATCAA ACGG	

TAR2-10
 GAGGTGCAGGCTGGAGGCTGGGGAGGGTACAGGCCCTGGGATTCCCTGTGAGCTGGCTCTCGGTCTCAGCTTCTAGGTAGCTGGTATTGGATGGGTGGTCCGC
 CAGGCTCCAGGGAGGGTCTAGAGTGGTCTCAGCTTCTAGGTAGCTGGTATTAGTGGTACTGGTGTAGCCACATACTACGAGACTCCGTGAAGGGCGGTCACCATCTCCCGGCCAACAAATCCAAGA
 ACACGGCTGATCTGCAAAATGAAACAGCCTGGTGTGGAGACCCGGAGAACCGGGCTAAATTACTGTGGAAAGTAAAGTTAGCTGGGGGGCTAAATTGTGACTACTGGGGGCCAGGGAAACCT
 GGTACCCGGTCTCGAGGCGCEYQLESGGGLVQPGGSLRLSCAASGFTTREWMGWVQAPGKGLEWVSAISGGGGSTYYADSVKGRFTISRDNSKNTLYLQMNSLRAEDTAVYYCA
 KVKLGGGPNNFDWNGQGTIVTVSS

10	1	E V Q L L E S G G G L V Q P G G S .
	51	GAGGTGCAGC TGGTGGAGTC TGGGGAGGC TTGGTACAGC CTGGGGGGTC
	101	CCTGGGTCTC TCCCTGTGAG CCTCCGGATT CACCTTGTGAG TCGGTATTGGCA G W V R Q A P G K G L E W V S A
	151	TGGGTGGGT CGGCCAGGCT CGAGGAAGG GTCTAGAGTG GGTCTAGCT I S G S G S T Y A D S V K G R
	201	ATTAGTGGTA GTGGTGGTAG CACATACTAC GCAGACTCCG TGAAGGGCCG F T I S R D N S K N T L Y L Q M N GTTCACCATC TCCCAGACA ATTCCAAGAA CACGCTGTAT CTGCAAATGA
	251	S L R A E D T A V Y C A K V K ACAGCCTGG TGCCGAGGAC ACCGGGTAT ATTACTGRC GAAAGTTAG L G G P N F D Y W G Q G T L V T
20	301	TTGGGGGGG GGCTTAATT TGACTACTGG GGCCAGGGAA CCTGGTCAC Ava I
	25	----- V S S CGTCTCGAGC

30 dAbs 1, 2 and 3 are the partner sequences from TAR1-5 dimers 1-6. dAb1 is the partner dAb to dimers 1, 5 and 6
 the second dabs are the same but the linker lengths are different, likewise, dAb2 is the partner dAb to dimers
 2 and 3, dAb3 is the partner dAb to dimer 4. * indicates the presence of an amber stop codon. Sequence homology
 between, TAR1-5, TAR1-5-19 and the unique second dabs is 88%
 35

TAR1-5-19
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I D S Y L H
 GACATCCAGA TGACCCAGTC TCCATCCCTCT CTGTCTGCACT CTGTAGGAGA CGGTGTCACT ATCACTGCC GGGCAAGTCA GAGGTTGAT AGTTATTAC
 CTGTAGGTCT ACTGGGTCAAG AGTGTAGGAGA GACAGACGTA GACATCCCTCT GGCAACAGGG TAGTGAACGG CCCGTTCACT CTGTTCACT
 W Y Q Q K P G K A P K L L I Y S A S E L Q S G V P S R F S G S

ATGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCTT GATCTATAGT GCATCCGAGT TGCAAACTGG GTCGACATCA CGTTTACGT GCAGTGGATC
 TAACCTGGT CCTCTTGGT CCCCCTGGG GATTGAGAA CTTAGATATCA CGTAGGCTAC AGTTTACCC CGAGGTAGT GCAAGTCAC CGTCACCTTG
 G T D F T L T Q S L Q P E D F A T Y C Q Q V W R P F T F G Q
 5 TGGACAGAT TTCACTCTCA CCATCGAGG TCTGCAACCT AGAGTGGAA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 ACCCTCTCA AAGTGGAGT GGTAGTGGTC AGACGTGGA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTTCCC ACCTTAGTT TGCG

10

TAR1-5
 D I Q M T Q S P S L S A S V G D R V T I T C R A S Q S I F M N L L
 GACATCCAGA TGACCGAGTC TCCATCCCTCC CGTGTGCAAT CTGTAGGAGA CGGTGTCACC ATCACTGCC GGGGAGTCA GAGGTTTTT ATGAAATTAT
 CTGTAGGTCT ACTGGTCTAG AGGTAGGG GACAGAGCTA GACATCCCTCT GGACAGCTGG TAGTGAACGG CGCGTTCACT CTGTAAAAA TACTTAATA
 W Y Q Q K P G K A P K L L I Y N A S V L Q S G V P S R F S G S G S
 TGTGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCTT GATCTATAGT GCATCCGAGT TGCAAACTGG GTCGACATCA CGTTTACGT GCAGTGGATC
 ACCCTCTCA AGTGGAGT GGTAGTGGTC AGACGTGGA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 G T D F T L T Q S L Q P E D F A T Y C Q Q V V W R P F T F G Q
 15 TGGACAGAT TTCACTCTCA CCATCGAGG TCTGCAACCT AGAGTGGAA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 ACCCTCTCA AAGTGGAGT GGTAGTGGTC AGACGTGGA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTTCCC ACCTTAGTT TGCG

25

Monomer - dab1: Partner dab in TAR1-5d1 (3U linker), d5 (5U linker), d5 (5U linker), d6 (7U linker)
 D I Q M T Q S P S L S A S V G D R V T I T C R A S Q S I Y D A L E
 GACATCCAGA TGACCGAGTC TCCATCCCTCC CGTGTGCAAT CTGTAGGAGA CGGTGTCACC ATCACTGCC GGGGAGTCA GAGGTTTTT GATGCGTTAG
 CTGTAGGTCT ACTGGTCTAG AGGTAGGG GACAGAGCTA GACATCCCTCT GGACAGCTGG TAGTGAACGG CGCGTTCACT CTGTAAATA CTACGATTC
 W Y Q Q K P G K A P K L L I Y T A S R L Q S G V P S R F S G S G S
 AGTGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCTT GATCTATAGT GCATCCGAGT TGCAAACTGG GTCGACATCA CGTTTACGT GCAGTGGATC
 30 TCACCTGTG CGTCTTGGT CCCTTCCGG GATTGAGAA CTTAGATATCA CGTAGGGCCA ACGTTTCAAC CGCGGGTAGT GCAGTGGAC CGTCACCTAG
 G T D F T L T Q S L Q P E D F A T Y C Q Q V M Q R P V T F G Q
 TGGACAGAT TTCACTCTCA CCATCGAGG TCTGCAACCT AGAGTGGAA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 ACCCTCTCA AAGTGGAGT GGTAGTGGTC AGACGTGGA CTTCTAAAC GATGATGAT GACAGTTGTC CGAGTGGAC GTCGGCCAA
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTTCCC ACCTTAGTT TGCG

35

Monomer - dab2: Partner dab in TAR1-5d2 (3U linker), d3 (5U linker)
 D I Q M T Q S P S L S A S V G D R V T I T C R A S Q S I Y D A L E
 GACATCCAGA TGACCGAGTC TCCATCCCTCC CGTGTGCAAT CTGTAGGAGA CGGTGTCACC ATCACTGCC GGGGAGTCA GAGGTTTTT GATGCGTTAG
 CTGTAGGTCT ACTGGTCTAG AGGTAGGG GACAGAGCTA GACATCCCTCT GGACAGCTGG TAGTGAACGG CGCGTTCACT CTGTAAATA CTACGAAATG
 W Y Q Q K P G K A P K L L I Y T A S R L Q S G V P S R F S G S G S
 AGTGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCTT GATCTATAGT GCATCCGAGT TGCAAACTGG GTCGACATCA CGTTTACGT GCAGTGGATC
 45 TCACCTGTG CGTCTTGGT CCCTTCCGG GATTGAGAA CTTAGATATCA CGTAGGGCCA ACGTTTCAAC CGCGGGTAGT GCAGTGGAC CGTCACCTAG

40

Monomer - dab2: Partner dab in TAR1-5d2 (3U linker), d3 (5U linker)
 D I Q M T Q S P S L S A S V G D R V T I T C R A S Q S I Y D A L E
 GACATCCAGA TGACCGAGTC TCCATCCCTCC CGTGTGCAAT CTGTAGGAGA CGGTGTCACC ATCACTGCC GGGGAGTCA GAGGTTTTT GATGCGTTAG
 CTGTAGGTCT ACTGGTCTAG AGGTAGGG GACAGAGCTA GACATCCCTCT GGACAGCTGG TAGTGAACGG CGCGTTCACT CTGTAAATA CTACGAAATG
 W Y Q Q K P G K A P K L L I Y T A S R L Q S G V P S R F S G S G S
 AGTGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCTT GATCTATAGT GCATCCGAGT TGCAAACTGG GTCGACATCA CGTTTACGT GCAGTGGATC
 45 TCACCTGTG CGTCTTGGT CCCTTCCGG GATTGAGAA CTTAGATATCA CGTAGGGCCA ACGTTTCAAC CGCGGGTAGT GCAGTGGAC CGTCACCTAG

G T D F T L T I S S L Q P B D F A T Y H C Q Q V M Q R P V T F G Q
 TGGACAGAT TTCACTCTCA CCATCAGAG TCTGCACCT GAAGATTTG CTAGTACCA CTGTCACAG GTTATGCCAG GTCCTGTAC GTTGGCCAA
 ACCCTGCTTA AAGTGAAGAT GTTACTGCTC AGACCTGGA CTTCTAACG GATGCTAGT GACAGTTC GAAATGTCG CAGGAAATG CAGGCCGGTT
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGTTCC ACCTTGTATT TGCC

Monomer - dab2: Partner dab in TAR1-5d4 (5U Linker)

10 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S V K E F L W
 GACATCCAGA TGACCCAGTC TCCATCCCTC CTGTCCTGCT ATCACCTGCC CGGCAAGTCA GAGGTTAAC GAGTTTTAT
 CTGTCAGTCT ACTGGGTCTAG AGGTAGGAGG GACAGACGTA GACATCCCTC GGACACAGTGG TAGTGAACGG CGCGTTCTAGT CTGCAATTCT CGCAAAATA
 W Y Q Q K P G K A P K L L I Y M A S N L Q S G V P S R F S G S G S
 GGTGGTACCA GGAGAAACCA GGGAAAGGCC CTAAGCTCT CTAAGCTCT GATCTATATG GCATCCAACT TGCAAAAGTGG GTCCTCCTAC CGTTCAGTGG GCAACTGGATC
 CCACATGGT CGTCCTGGT CCCTTTCGG GATTGAGGTA CTAGATATAC CGTAGGTTA ACGTTTCAAC CGAAGTCTAGT CGTACACTG CGTACACTG
 15 G T D F T L T I S L Q P C T A T Y Y C Q Q K P K L P R T F G Q
 TGGACAGAT TTCACTCTCA CCATCAGAG TCTGCACCT GAAGATTTG CTAGTACTA CTGTCACAG AAGTTAAC TGCCCTGTAC GTTGGCCAA
 ACCCTGCTTA AAGTGAAGAT GTTACTGCTC AGACCTGGA CTTCTAACG GATGCTAGT GACAGTTTC TCAAAATTG ACAGGAGTAC CAGGCGGTT
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGTTCC ACCTTGTATT TGCC

TAR1-27 and related partner dab amino acid and DNA sequences (* indicates the presences of an amber stop codon).
 25 Sequence homology between TAR1-27 and the unique second dabs is 90.4%.

TAR1-27

D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I W T K I I H
 GACATCCAGA TGACCCAGTC TCCATCCCTC CTGTCCTGCT ATCACCTGCC CGGCAAGTCA GAGGATTGG ACCAGATTAC
 CTGTCAGTCT ACTGGGTCTAG AGGTAGGAGG GACAGACGTA GACATCCCTC GGACACAGTGG TAGTGAACGG CGCGTTCTAGT CTGCAATTCT CGCAAAATA
 W Y Q Q K P G K A P K L L I Y M A S S L Q S G V P S R F S G S G S
 ATGGTACCA GGAGAAACCA GGGAAAGGCC CTAAGCTCT CTAAGCTCT GATCTATATG GCATCCAGT TGCAAAAGTGG GGTGCCCCATCA CGTTCAGTGG GCAACTGGATC
 TAACCTGCTGT CGTCCTGGT CCCCTTCCGG GATTGAGGTA CTAGATATAC CGTAGGTTA ACGTTTCAAC CGAAGTCTAGT CGTACACTG CGTACACTG
 30 G T D F T L T I S S L Q P E D F A T Y Y C Q Q W F S N P S T F G Q
 TGGACAGAT TTCACTCTCA CCATCAGAG TCTGCACCT GAAGATTTG CTAGTACTA CTGTCACAG TGTGTTAGTA ATCTAGTAC GTTGGCCAA
 ACCCTGCTTA AAGTGAAGAT GTTACTGCTC AGACCTGGA CTTCTAACG GATGCTAGT GACAGTTTC ACCAAATCAT TAGGATCATG CXAGGCCGGTT
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGTTCC ACCTTGTATT TGCC

Partner dab monomer in TAR1-27d1 (3U Linker)

D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I * P I L C
 GACATCCAGA TGACCCAGTC TCCATCCCTC CTGTCCTGCT ATCACCTGCC CGGCAAGTCA GAGGATTGG CGCGTTCTAGT CTGCAATTCT CGCAAAATA
 CTGTCAGTCT ACTGGGTCTAG AGGTAGGAGG GACAGACGTA GACATCCCTC GGACACAGTGG TAGTGAACGG CGTCAAAATC GGCTAAATA
 W Y Q Q K P G K A P K L L I Y A A S S L Q S G V P S R F S G S G S

GTGGGTACCA GCGAGAACCA GGGAAAGGCC CTAAGCTCT GATCPATGCT GCATCCPAGT
 CAACCATGCT CGTCCTTGCTT GCTCTTGGG GATTGCGGAA CTAGATACGA CGTAGGGCTAA ACGTTCACCC
 G T D F T L T I S S L Q P E D F A T Y Y C Q Q I Q H I P V T F G Q
 GGGACAGAT TTCACTCTCA CTCATCAGCAG TCTGCAACCT GAAGATTTCG CTAGTACTAA CTGTCAACAGS ATTACGATTA
 CTCCTGCTTA AAGTGAAGT GGTAGTCGTC AGACGTGGA CTCCTAAAC GATGCTGATG GTCAGTGTGTC ATTACGATTA
 G T K V E I K R

partner dAb monomer in TAR1-27d2 (3U linker)
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I G * D L H
 GACATCCAGA TGACCCAGTC TCCATCCTCC CTCCTGCTCAT CTCCTGAGA CGCTAGGAGA ATCTACTGCC GGGCAAGTC GAGCATTTGGG TAGGATTAC
 CTGTTAGTCAT ACTGTTAGC AGCTTGGGG GACCTGCTCAT GACATCTCT GGCACATCTGG TAGTGTACGG CCGGTTACCC ATCTAAATG
 W Y Q Q K P K L L I Y T A S L L Q S G V P S R F S G S G S
 ATGGTACCA GCAGAAACCA GCGAAAGGCC CTAAGCTCT GATCTTATAG GCATCCCTT TGCCTAGTGS GCTTTCATGTG GCAGTGGATC
 TRACCATGGT CGTCCTGGT CCTTCTGGG GATTCGAGA CTAGATATGC CGTAGGAAA ACCTTCTCC CCAAGGTGATC CTCACCTAG
 G T D F T L T I S S L Q P E D F A T Y Y C Q Q Q S A F P N T L G Q
 TGGGACAGAT TTICACTCTCA CCTTCAGCAG TCTGCACT GAAAGATTTT CTAGCTATA CTGTCAGCAG CAGAGTCCTTAC GCTCCGCTTA
 ACCCTGTTCA AAGTGAGAT GGTAGTC GTCAGCTGC AGACGTTGGA CTTCCTAAC GATGCCAT GACAGTTGTC GTCTCACGAA AAGGATTATG CGAGCCGGTT

25 Partner dAb monomer in TARI-27d7 (3' linker)
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I T K N L L
 GACATCGAGA TGACCCAGTC TCCATCCTCC CTCCTCGAT CGCTAGGAGA CGGCAAGTCG ATCACCTGCC CCGTGTCCAC
 CTCCTAGGCTC ACTGGTCA GAGTAGGGG GACAGAGCTA GGCACTCTCT GGCACTAGTG TAGTGAAAGGG CCCGTTCACT CTCCTAAATG
 30 W Y Q K P G K A P K L L I Y * A S S L Q S G V P S R F S G S G S
 TTGGTACCA GCGAAACCA GGGAAAGCCC CTAAGCTCT GATCTATTAG GCATCCCTCT TGCAAAAGTGG GGTCCCACTCA CTTTCAGTGC
 AAACCTGGT CTCCTGGT CCCTTCGGG GATTGAGGA CTAGATATTC CGTAGGAGAA ACGTTCACCC CCAAGGTGAT GCAAGTCAC CGTCACTCTAG
 G T D F T L T I S L Q P B D F A T Y Y C Q Q L R H K P P T F G Q
 TGGGACAGAT TTCTCTCA CCTATCGAGA TCCTGCAACTA GAGAGTTCTA CTCTGACTA CCTGTCACAG CCTGTCACAG CCTGTCACAG CCTGTCACAG
 ACCCTGCTCA AAGTAGAGT GTCAGTGTCTA GTCAGTGTCTA GTCAGTGTCTA GTCAGTGTCTA GTCAGTGTCTA GTCAGTGTCTA GTCAGTGTCTA
 35 G T K V E I K R
 GGACCAAGG TGGAAATCAA ACGG
 CCCTGGTCC ACCTTGTGTT TGCC

40 Partner dab monomer in TAR1-27d8 (3D linker) 45

D	I	Q	M	T	Q	S	P	S	L	S	A	S	V	G	D	R	V	T	I	T	C	R	A	S	Q	S	I	*	K	S	L	R
G	R	A	T	G	C	T	C	T	T	C	T	G	A	G	T	T	G	A	G	T	C	G	G	C	T	G	T	A	T	A		
C	T	G	T	C	T	G	A	T	C	T	G	A	G	T	T	G	A	G	T	C	G	G	C	T	G	T	A	T	A			
G	T	A	T	C	T	G	A	T	C	T	G	A	G	T	T	G	A	G	T	C	G	G	C	T	G	T	A	T	A			
C	T	G	T	C	T	G	A	T	C	T	G	A	G	T	T	G	A	G	T	C	G	G	C	T	G	T	A	T	A			
C	T	G	T	C	T	G	A	T	C	T	G	A	G	T	T	G	A	G	T	C	G	G	C	T	G	T	A	T	A			

G T D P T L T I S S L Q P E D F A T Y C Q Q M V N S P V T F G Q
 TCTGCGAGAT TTCACTCTCA CTCATCGAGC TCTGCAACCT GAAAGATTG CTCGCTACTA CTCGTCACAG ATGGTGTATA GTCCTGTTAC GTCGGCCCAA
 CCGCTGCTCA AAGCTGAGT GTTAGTGTCTC AGACGTGGA CTCTCTAAAC GATGCTGTC TACCATGAT CAGGACAATG CAGCGCGTT
 T K V E I K R

5

5	Partner dab monomer in TARI-27d16 (3U linker)	D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I G . P N L E GACATCCAGA TGACCCAGTC TCATCCCTCC CTGTCTGCA TCTGAGGAGA CGGCAAGTC AATCACTGCC GGGCAAGTC GAGCATGGG CGCAATTAG CTGTAGCTCT ACTGGCTCAG AGGTAGGGG GACAGACGTA GACATCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT W Y Q O K P G K A P K L I Y A A S S L Q S L Q S G V P S R F S G S G S AGTGGTACCA GCAGAAACCA GGAAAGGCC CTAAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT TCACCTGGT CGTCCTGGT CCCTTCGGG GATTCGAGGA CTAGATAGCA CGTGGTCA AGCTTCACCA AGCTTCACCA AGCTTCACCA AGCTTCACCA G T D F T L T I S S L Q P E D F A T Y Y C O Q Q M G R P R T F G Q TGGGAGAT TTCACTCTCA CCATCAGCAG TCTGCAACT GAGATTTG CTAGCTACTA CTGTCAACAG CAGATGGGGC GTCTCGGCAG GTTCGGCCAA ACCCCTCTCA AAGTAGAGT GGTAGTCAG AGACGTTGGA CTCTAAAC GATGCTGTC GACAGCTCTC GTCTACCCCG CAGGAGCTG CAAAGCCGTT G T K V E I K R GGGACCAAGG TTGAAATCA ACGG CCCTGGTCC ACCTTGTAGT TGGC
10	Partner dab monomer in TARI-27d23 (5U linker)	D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I K H * L A GACATCCAGA TGACCCAGTC TCATCCCTCC CTGTCTGCA TCTGAGGAGA CGGCAAGTC AATCACTGCC GGGCAAGTC GAGCATGGG CGCAATTAG CTGTAGCTCT ACTGGCTCAG AGGTAGGGG GACAGACGTA GACATCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT W Y Q O K P G K A P K L I Y K A S V L Q S L Q S G V P S R F S G S G S CTTGGTACCA GCAGAAACCA GGAAAGGCC CTAAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GAACCTGGT CGTCCTGGT CCCTTCGGG GATTCGAGGA CTAGATAGCA CGTGGTCA AGCTTCACCA AGCTTCACCA AGCTTCACCA AGCTTCACCA G T D F T L T I S S L Q P E D F A T Y Y C O Q Q M G R P R T F G Q L R R R P T T F G Q
15	Partner dab monomer in TARI-27d23 (5U linker)	D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I K H * L A GACATCCAGA TGACCCAGTC TCATCCCTCC CTGTCTGCA TCTGAGGAGA CGGCAAGTC AATCACTGCC GGGCAAGTC GAGCATGGG CGCAATTAG CTGTAGCTCT ACTGGCTCAG AGGTAGGGG GACAGACGTA GACATCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT W Y Q O K P G K A P K L I Y K A S V L Q S L Q S G V P S R F S G S G S CTTGGTACCA GCAGAAACCA GGAAAGGCC CTAAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GGTAGCTCT GAACCTGGT CGTCCTGGT CCCTTCGGG GATTCGAGGA CTAGATAGCA CGTGGTCA AGCTTCACCA AGCTTCACCA AGCTTCACCA AGCTTCACCA G T D F T L T I S S L Q P E D F A T Y Y C O Q Q M G R P R T F G Q L R R R P T T F G Q

TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAGGATTG CTAGCTACTA CTGTCACAG CTAGGCCTAC GTOCTACTAC GTGGGCCAA
 ACCCTGCTCA AAGTGAGAGT GGTAGCTGTC AGACCTGGA CTTCTAAAC GATGCTATG GACAGTGTG GAGGATGATG CAGGCGGT
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGGTTC ACCTTGTGTT TGCC

5

10 Partner dab monomer in TAR1-27d30 (7U linker)
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S V K A * L T
 GACATCCGAA TGACCCGTC TCCATCCTTC CGTCTGCTAT CTGTAGGAGA CGCTGTCAACG ATCACTGCC GGGCAAGTCA GAGGGTTAAG GCTTAGTAA
 CTGTAAGCTCT ACTGGCTCTAG AGGTAGGAGG GACAGACCTA GACATCCCTCT GGACAGTGG TAGTGAACGG CCCGTTCACT CTGCGAATTG CGATCAATT
 W Y Q K P G K A P K L L I Y K A S T L Q S G V P S G R F S G S
 CTGTTGACCA GCGAGAACCA GGGAAGGCC CTAAGCTCTT GATCTACTAG GATCTCCTCTT GCGAGCTCTCA CGTTTCAGTG GCGAGGAGTC
 GAACCATGCT CGCTCTTGTGTC CCCTTTCGGG GATTGAGGA CTAGATATTG CGTAGGTGAA AGCTTCAACCC CGTTCACTCTG GCGAGCTCTAG
 G T D F T L T I S S L Q P E D F A T Y Y C Q Q H S S R P Y T F G Q
 TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAGGATTG CTAGCTACTA CTGTCACAG CTTAGTCTA GGCCTTATAC GTTCGGCCAA
 ACCCTGCTCA AAGTGAGAGT GGTAGCTGTC AGACCTGGA
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGGTTC ACCTTGTGTT TGCC

20

25 Partner dab monomer in TAR1-27d31 (7U linker)
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I E N R L G
 GACATCCGAA TGACCCGTC TCCATCCTTC CGTCTGCTAT CTGTAGGAGA CGCTGTCAACG ATCACTGCC GGGCAAGTCA GAGGGTTAAG ATCCGGTTAG
 CTGTAAGCTCT ACTGGCTCTAG AGGTAGGAGG GACAGACCTA GACATCCCTCT GGACAGTGG TAGTGAACGG CCCGTTCACT CTGCGAATTG CTAGCCAAATC
 W Y Q K P G K A P K L L I Y * A S L L Q S G V P S G S R F S G S
 GTGGTACCA GCGAGAACCA GGGAAGGCC CTAAGCTCTT GATCTACTAG CGTCTCTGTG TGCAAGTGG GGTGCCCCATCA CGTTCACTTG GCGAGGAGTC
 CAACCATGCT CGCTCTTGTGTC CCCTTTCGGG GATTGAGGA CTAGATAATC CGCAGGAAACA AGCTTCAACCC CGAGGGTGTG GCGAGTCACTCG
 G T D F T L T I S S L Q P E D F A T Y Y C Q Q D S Y F P R T F G Q
 TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAGGATTG CTAGCTACTA CTGTCACAG GATTCGTTATTCCTGTTAC GTTCGGCCA
 ACCCTGCTCA AAGTGAGAGT GGTAGCTGTC AGACCTGGA CTTCTAAAC GATGCTATG GACAGTGTG CTTAGCTAA AAGGAGCAT GAAAGCTGTT
 G T K V E I K R
 GGGACCAAGG TGGAATCAA ACGG
 CCCCTGGTTC ACCTTGTGTT TGCC

35

40 Partner dab monomer in TAR1-27d36 (7U linker)
 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I M D K L K
 GACATCCGAA TGACCCGTC TCCATCCTTC CGTCTGCTAT CTGTAGGAGA CGCTGTCAACG ATCACTGCC GGGCAAGTCA GAGGGTTAAG GATAAGTAA
 CTGTAAGCTCT ACTGGCTCTAG AGGTAGGAGG GACAGACCTA GACATCCCTCT GGACAGTGG TAGTGAACGG CGCTTCACT CTGCGAATTG CTAGCAATT
 W Y Q K P G K A P K L L I Y * A S I L Q S G V P S R F S G S G S
 AGTGGTACCA GCGAGAACCA GGGAAGGCC CTAAGCTCTT GATCTACTAG CGTCTCTGTG GGTGCCCCATCA CGTTCACTTG GCGAGGAGTC
 TCACCATGCT CGCTCTTGTGTC CCCTTTCGGG GATTGAGGA CTAGATAATC CGAGGGTGTG GCGAGTCACTCG
 G T D F T L T I S S L Q P E D F A T Y Y C Q Q D S G G P N T F G Q

45

5' TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAAGATTG CTACGACTA CTGTCACAG GATAGTGGGG GTCCTAATAC GTTCGGCCAA
ACCTGGCTTA AAGTGAGAT GGTAGTCGTC AGACGTGGA CTCTAAAC GATGCTGAT GACGATGAT GATACCCC CAGGATGAT CAAGCCGGTT
G T K V B I K R
GGGACCAAGG TGGAAATCAA ACGG
CCCTGGTTC ACCTTAGTT TGCC

Partner dab monomer in TAR1-27d37 (7U linker)

10 D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I G R N L E
GACATCCAGA TGACCCAGTC TCCATCCTTC CTCCTCTC CTGAGGAGA CGCAGCTCAT CTGAGGAGA CGCAGCTCAT
CTGAGGCT ACTGGTCTG AGGTAGGG AGCAGAGTA GACATCTCT GGAGCTGAGG TAGTGAACGG CCGGTCTAGT CTCGTAACCC TCTTAAATC
W Y Q Q K P G K A P K L I Y D A S H L Q S G V P S R F S G S G S
AGTGGTACCA GCGAAACCA GCGAAAGCC CTTAGCTCT GATCATGAT GCATCCATT TGCAAAAGTG GGTCCTACCT TGCAAAAGTG
TTCACCTGT CGCTCTGGT CCCTTGGG GATTCGAGA CTAGTACTA CGTAGGGTA ACGTTTACCC CGAGGGTAGT GCAAGTCA CGTCACCTAG
G T D F T L T I S S L Q P E D F A T Y Y C C Q Q S R W L P R T F G Q
TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAAGATTG CTACGACTA CTGTCACAG TGCTGTGGC TTCCCTGTAC GTTCGGCCAA
ACCTGGCTTA AAGTGAGAT GATACCCC CAGGATGAT GACGATGAT GACGATGTC AGCGCAACG AAGGAGCTG CAGGCGGGTT
G T K V B I K R
GGGACCAAGG TGGAAATCAA ACGG
CCCTGGTTC ACCTTAGTT TGCC

25 Partner dab monomer in TAR1-27d39 (7U linker)

D I Q M T Q S P S S L S A S V G D R V T I T C R A S Q S I R K M L V
GACATCCAGA TGACCCAGTC TCCATCCTTC CTCCTCTC CTGAGGAGA CGCAGCTCAT CTGAGGAGA CGCAGCTCAT
CTGAGGCT ACTGGTCTG AGGTAGGG GACATGAT GACATCTCT GGAGCTGAGG TAGTGAACGG CCGGTCTAGT CTCGTAACCC TCTTAAATC
W Y Q Q K P G K A P K L I Y R A S Y L Q S G V P S R F S G S G S
TTCGAGGATCA GCGAAACCA GCGAAAGCC CTTAGCTCT GATCATGAGG TGCAAAAGTG GGTCCTACCT TGCAAAAGTG
AACCATGGT CGCTCTGGT CCCTTGGG GATTCGAGA CTAGTACTA CGTAGGGTA ACGTTTACCC CGAGGGTAGT GCAAGTCA CGTCACCTAG
G T D F T L T I S S L Q P E D F A T Y Y C C Q Q A F R R P R T F G Q
TGGGACAGAT TTCACTCTCA CCATCAGGAG TCTGCAACCT GAAGATTG CTACGACTA CTGTCACAG GCTTGTGGC GCCTCTAGGAC GTTCGGCCAA
ACCTGGCTTA AAGTGAGAT GATACCCC CAGGATGAT GACGATGTC AGCGATGTC CGAAAGGCC CGGATCTG CAGGCGGGTT
G T K V B I K R
GGGACCAAGG TGGAAATCAA ACGG
CCCTGGTTC ACCTTAGTT TGCC

40 TAR2 dab amino acid and DNA sequences (*) indicates the presences of an amber stop codon) sequence homology
between TAR2h-5 and the unique second dabs is 83.5%

TAR2h-5

E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T P D L Y N M
GAGGTGAGGAGTC TTGGTGGAGTC TTGGTACAGC CTGGGGGGTC CCTGGGTCTC TCCTGTGAGG CACCTTGAT CTTATAATA
CTCCCACTCTCA ACCTACCTCA ACCCCCTCG AACCATGTCG GACCCCTCG AGACACGTG AGACGCAAG AGGGGCTAA GTGGAAACTA GAAATATTAT
F W V R Q A P G K G L E W V S F I S Q T G R L T W Y A D S V K G R

TGTCTGGGT CGCCCGAGGT CCAGGGAGG GTCTAGAGT GGTCTCATTT ATTAGTCGA CTGGTAGGTT TACATGGTAC CGAGACTTCG TGAAGGGCCG
 ACAAACCCA GCGCGTCGGA GTCGCCCTTC CAGATCTAC CCAGAGTAA TAATCAGTCA GACCATCGGA ATGTTACCATG CTCCTGAGG ACCTTCGGGC
 F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y C A K T L
 GTCACCATC TCCCGGACA ATTCCAGAA CACGCTGTAT CTGCAATATGA ACAGGCTGG TGCCGAGGAC ACCGGGTAT ATTACTGTCG GAAAACGCTG
 CAAGCTGTAG AGGGCTGTAT TAAGGTCTT GTEGCTGTAT GACGTTACT TGTGCTGTAG TGCCGAGGAC AGGGCTCCG TGGCCATATAATGACAGC
 E D F D Y W G Q G T L V T V S
 GAGGTTTG ACTACTGGG CCAGGAACCG CTGGTACCCG TCTCG
 CTCCTAAC TGATGACCC CGTCCTCG GACCACTGGC AGAGC

5

10 Partner dab monomer in TAR2h-5d1 (3U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F P V Y M M
 GAGGTGAGC TTGGTACCG CTCGGGGGTC CCTGCGTC TCCTGTCAG CCTCCGGATT CACCTTTTCG GTTTATGTA
 CTCACCTCG ACAAACCTAG ACCCCCTCG AACCCTCTCG GACCCCGAGG AGCACAGTC GAGACGCTAA GTCGAAAGGC CAAATATGTA
 G W V R Q A P G K G L E W V S S I D A L G G R T G Y A D S V K G R
 TGGCTGGGT CGGCCAGGT CCAGGGAGG GTCTAGAGT GGTCTCATCG ATTGTATGTC GACAGGTTAC GCAAGCTTCG TGAAGGGCCG
 ACCCAACCA GGCAGCTCGA GTCGCCCTTC CAGATCTAC CCAGAGTACG TAATACAGG AACCAACCGC CTGTCGATG CGTGTGAGG ACCTTCGGGC
 F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y C A K T M
 GTTCACCATC TCCCGGACA ATTCCAGAA CACGCTGTAT CTGCAATATGA ACAGGCTGTAG TGCCGAGGAC ACCGGGTAT ATTACTGTCG GAAAACGCTG
 CAAGCTGTAG AGGGCTGTAT TAAGGTCTT GTEGCTGTAT GACGTTACT TGTGCTGTAG TGGCCATATAATGACAGC CTTCGATAC
 S M K T H T F D Y W G Q G T L V T V S
 TCGAATAAGA CGCATAGTT TGTACTCTG GGCAGGGGA CCCTGGTAC CGTCTCG AGCTTATCT
 AGCTTATCT CGCTATGACCC ACTGATGACCC CGCTATGACCA AGCTTATCT
 GCTATGACCA

20

25 Partner dab monomer in TAR2h-5d2 (3U linker)
 E V Q L L B S G G G L V Q P G G S L R L S C A A S G F T F V A Y N M
 GAGGTGAGC TTGGTGGAGTC TGGGGGGTC TGGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC
 CCTCAACCTCG ACAAACCTAG ACCCCCTCG AACCCTCTCG GACCCCGAGG GAGACGCTTC GAGACGCTAA GTCGAAAGGC CAAATATGTA
 T W V R Q A P G K G L E W V S S I N T F G N * T R Y A D S V K G R
 TGACTTGTT CGGCCAGGT CCAGGGAGG GTCTAGAGT GGTCTCATCG ATTATATCTT TTGTAATTA GCAAGGTTAC GCAAGCTTCG TGAAGGGCCG
 ACTGAACCA GGGGGTCTGA GGTCCTTC CAGATCTAC CCAGAGTCA TAATTATGAA ACCATTATCTT CTTCCTCATG CGTGTGAGG ACCTTCGGGC
 F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y C A K G S
 GTTCACCATC TCCCGGACA ATTCCAGAA CACGCTGTAT CTGCAATATGA ACAGGCTGTAG TGCCGAGGAC ACCGGGTAT ATTACTGTCG GAAAACGCTG
 CAAGCTGTAG AGGGCTGTAT TAAGGTCTT GTEGCTGTAT GACGTTACT TGTGCTGTAG TGGCCATATAATGACAGC CTTCGATAC
 R P F D Y W G Q G T L V T V S
 AGCTTATCT ACTACTGGG CCAGGAACCG CTGGTACCCG TCTCG
 TCCGAAAC TGATGACCC CGTCCTCG GACCACTGGC AGAGC

30

35 Partner dab monomer in TAR2h-5d3 (3U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F * G Y R M
 GAGGTGAGC TTGGTGGAGTC TGGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC CCTGGGGGTC
 CCTCAACCTCG ACAAACCTAG ACCCCCTCG AACCCTCTCG GACCCCGAGG GAGACGCTTC GAGACGCTAA GTCGAAAGGC CAAATATGTA
 G W V R Q A P G K G L E W V S W I T R T G G T Q Y A D S V K G R
 TGGGTGGGT CGGCCAGGT CCAGGGAGG GTCTAGAGT GGTCTCATCG ATTACGCTTA GCAAGGTTAC GCAAGCTTCG TGAAGGGCCG
 ACCCAACCA GGCGGTCC CAGATCTAC CCAGAGTAC CGTCCTCATG GAAAACGCTG CTTCGATAC GTCGATGACCC ACTTCGGGC

40

45

P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K P A
 GTCACCATC TCCCGGACA ATTCCAAAGA CACGGTGTAT CTGCAATGA ACGCCCTGCG TGCCGAGGAC ACCGGCTAT ATTACTGTC GAAACGGCG
 CAAGCTGCTAG AGGGCCCTGT TAACTGTTCTT GTGCGACATA GACGTTACT TGTGAGCGC ACGGCTCCTG TGGCCTATA TAATGACACG CTTGGCCG
 K L V G V G F D Y W G T L V T V S
 AGCTCTTGGGTGGCTT TCACTACTGG GGGCGGGAA CCTGTCAC CCTGTCAC
 TCTCCAAAC CCCAACCCAA ATCTGATGACC CGGTGTCCTT GGGACCAAGT GCGAACGG
 5

25	30	35	Partner dab monomer in Tar2h-5ds (3' linker)
			E V Q L L E S G G L V Q P G G S L R L S C A A S G P T F R R Y S M
			GAGGTGAGC TGTTGGAGTC TGGGGGGGGC TTGGTACAGGC CTCCTGGGATT CACCTTCGG CGCTATAGTA
			CTCCACCTCG ACAPACCTCG ACCCCCTCCG AACCATGTCG GACCCCGAG AGGAACACCTG GCAATATCAT
			S W R Q A P G L E W V S D I S R S G R Y T H Y A D S V K G R
			TGTCTGGGT CCGCAGGGT CGGGAAAGG GTCTAGAGT GGTCTAGAGT ATTTCCTGT CTGGTCTGGTA TACACATAC GCAGACTCTCC TAGACATAC CGTCTGAGGC ACTTCCGGC
			ACAGCACCCA GGGGTCTCGA GTTCCCTTCC CAGATCTCAC CCAGACTCTA TAAGAGCA GACCAAGCTA ATGTGTAACTGCTA CGTCTGAGGC ACTTCCGGC
			F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K R I
			GTTCAACATC TCCCGCAGCA ATTCCAAAGAA CACGGCTATG ACAGCTCTGC TGCCGGGAGC ACCGGGGAT ATTACTGTC GAAAGCTTATT
			CAAGTGGTAG AGGGCTGT TAAGGAGACATA GACGTTACTG TGCTGGAGGC ACGGCTCTCG TGGGGCCATA TAATGACACG CTTGCTATA
			D S S Q N G F D Y W G Q G T L V T V S
			GATTCCTCTC AGAAATGGGTT TGAATCTGG GGCCTGGGAA CCCCTGTCAC CGTCTCG
			CTAAGAAGAG TCTAACCAA ACTGTATGACC CCCTCTCCCTT GGGACCGTGC GCAAGGC

P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K Q K
 GTCACCAT TCCCGGACA ATCCGAGAA CACGCTGAT CTGCAAATGA ACAGCCTCG TGCCGAGAC ACCGGGTAT ATTACTGTG GAAACAGAG
 CAGGGTAG AGGGGCTGT TAGGTTCTT GTGCGACATA GACGTTACT TGTGCGACGC ACGGGCTCTG TGGGCCATA TAATGACAG CTTGGTCCTC
 E N F D Y W G Q T L V T V S
 5 GAGGTTTG ACTACTGGG CGAGGAAC CTCGTCACG TCTCG
 CTCCTAAC TGTGACCC GGTCCCTTG GACCGTGGC AGAGC

10 Partner dab monomer in 'tar2h-5d7' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F G D Y A M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTGGGGGTG TCCTGGCTTC CCTCCGGATT CACCTTGGG GATATGCTA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTAATACGAT
 W W V R Q A P G K L E W V S V I S S N G G S T F Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGT ATTAGTTCGA ATGGTGGGGAT GCAATTAC CGAGACTCCG TAGAGGGCCG
 ACACACCCCA GGCGCTCGAC GGTCCTCTTC CAGATCTAC TAATGCTAC TACCTACGCT TACCTACGCT ATGTAATATG CGTCTGAGGC ACTTCGGG
 P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K R V
 GTCACCATC TCCCCGACA ATCCGAGAA CACGCTAT CTGCAAATGA ACAGCCTCG TGCCGAGAC ACCGGGTAT ATTACTGTG GAAACGTTG
 CAGGGTAG AGGGGCTGT TAGGTTCTT GTGCGACATA GACGTTACT TGTGCGACGC ACGGGCTCTG TGGGCCATA TAATGACAG CTTGGACAA
 R K R T P E F D Y W G Q G T L V T V S
 15 CGTAAAGGAA CTCCCTGAGT TGACTACTGC GGCCGAGGAA CCCTGGTAC CGTCTCG
 CGATCTCTC GAGGACTAA ACTGAGAC CCGGTCCTT GGGACCGTGTG GCAGAGC

20 Partner dab monomer in 'tar2h-5d8' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F R R Y K M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 G W V R Q A P G K L E W V S A I G R N G T K T N Y A D S V K G R
 TGGGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACCAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG
 P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K I Y
 GTCACCATC TCCCCGACA ATCCGAGAA CACGCTAT CTGCAAATGA ACAGCCTCG TGCCGAGAC ACCGGGTAT ATTACTGTG GAAATTAT
 CAGGGTAG AGGGGCTGT TAGGTTCTT GTGCGACATA GACGTTACT TGTGCGACGC ACGGGCTCTG TGGGCCATA TAATGACAG CTTTAATA
 T G K P A A F D Y W G Q G T L V T V S
 25 Partner dab monomer in 'tar2h-5d9' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F K K Y * M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 S W V R Q A P G K L E W V S A I S G S T Y Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACAGAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG

30 Partner dab monomer in 'tar2h-5d9' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F K K Y * M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 S W V R Q A P G K L E W V S A I S G S T Y Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACAGAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG

35 Partner dab monomer in 'tar2h-5d9' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F K K Y * M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 S W V R Q A P G K L E W V S A I S G S T Y Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACAGAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG

40 Partner dab monomer in 'tar2h-5d9' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F K K Y * M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 S W V R Q A P G K L E W V S A I S G S T Y Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACAGAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG

45 Partner dab monomer in 'tar2h-5d9' (3'U linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F K K Y * M
 GAGGGCAGC TGTGGAGTC TGGGGAGGCT TGGTAAGGC ACGCTGCTTC CCTCCGGATT CACCTTGGG AGGTATATAAGA
 CTCCAGCTCG ACACCTCG ACCCCCTCG AACCATCTCG GACCCCTCG GACCCCGAG AGAACACGTC GGAGGCCATA GTGGAAACCC CTCATATCT
 S W V R Q A P G K L E W V S A I S G S T Y Y A D S V K G R
 TGTGGGGT CGCCGAGGT CCAGGGAGG GTCTGAGGT GGTCCTGAGC ATGGTGGGGAA GCAAAATAC CGAGACTCCG TAGAGGGCCG
 ACAGAACCA GGCGCTCGAA GGTCCTCTTC CAGATCTAC TAACCCCTCTT TACCATGCTT CTGTTATG CGTCTGAGGC ACTTCGGG

P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y C A K M L
 GTTCACATC TCCCCGACA ATTCCAAGA CACGCTGTAT CTGCAAATGA ACAGCTCTCG TGCCGAGGAC ACCGGGTAT ATTACTGTGC GAAATGCTG
 CTAAGTGTAG AGGGGCTGT TAAGTTCTT GTGCGACATA GACGTCTACT TGTGACGC ACGGCTCCCTG TGGCGCCATA TAATGACAGC CTTTACGAC
 R T K N K V F D Y W G Q T L V T V S
 AGCTTAAAG ATATGGTAA TGCACTACG GGCCTGGGAA CCCTGGTCTAC CGTCCTCG
 TCCGGAATTCT TAATCCACAA ACTGATGACCC CGGGTCCCTT GGGACCAAGT GGAGGCC

Ab monomer in Targh-5d11 (5' linker)
 E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F * S Y R M
 GAGSTGCGC TGTGTGGAGTC TGGGGGAGGC TTGGTACAGC CTGGGGGGTC CCTGGGGTCTC TCCTGGGGAG CCTCTGGTCAG
 CTCAGCTCG ACCAACCTCG ACCCTCG AACCAGTGC GACCCCTCG AGGAGGAG AGGAGAACCTC
 G W V R Q A P G G L E W V S S I S S R G R H T S Y A D S V K G R
 TGGGGGGT CCAGGGGAGG GTCTAGAGTG GGTCTCAAGT ATTCTGTTGA GGGGGTGGCA TACATCTTCG TGAGGGGGCG
 ACCAACCCA GGGGGTCCA GTTCCCCTTC CAGATCTCAC CGAGGTTCA TAMAGCTCA CGTCTGAGGC ACTTCCGGCG
 P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K R V
 GTTCACCATC TCCCCGACAA ATTCCAAAGGA CACGGCTGTAT CTGCAATGA ACAGCTGCG TGCCGGAGAC ACCGGGGTAT ATTACTGTGC GAAAGGGTT
 CAAGCTGTAG AGGGGGCTT TAAGGTCTT GTGGACATA GACGTTCAT TCTGGACGC AGGGCTCTG TGGGCCATA TAATGACAGC CTTTCCCAA
 P G R G G S P D Y W G G Q T L V T V S
 CGGCTGGG CGCGTCTT TTGACTACTGGG GCGCAGGGAA CCCTGGTCAC CGTCTCG
 CGCCCAAGCC CGGAAGAAA ACTGATGACC CGGTTCCCTT GGAGCACTG GCGAGAC

40

P T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K G E
 GTCGACCATC TCCCGGCGACA ATTCCAGAA CACGGCTTAT CTGCAATGA ACGCGCTGCG TGCGGAGGAC ACCGGCTAT ATTACGTCG
 CAAGTGCTAG AGGGCGCTGT TAAGGTCTT GTGCGACATA GACGCTTACT TGTGGACGC AGCGCTCTG TGCGGCATA TAATGACAG
 G G A S S A F D Y W G Q G T L V T V S
 GCGGGGGGCGA GTTCGCGCTT TGTACTTGC GCGCGCGGAA CGCGCGTCA CGCGCG
 CCCCCCCCCCTT CAAGACGCGA ACTGATGACC CGGTTCCTT GGACGACGTC CGACAGGC

10	Partner dab monomer in TAR2b-5d13 (5U linker)	E V Q L L E S G G G L V Q P G G S L R L S C A A S G F T F * R Y G M
15		GAGGTGCAAGC TGTGTGAGTC TCGCGGAGGC TTGTGTACAGC CTTGGGGGTC CCTGGTGTCTC TCTGTGTCAG CCTCGGGATT CACCTTTAG CGGTATGGGA CTCACTCGG ACCTACCTCG ACGACTGTG ACGACTCTCG ACGACTCGG ACGACTCGG AGAACAGCTG GAGACAGCTG GAGACAGCTA GTGGAAATC GGCATACCTT V W V R Q A P G K R L B W V S A I I S G S G T Y Y A D S V K G R TGGTTGGGT CGCGCGAGGT CCGCGGAAGG GTCTGTAGGT ATTAGTGTTGA GTGGGTGGTAG CACATACATC GCGAGCTCCG TGAGGCGGCG ACCARACCA GCGCGTCGA GTTCCTTC CAGATCTAC CCAAGATCGA TAATCACCCT CACCCACCAT GGTGTGATG CGTCGAGGC ACTTCCGGC F T I S R D N S K N T L Y L Q M N S L R A E D T A V Y Y C A K R H GTCACCATC TCCCGGACA ATTCCAGAA CACCGCTGTAT CTCGAATAGA ACAGCTCTGG AGGGCTGGAC ACCCGGTAT ATTACTCTGC GAAACGGCAT CAAGCTGGT AGGGCTGGTG TGTGGTACT GTGGACATA GACGTACT TGTGGTACT GCGGTGAGC AGGGCTCTG TGGCGCCATA TAATGACAGC CTTCGCGCTA S S A R Q F D Y W G Q G T L V T V S AGTTCTGAGG CTAGGCAAGT TGACTACTGG GGCCTGGGAA CCCTGGTCAC CGTCCTCG TCAAGACTCC GATCCGTCAA ACTGATGACC CGGGTCCCTT GGGACCACTG GCAGAGC
20		

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(10) International Publication Number
WO 2004/003019 A3

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(74) Agents: MASCHIO, Antonio et.al.; D Young & Co, 21 New Fetter Lane, London EC4A 1DA (GB).

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 2004/003019 A3

(54) Title: IMMUNOGLOBIN SINGLE VARIANT ANTIGEN-BINDING DOMAINS AND DUAL-SPECIFIC CONSTRUCTS

(57) Abstract: The invention provides a dual-specific ligand comprising a first immunoglobulin variable domain having a first binding specificity and a complementary or non-complementary immunoglobulin variable domain having a second binding specificity.

INTERNATIONAL SEARCH REPORT

Int'l Application No
PCT 3 03/02804

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C07K16/24 C07K16/18 C07K16/28 A61K39/395 C07K16/46
C12N15/63 C12N15/62 C12N15/13

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, WPI Data, PAJ, MEDLINE, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	ELS CONRATH K ET AL: "Camel single-domain antibodies as modular building units in bispecific and bivalent antibody constructs" JOURNAL OF BIOLOGICAL CHEMISTRY, AMERICAN SOCIETY OF BIOLOGICAL CHEMISTS, BALTIMORE, MD, US, vol. 276, no. 10, 9 March 2001 (2001-03-09), pages 7346-7350, XP002248402 ISSN: 0021-9258 cited in the application page 7350 page 7349	1-21, 47-56, 60, 111, 112
X		113-117
Y		24-26, 32-35, 38, 40, 47-56, 64-67, 70-72, -/-

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the International filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the International filing date but later than the priority date claimed

- *T* later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *&* document member of the same patent family

Date of the actual completion of the International search

8 June 2004

Date of mailing of the International search report

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INTERNATIONAL SEARCH REPORT

Int'l Application No
PCT/GB 03/02804

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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X	WO 00/29004 A (PEPTOR LTD ; PLAKSIN DANIEL (IL)) 25 May 2000 (2000-05-25) example 2	22, 23, 25, 29-31, 38, 40, 47-56, 61-63, 70-72, 111, 112
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Int'l Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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INTERNATIONAL SEARCH REPORT

Inte ~~l~~ Application No
PC., JB 03/02804

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	MUYLDERMANS S ET AL: "UNIQUE SINGLE-DOMAIN ANTIGEN BINDING FRAGMENTS DERIVES FROM NATURALLY OCCURRING CAMEL HEAVY-CHAIN ANTIBODIES" JOURNAL OF MOLECULAR RECOGNITION, HEYDEN & SON LTD., LONDON, GB, vol. 12, no. 2, March 1999 (1999-03), pages 131-140, XP009012180 ISSN: 0952-3499 abstract ---	1-21, 47-56, 60,111, 112
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INTERNATIONAL SEARCH REPORT

In National Application No
PCT/GB 03/02804

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>TANHA J ET AL: "Optimal design features of camelized human single-domain antibody libraries." THE JOURNAL OF BIOLOGICAL CHEMISTRY. UNITED STATES 6 JUL 2001, vol. 276, no. 27, 6 July 2001 (2001-07-06), pages 24774-24780, XP002283749 ISSN: 0021-9258 the whole document</p> <p>---</p>	1-117
P,A	<p>WO 02/072141 A (HERMAN WILLIAM) 19 September 2002 (2002-09-19) the whole document</p> <p>-----</p>	1-117

INTERNATIONAL SEARCH REPORTInternational application No.
PCT/GB 03/02804**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this International application, as follows:

see additional sheet

1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-7, 8-10, 11-21, 47-56 (all 10 in part),
60 (in part), 111-112 (both in part)

Ligand comprising a first immunoglobulin variable domain having a first antigen or epitope specificity and a second immunoglobulin variable domain having a second antigen or epitope specificity wherein one or both of the antigens or epitopes acts to increase the half-life of the ligand in vivo, and the variable domains are not complementary to one another.

2. Claims: 22, 23, 25, 29-31, 38 (in part), 40,
47-56 (all 10 in part), 61-63, 70-72 (all 3 in part),
111-112 (both in part)

Single domain antibody monomer ligand specific for TNF-.

3. Claims: 24, 26, 25, 32-35, 38 (in part), 40,
47-56 (all 10 in part), 64-67,
70-72 (all 3 in part), 111-112 (both in part)

Single domain antibody monomer ligand specific for TNF receptor 1.

4. Claims: 27, 28, 36, 37, 38 (in part) 40,
47-56 (all 10 in part), 64-67,
70-72 (all 3 in part), 111-112 (both in part)

Single domain antibody monomer ligand specific for serum albumin.

5. Claims: 39, 47-56 (all 10 in part),
111-112 (both in part).

Single domain antibody monomer ligand comprising a terminal Cys residue.

6. Claims: 41-46, 47-56 (all 10 in part), 60 (in part).

A dual specific ligand comprising at least a single domain antibody monomer according to inventions 2-5.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

7. Claims: 73-81, 82-104, 105-110

A closed conformation multispecific ligand comprising a first epitope binding domain having a first epitope binding specificity and a non-complementary second epitope binding domain having a second epitope binding specificity, wherein the first and second binding specificities are capable of competing for epitope binding such that the closed conformation multi-specific ligand cannot bind both epitopes simultaneously. Methods for producing the closed conformation multispecific ligand. Methods using the closed conformation multispecific ligands.

8. Claims: 113-117

Method for preparing a chelating multimeric ligand, specific for adjacent epitopes.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 03/02804

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INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 03/02804

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